

NTM Lecture Series for Providers

April 27-28, 2023
NATIONAL JEWISH HEALTH



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Professor of Medicine
National Jewish Health
University of Colorado

Disclosures

Conflicts

- none

Supported by:

CFF

- Colorado NTM Core Clinical Research Service NRC
- NTM Outcome Measure Advancement Core NRC
- NICK20A0 Urine Lipoarabinomannan as a Marker for Low-risk of NTM Airway Infection
- NICK17K0 NTM-OB-17 Colorado Adult
- NICK21K0 Prospective Evaluation of Markers of NTM and Host Response in Saliva

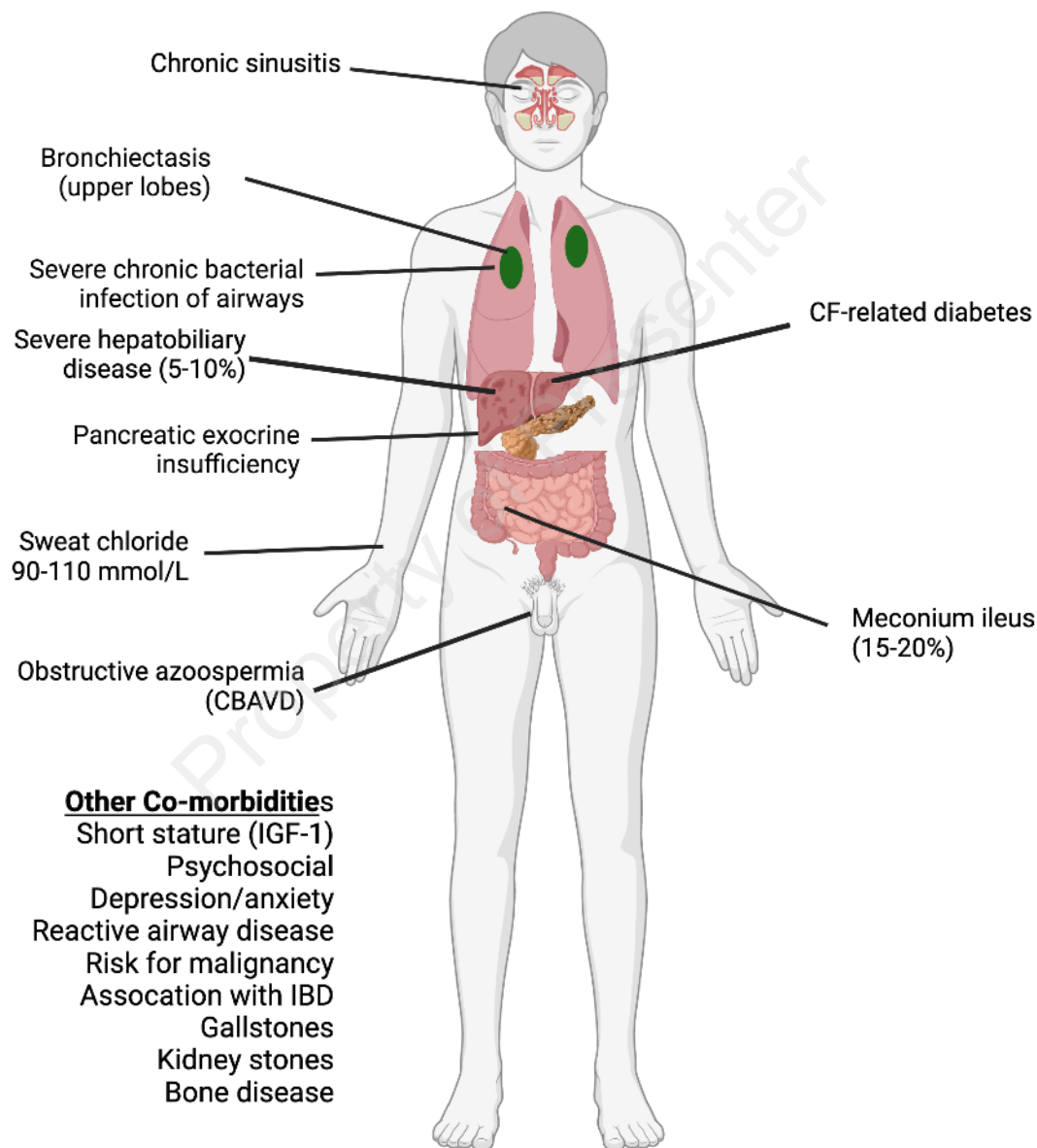
NIH

- R01HL146228 Longitudinal Assessment of Culture-Independent Molecular Airway Markers of Nontuberculous Mycobacteria

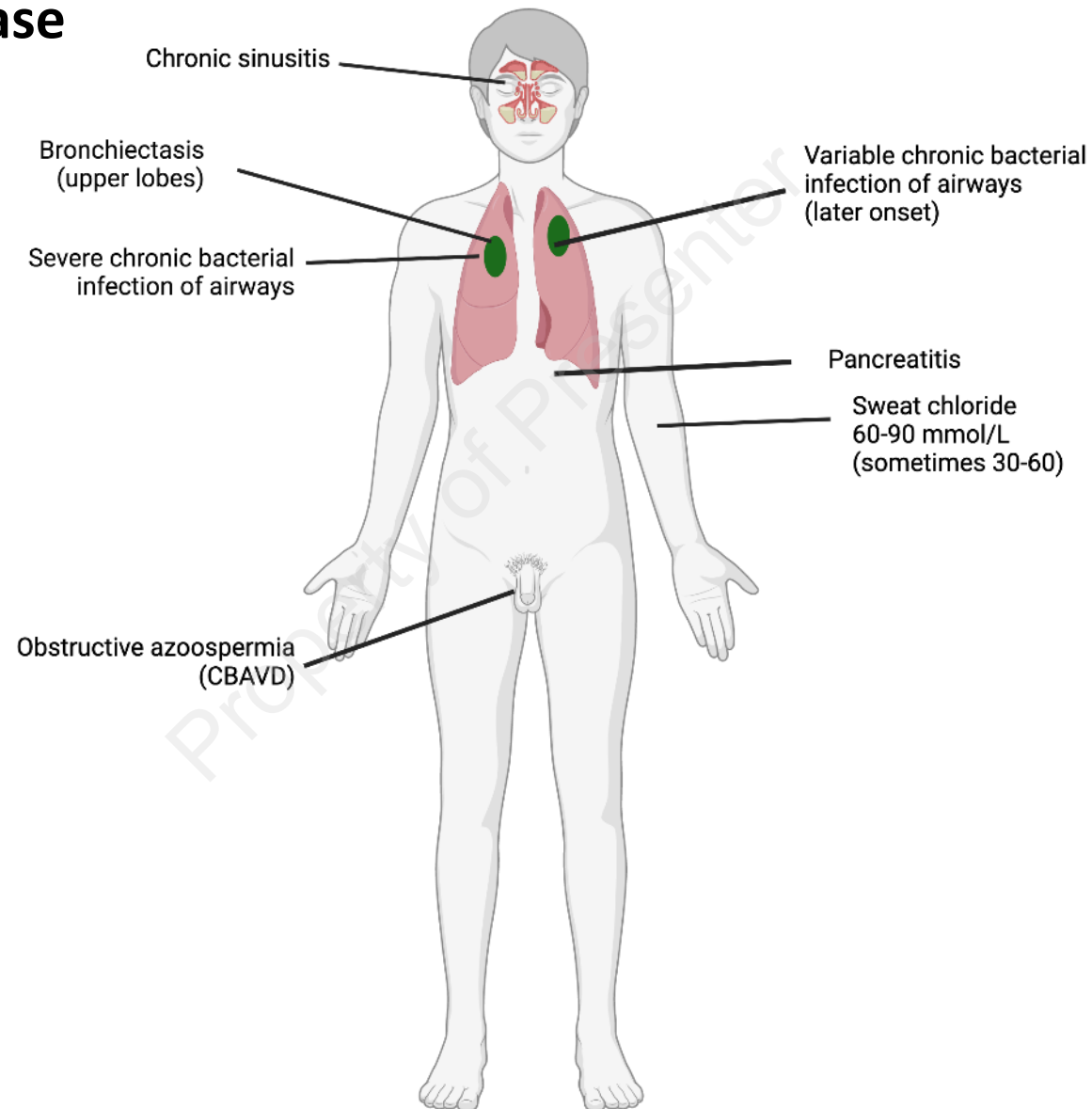
FDA

- R01FD-R-6848 A Phase 1b, Multi-center Study of IV Gallium Nitrate in Patients with Cystic Fibrosis who are colonized with NTM

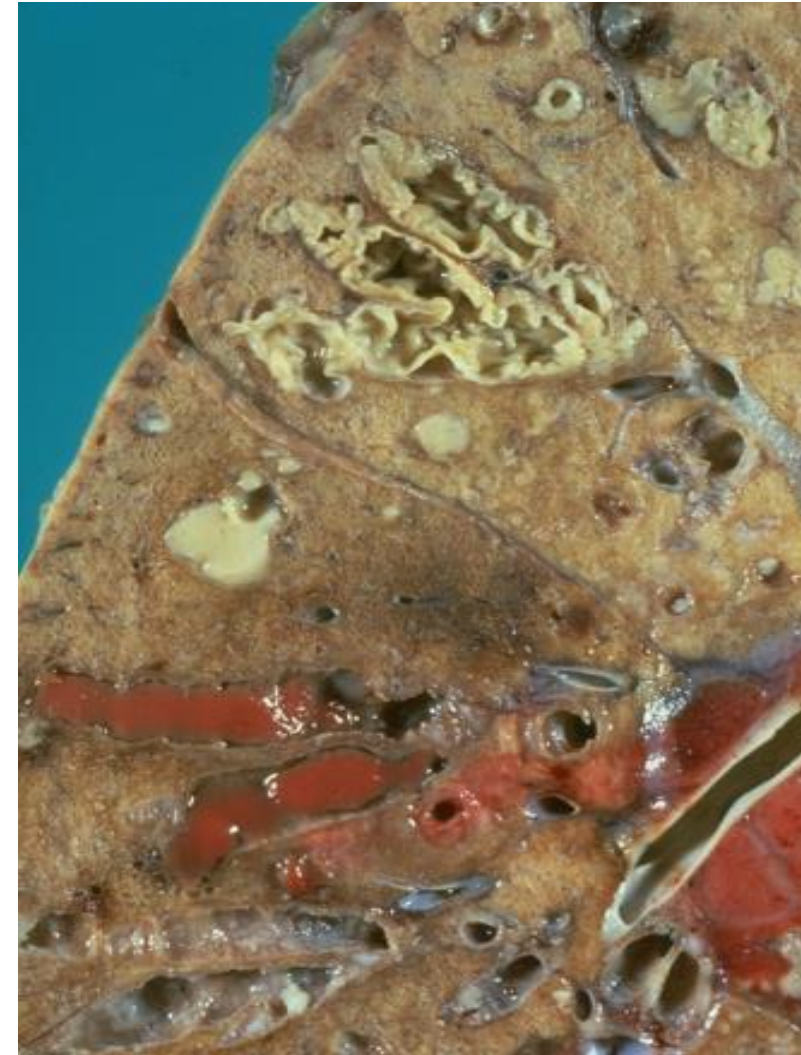
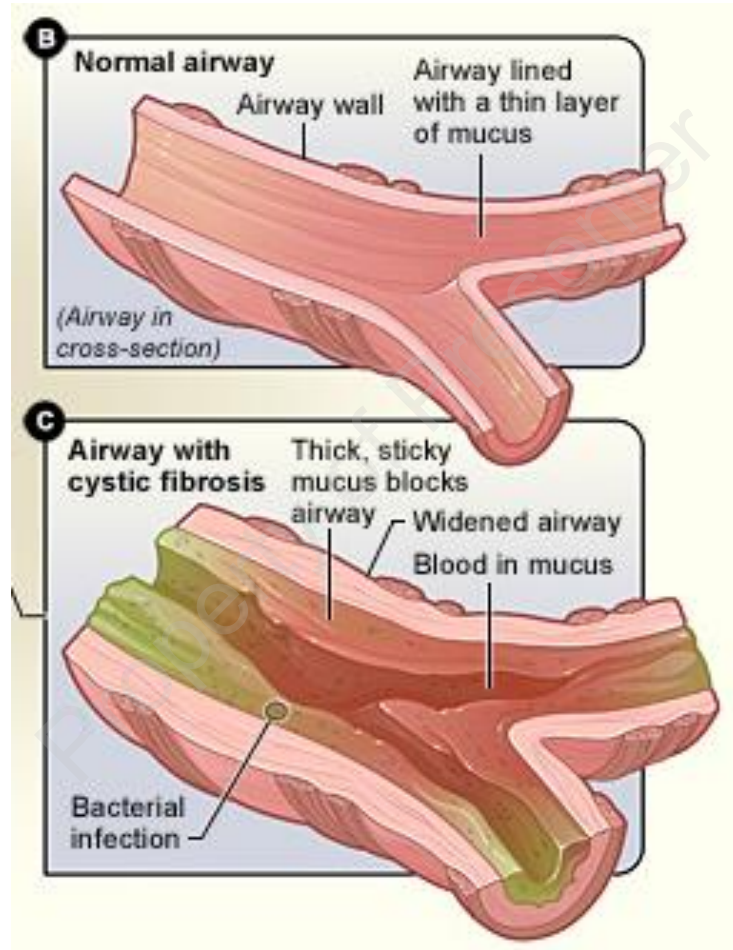
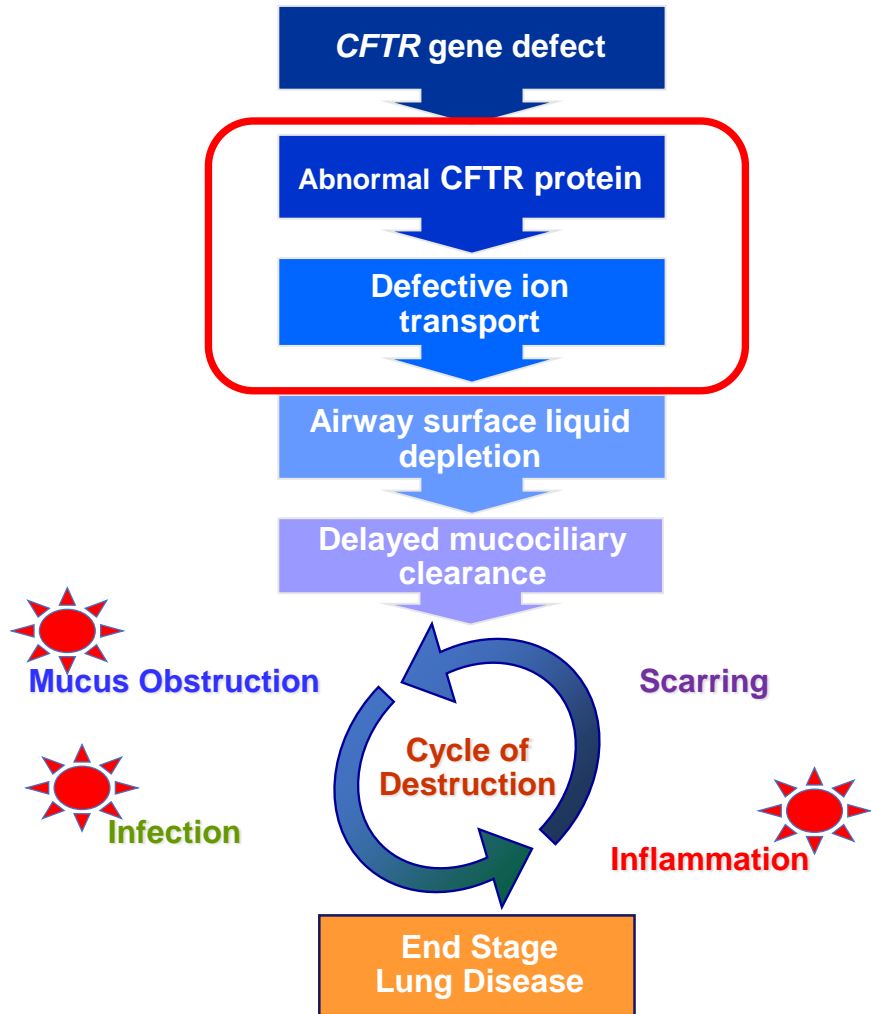
Classic (severe) Disease -childhood diagnosis ($\approx 90\%$)



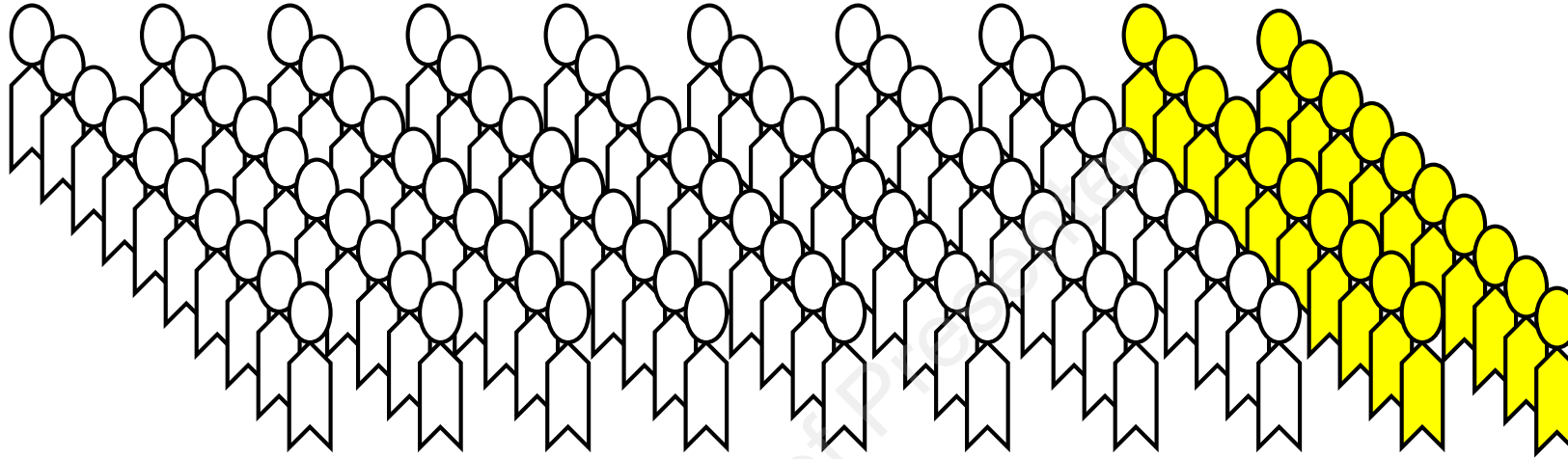
Nonclassic (delayed) Disease -adult diagnosis ($\approx 10\%$)



Pathogenesis of CF lung disease

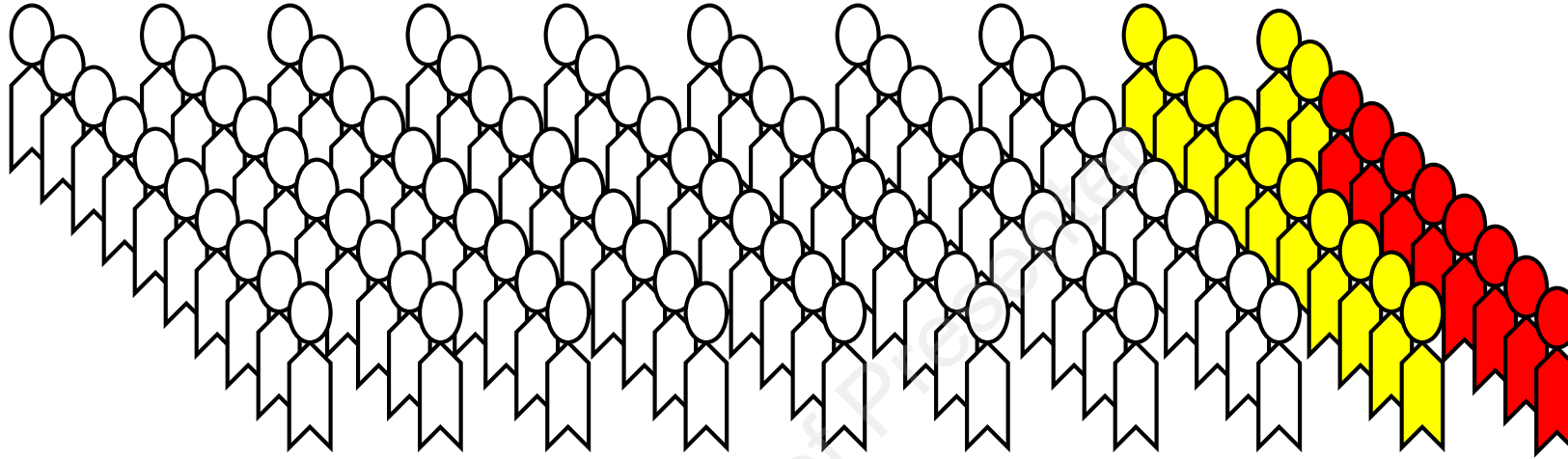


Epidemiology of NTM in the United States CF Population



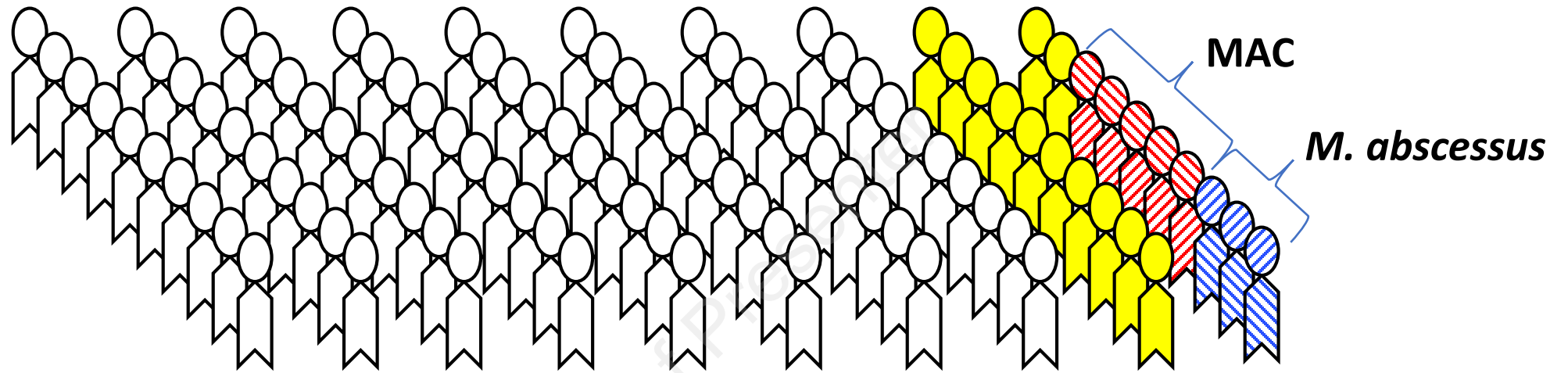
Approximately 20% of CF population will grow NTM over a 5-year period

Epidemiology of NTM in the United States CF Population



25-40% will meet criteria for NTM pulmonary disease

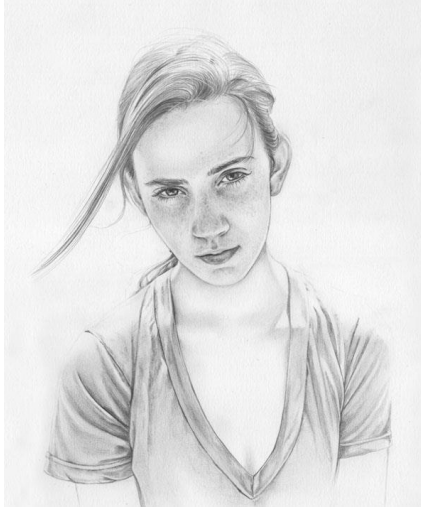
Epidemiology of NTM in the United States CF Population



MAC is the predominant NTM in the United States CF population

NTM pulmonary infection and disease: Cystic Fibrosis vs. Non-CF

Cystic Fibrosis



- CF Care Center
- Lifelong sputum culture surveillance = Earlier detection without clinical features
- MAC or *M. abscessus*
- Always co-infection with other bacteria
- Uniform pathogenesis
- Greater variability in drug PK
- Evidence of diminished response to treatment
- Uncertainty regarding the effect of highly effective modulator therapy

Non-CF Bronchiectasis

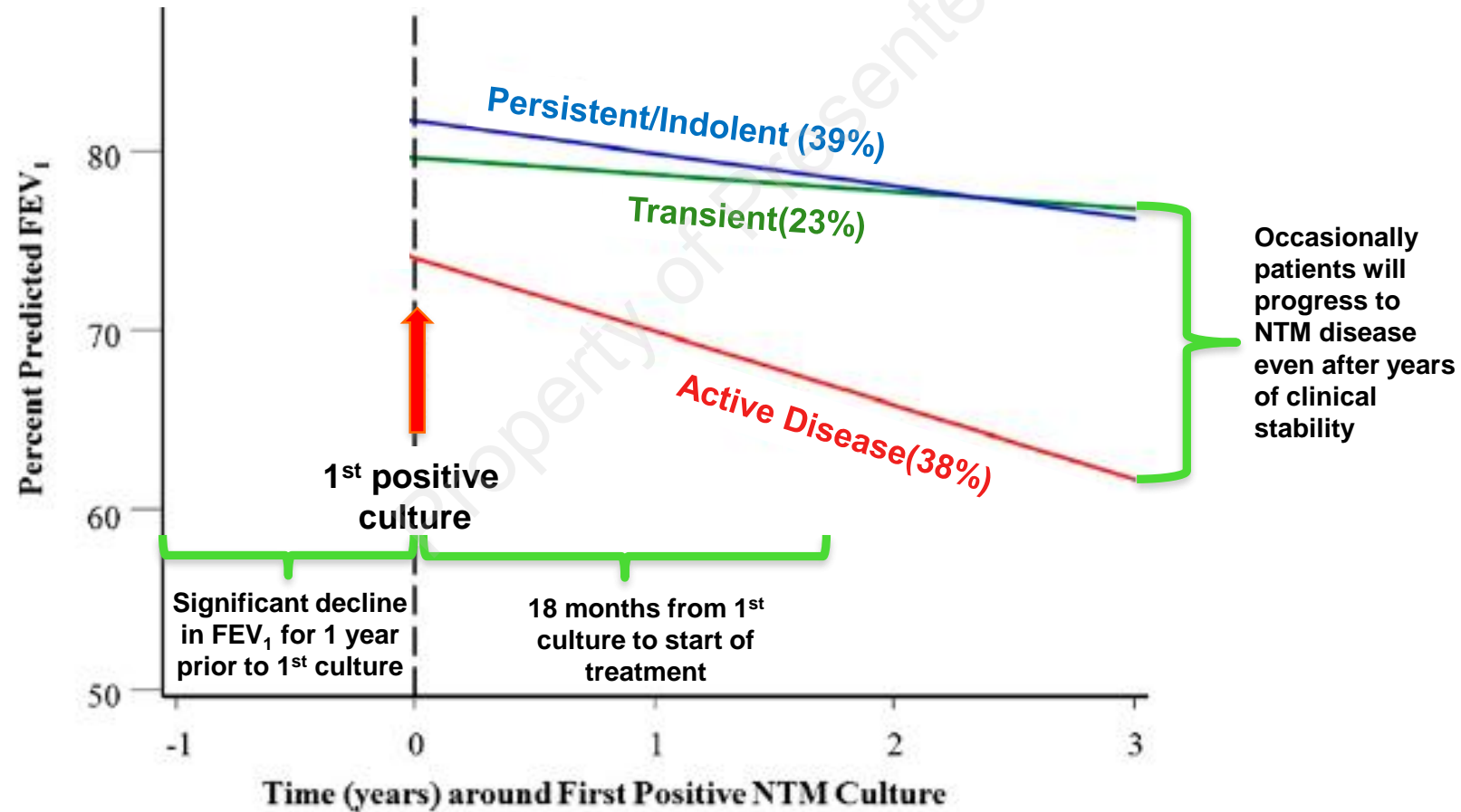


- Community-based care
- Infrequent airway cultures= Clinical symptoms prompt cultures
- Greater spectrum of NTM infections
- NTM may be primary infection
- Mix of underlying diseases and co-morbidities

Clinical Significance of a First Positive NTM Culture in CF

Change in FEV₁ pre- and post first positive NTM culture

Average time of follow-up 4.4 yrs. (n=96)



Diagnosis of NTM Pulmonary Disease: CF-specific considerations

Other CF pathogens and co-morbidities should be considered as potential contributors to a patient's symptoms and radiological features when determining the clinical significance of NTM positive cultures

CFF/ECFS Guidelines: NTM treatment **should be considered** for individuals with CF who have ATS/IDSA defined NTM pulmonary disease.

Suboptimal CF care

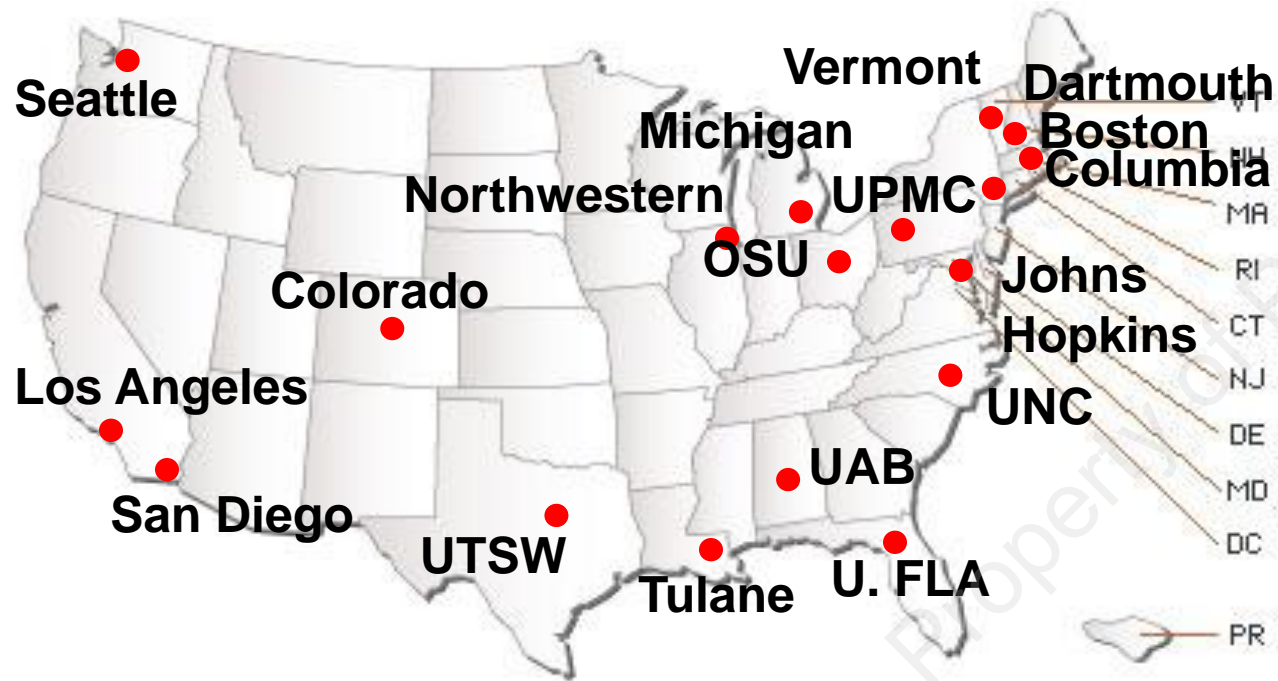
Pulmonary Exacerbations

- Usual CF pathogens
- New bacterial infection

Co-morbidities:

- Allergic bronchopulmonary aspergillosis (ABPA), asthma
- CF-related diabetes
- Sinus disease
- Gastroesophageal reflux
- Chronic aspiration
- Nutritional deficiencies

Cystic Fibrosis NTM Consortium 2023



2013

- National Jewish Health, Children's Hospital Colorado

2018

- University of Washington
- University of Michigan
- University of Texas Southwestern
- University of Alabama Birmingham (Adult Program)
- Johns Hopkins University
- University of North Carolina
- Columbia University

2022

- University of Florida (Gainesville)
- Rady Children's Hospital, University of California San Diego
- Children's Hospital of Pittsburgh of UPMC & University of Pittsburgh Medical Center
- University of Vermont Children's Hospital (Adult)
- Nationwide Children's Hospital (Columbus, OH)
- Dartmouth Hitchcock Medical Center
- Northwestern University (Adult)
- Boston Children's
- Los Angeles Children's Hospital, USC Adult Program
- Tulane University

Prospective Evaluation of NTM Disease in Cystic Fibrosis (PREDICT Study)



- Prospective, single-center, observational trial at the Pediatric and Adult Colorado CF Care Center
 - Single diagnostic algorithm based on CF Foundation and European CF Society Guidelines (*Thorax*, 2016)
- **Inclusion:**
 - Diagnosis of CF
 - Positive NTM culture (last 2 years) with *M. avium* complex (MAC) or *M. abscessus* complex (MABSC)
- **Exclusion:**
 - Recent or current treatment of NTM



PATIENCE: Objectives

Prospective, open-label, treatment trial

Primary Study Goals:

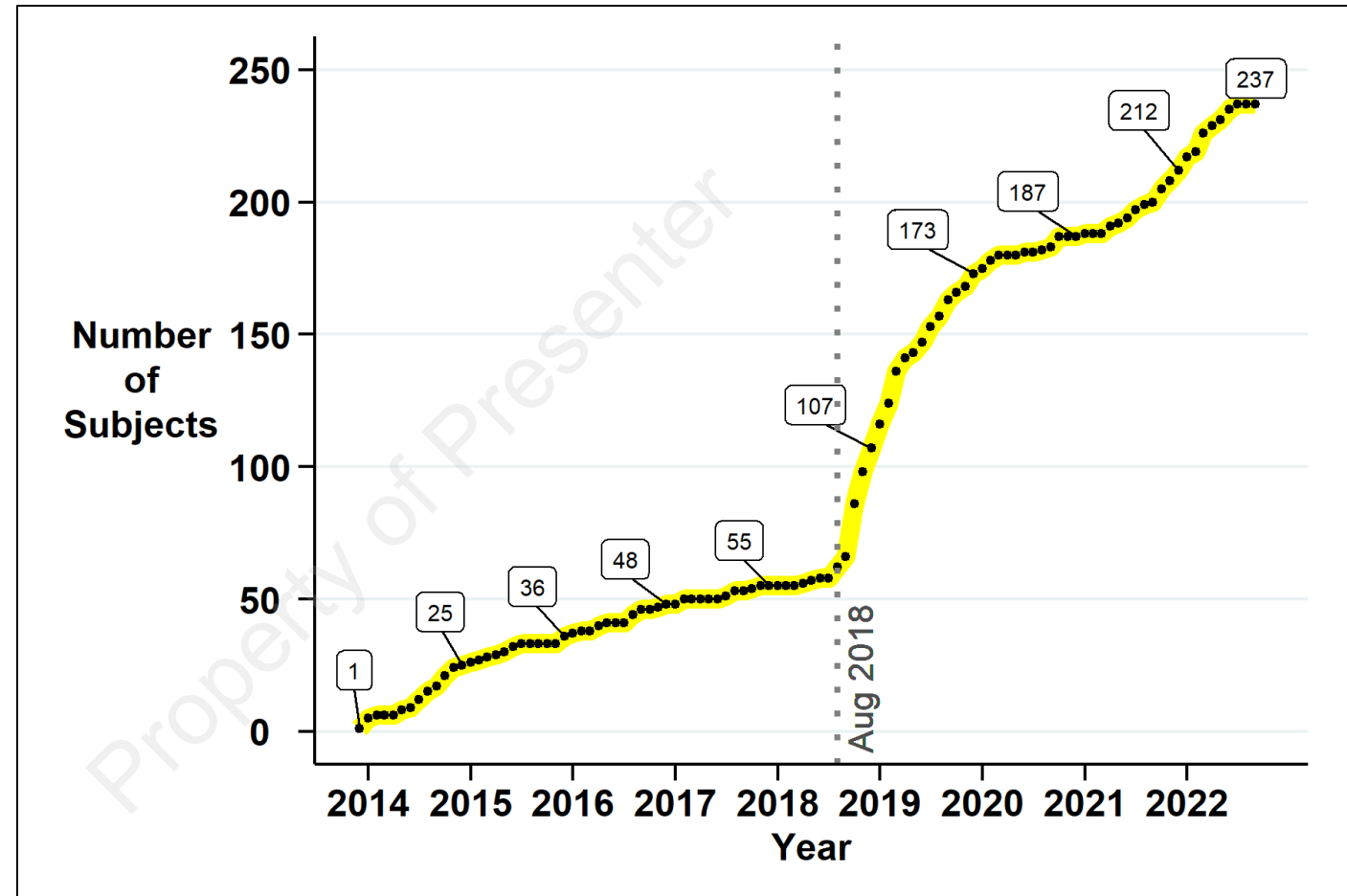
- To implement a standardized approach to the initial treatment of NTM pulmonary disease (NTM-PD) in CF patients

Secondary Study Goals:

- Define an expected rate of clinical response and tolerance of treatment of NTM-PD
- Establish foundation for future NTM treatment trials

Current number of
PREDICT participants
“ongoing study
procedures”:
n=151

PREDICT: Ever Enrolled

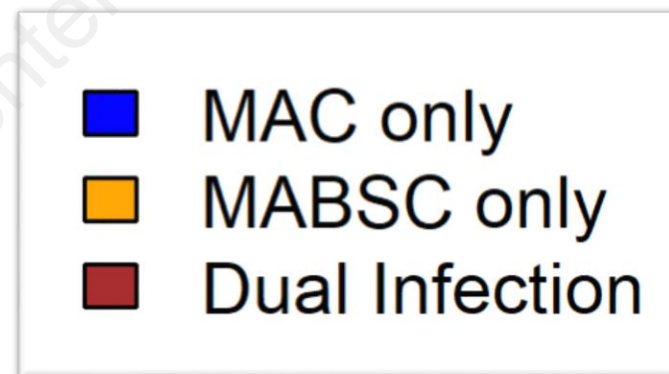
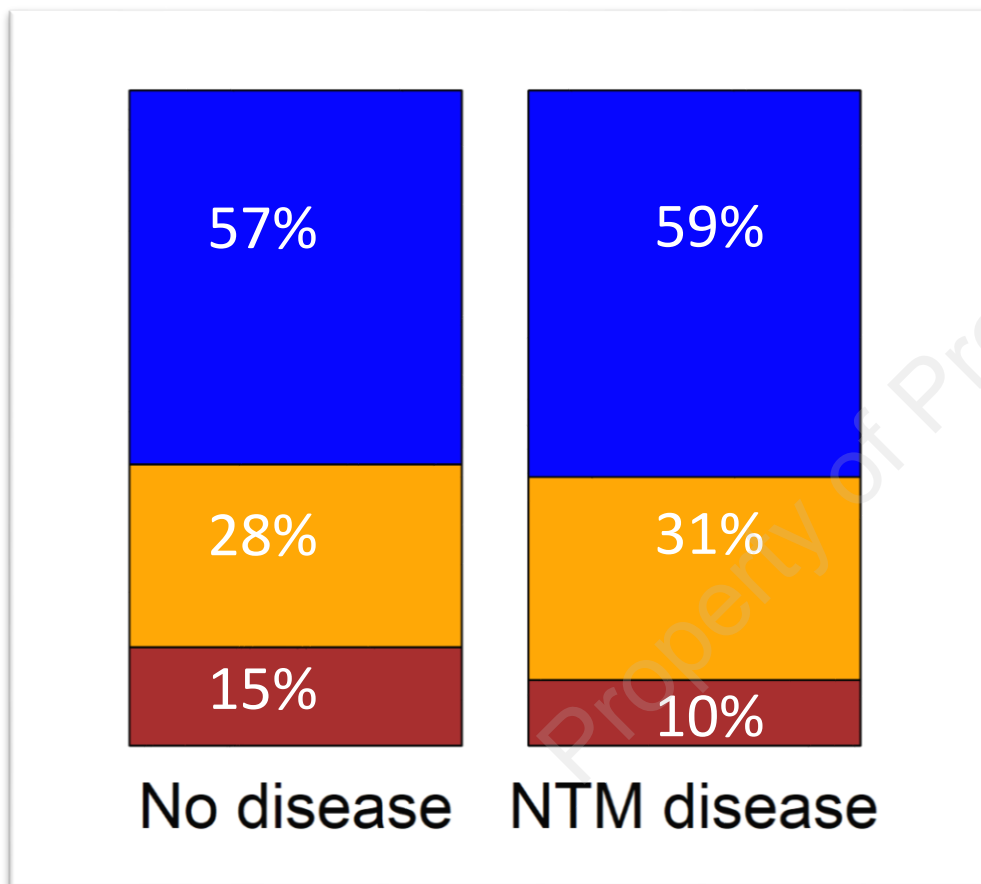


NTM PREDICT: Clinical Comparisons by NTM Disease Status

	Total (n=237)	No NTM disease (n=181; 76%)	NTM disease (n=56; 24%)	P-value
Male sex (%)	40%	43%	30%	0.58
Enrollment FEV% predicted, mean (SD)	79 (24)	80 (24)	75 (23)	0.18
Enrollment age, mean (SD)	30 (14)	31 (14)	25 (13)	0.01
Enrollment age distribution				
6 to 12 years	8%	7%	13%	0.05
>12 to 18 years	11%	9%	18%	
>18 to 30 years	43%	43%	43%	
> 30 years	38%	42%	27%	
First Lifetime NTM culture age, mean (SD)	27 (13)	27 (13)	22 (13)	0.02

* No differences in CF genotype, race, ethnicity

NTM Species Distribution

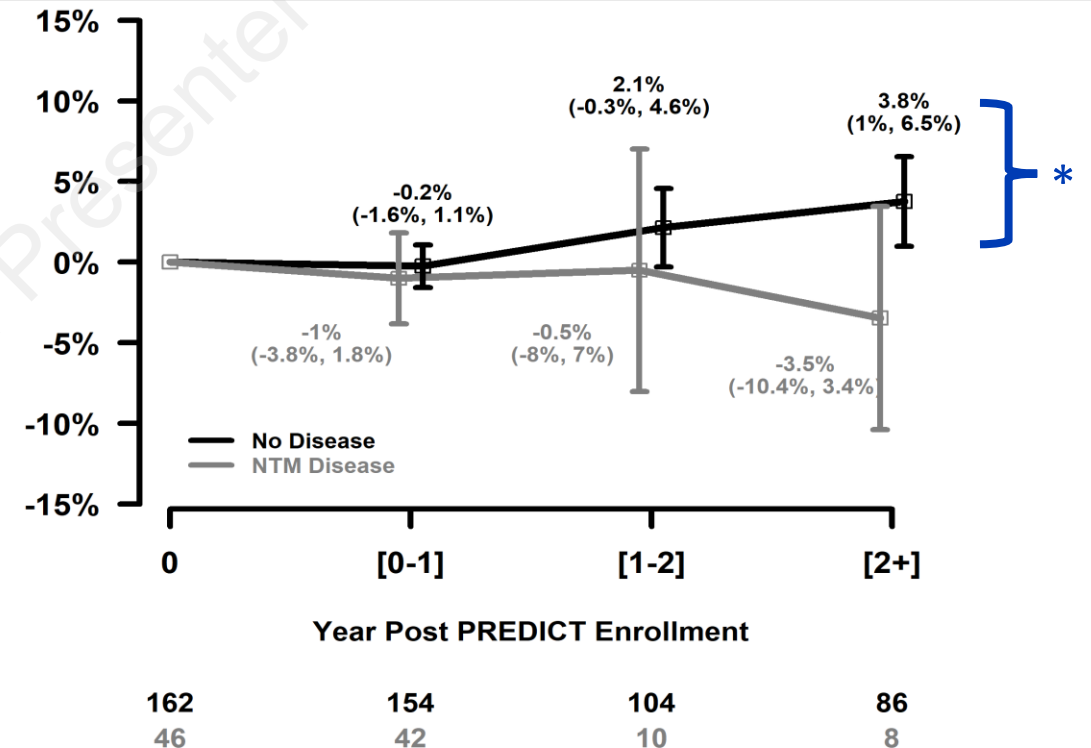


PREDICT: Follow Up & Change in FEV1% Predicted

Mean Follow-up Time:

No NTM Disease:
30 months
4 clinic visits per year

NTM Disease Diagnosis:
12 months
6 clinic visits per year



* For those with NTM-PD diagnosis 2 years post enrollment, FEV1pp was 7 % lower (95% CI 0% to 14%, p=0.05) compared to no NTM-PD

Prospective Algorithm for Treatment of NTM in Cystic Fibrosis (PATIENCE)



Prospective, open-label, treatment trial at the Colorado CF Care Center

- Single treatment algorithm based on CF Foundation and European CF Society Guidelines (*Thorax*, 2016)

Inclusion: confirmed diagnosis of CF

- Age 7 years or greater
- Diagnosis of NTM disease (via PREDICT or Provider)
- Intention to treat the NTM disease, based on the judgment of the CF clinic physician that the patient may benefit from treatment

Exclusion:

- Pregnant
- History of transplantation
- Currently undergoing treatment for NTM infection
- Prior treatment failure for current NTM species, as defined by positive sputum cultures within 12 months of discontinuation of antibiotic treatment



I. Stenzel

Photo by Derek Powazek

<http://www.thebreathingroom.org/>

PATIENCE: Objectives

Prospective, open-label, treatment trial

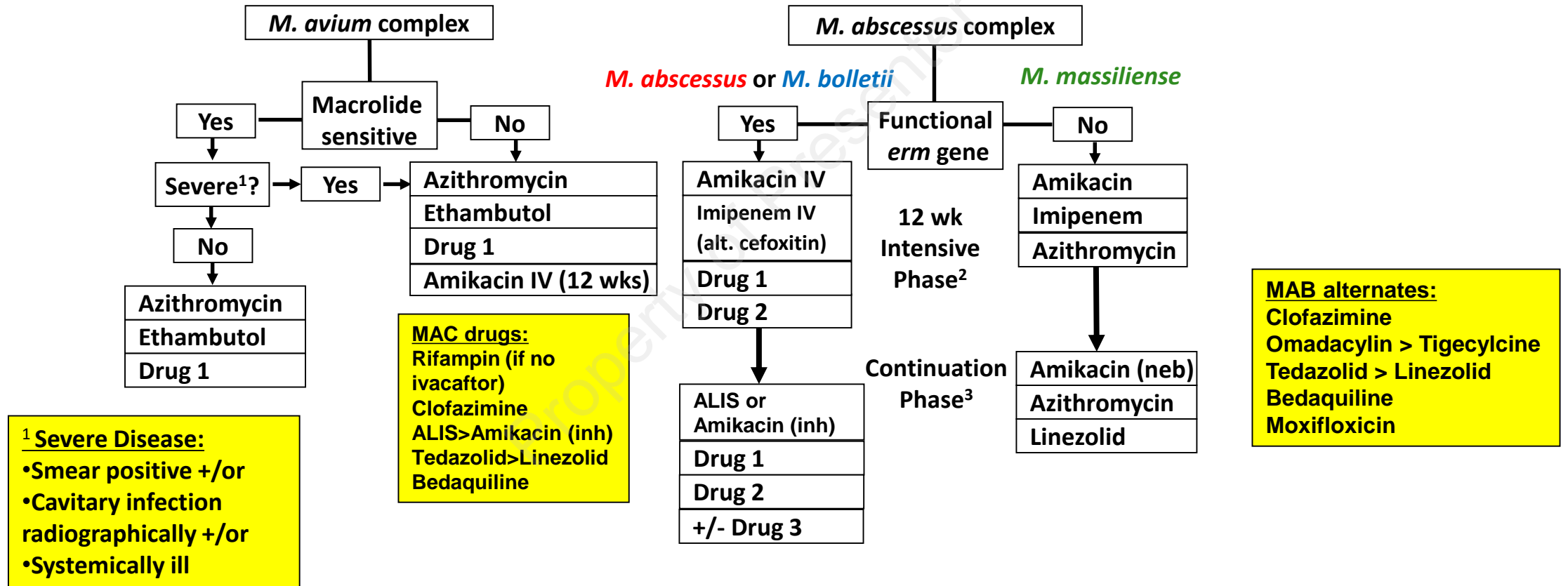
Primary Study Goals:

- To implement a standardized approach to the initial treatment of NTM pulmonary disease (NTM-PD) in CF patients

Secondary Study Goals:

- Define an expected rate of clinical response and tolerance of treatment of NTM-PD
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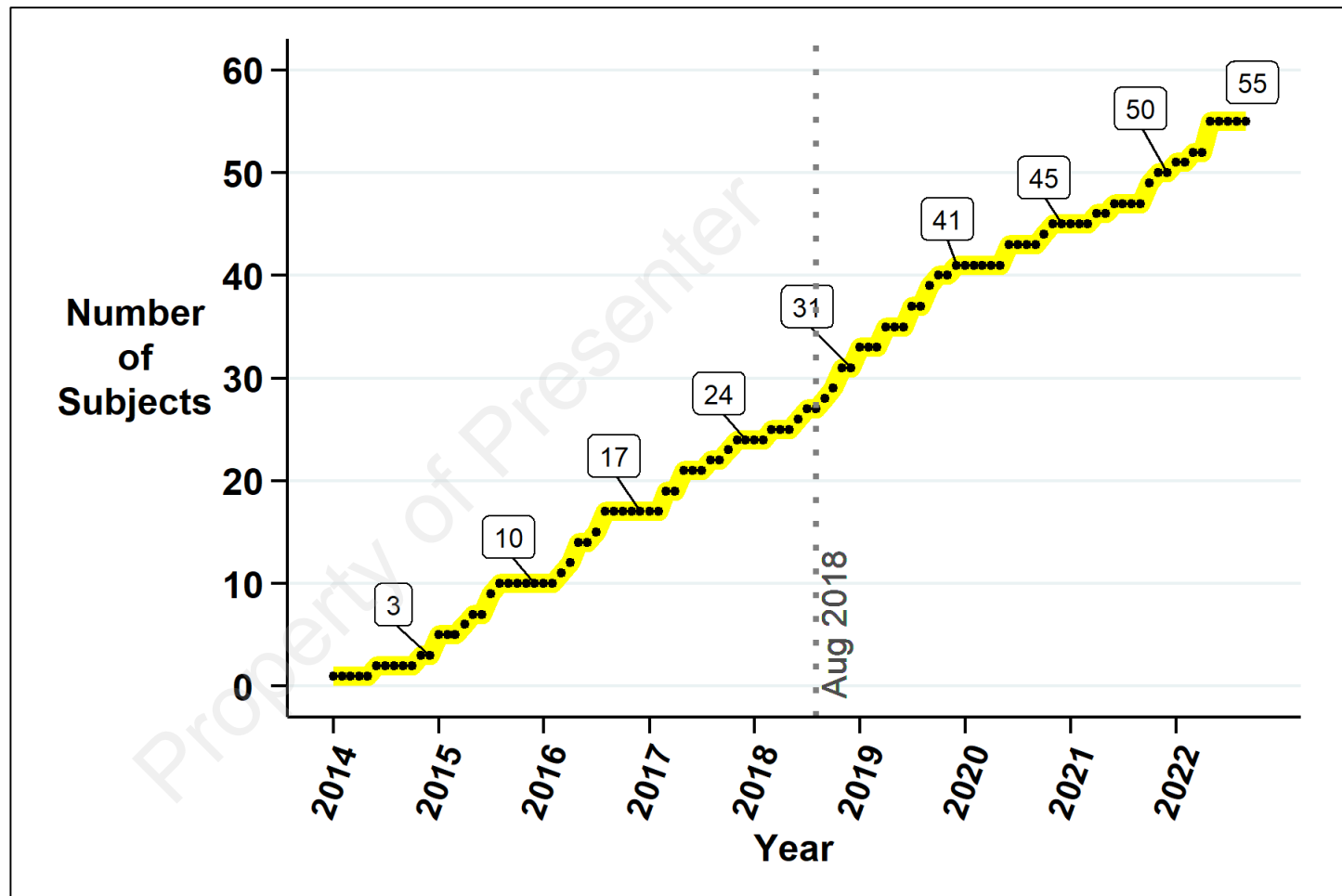
PATIENCE Treatment Protocols



All NTM isolates are evaluated by WGS by the Colorado NRC Molecular Core

PATIENCE: Ever Enrolled

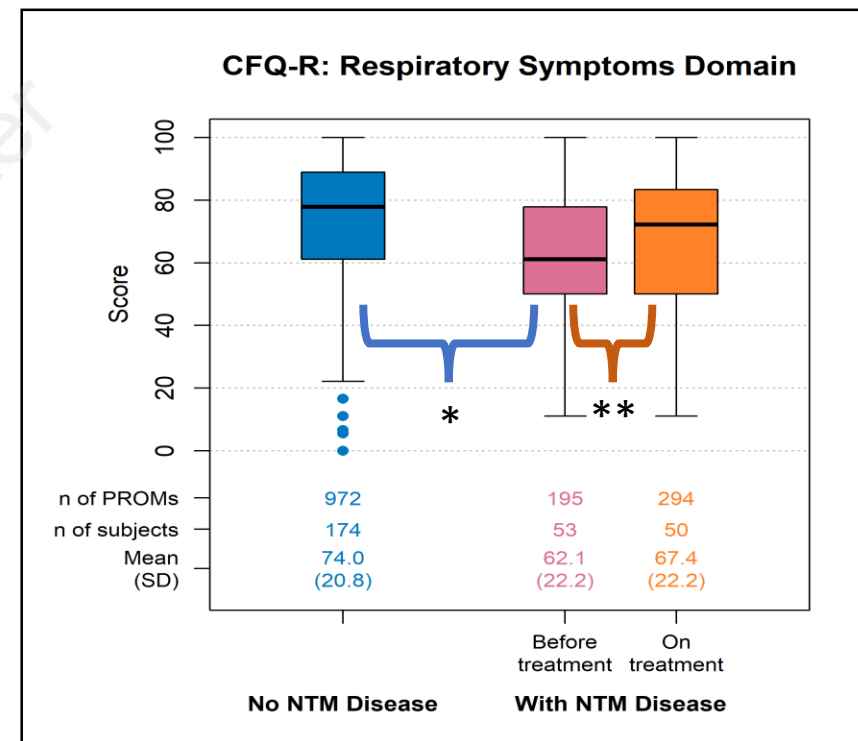
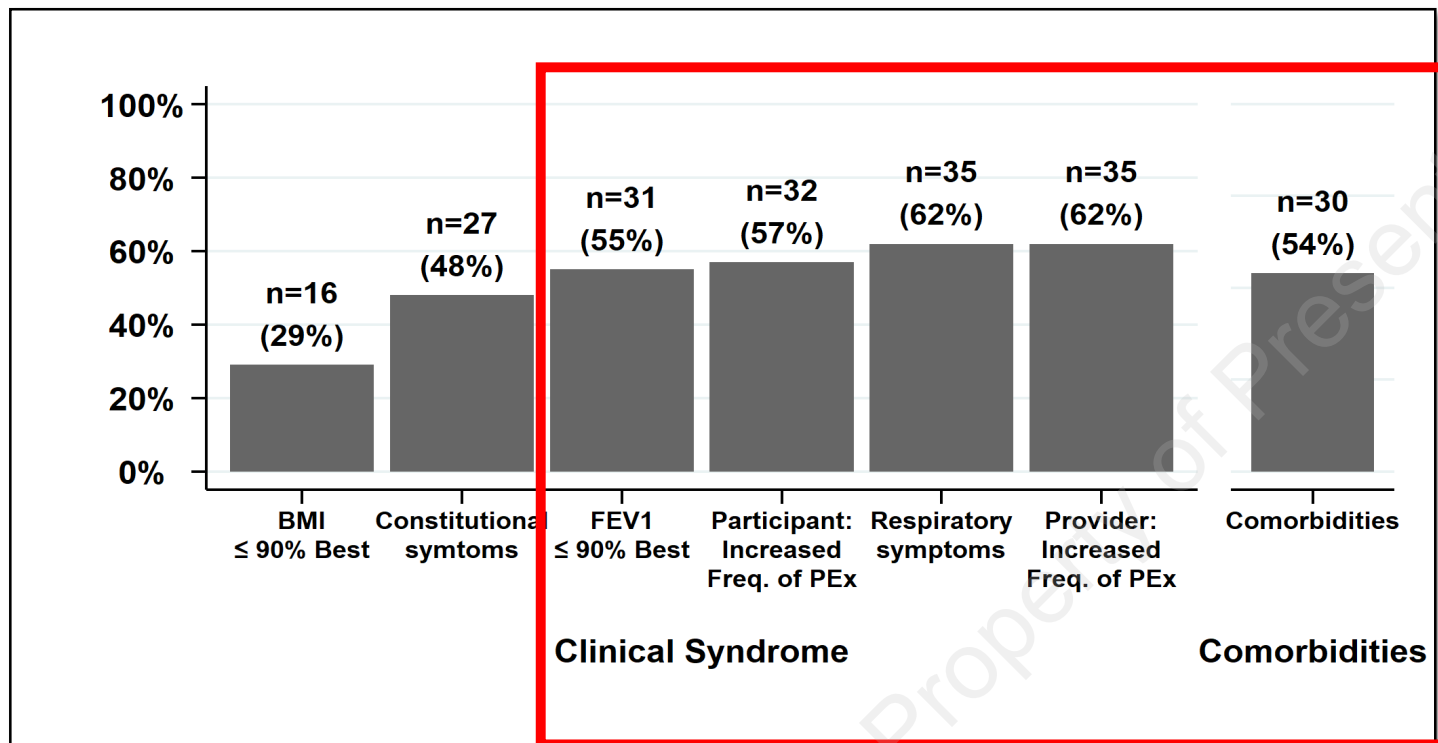
Current number of
PATIENCE participants
"ongoing study
procedures":
n=15



PATIENCE Demographics: MAC & MABSC

	Total (n=55)	MAC (n=33; 60%)	MABSC (n=22; 40%)
Male sex (%)	31%	24%	41%
Enrollment FEV% predicted, mean (SD)	72 (24)	76 (24)	64 (21)
Enrollment age, mean (SD)	25 (12)	26 (14)	25 (10)
Enrollment age distribution			
6 to 12 years	9%	12%	5%
>12 to 18 years	18%	18%	18%
>18 to 30 years	49%	42%	59%
> 30 years	24%	27%	18%
CF Genotype			
F508del homozygous	46%	46%	46%
F508del heterozygous	38%	39%	36%
other	17%	12%	18%

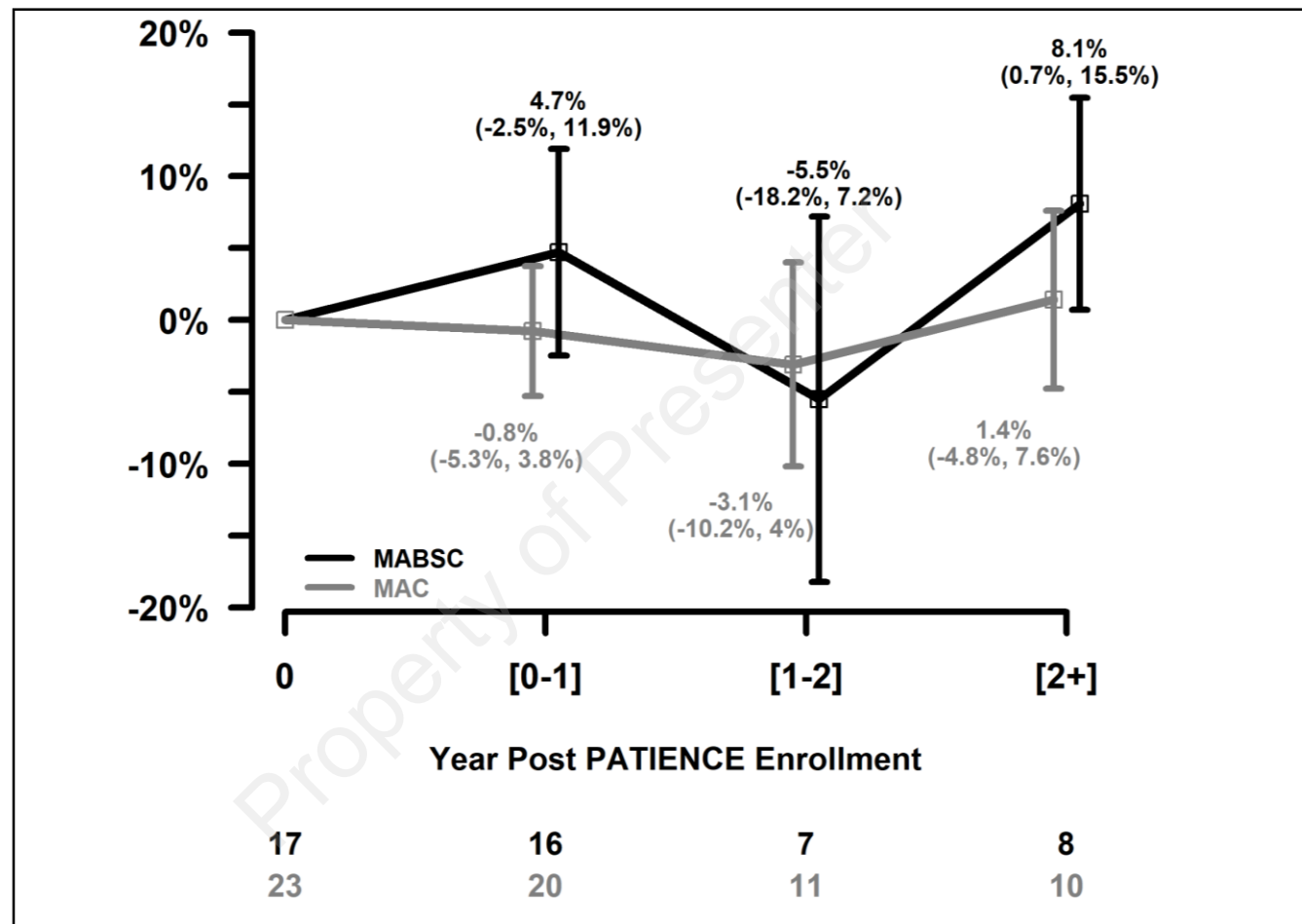
Clinical Syndrome & CFQ-R at Time of NTM Disease Diagnosis



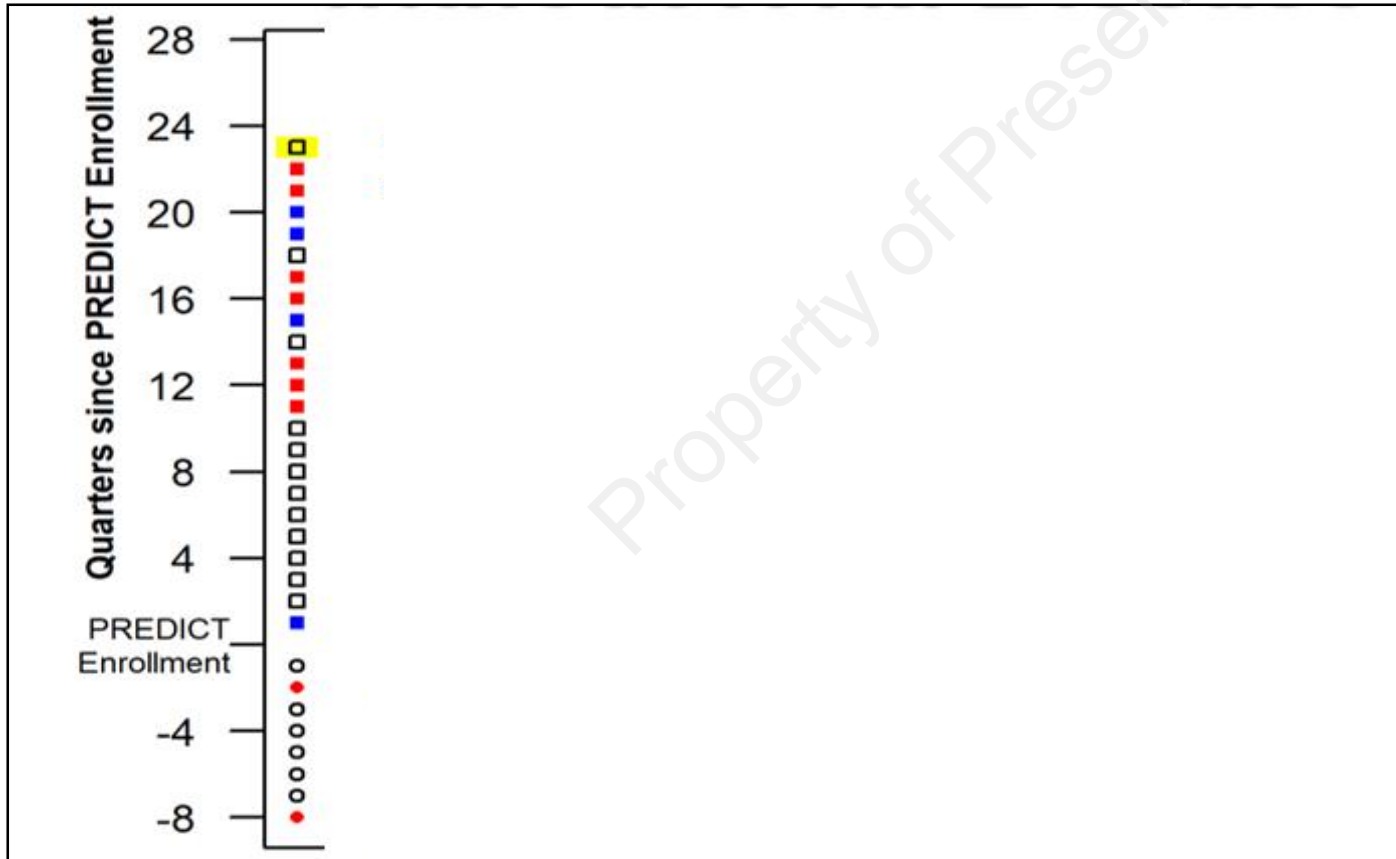
*Respiratory symptom domain is 14 points lower after NTM disease onset prior to treatment, 95% CI 8 to 20, $p < 0.001$.

**Respiratory symptom domain 10 points higher post treatment in those with NTM disease, 95% CI 6 to 13, $p < 0.001$.

PATIENCE: Change in FEV1 % Predicted



Culture Data among PREDICT Subjects without NTM Disease

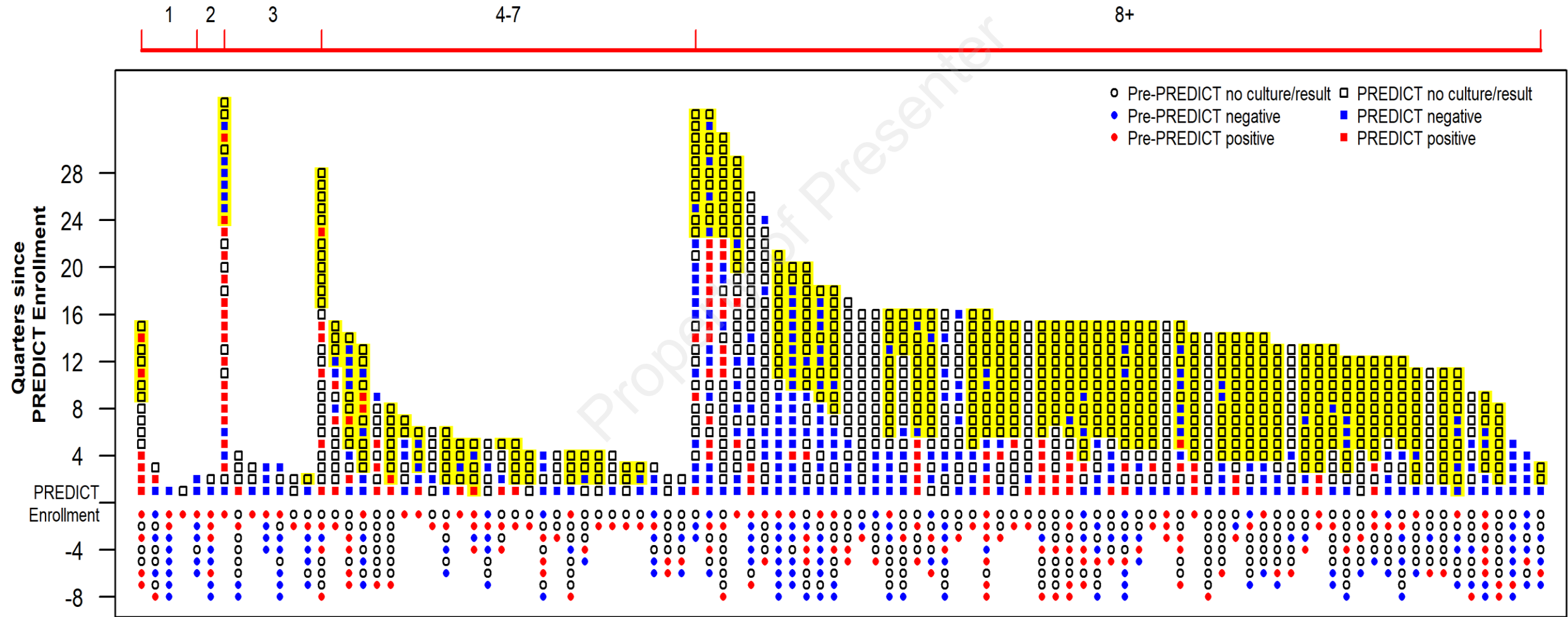


Culture results: PREDICT subjects without NTM Disease

MAC: No NTM Disease

Only 13% of participants have had a positive culture in the past year

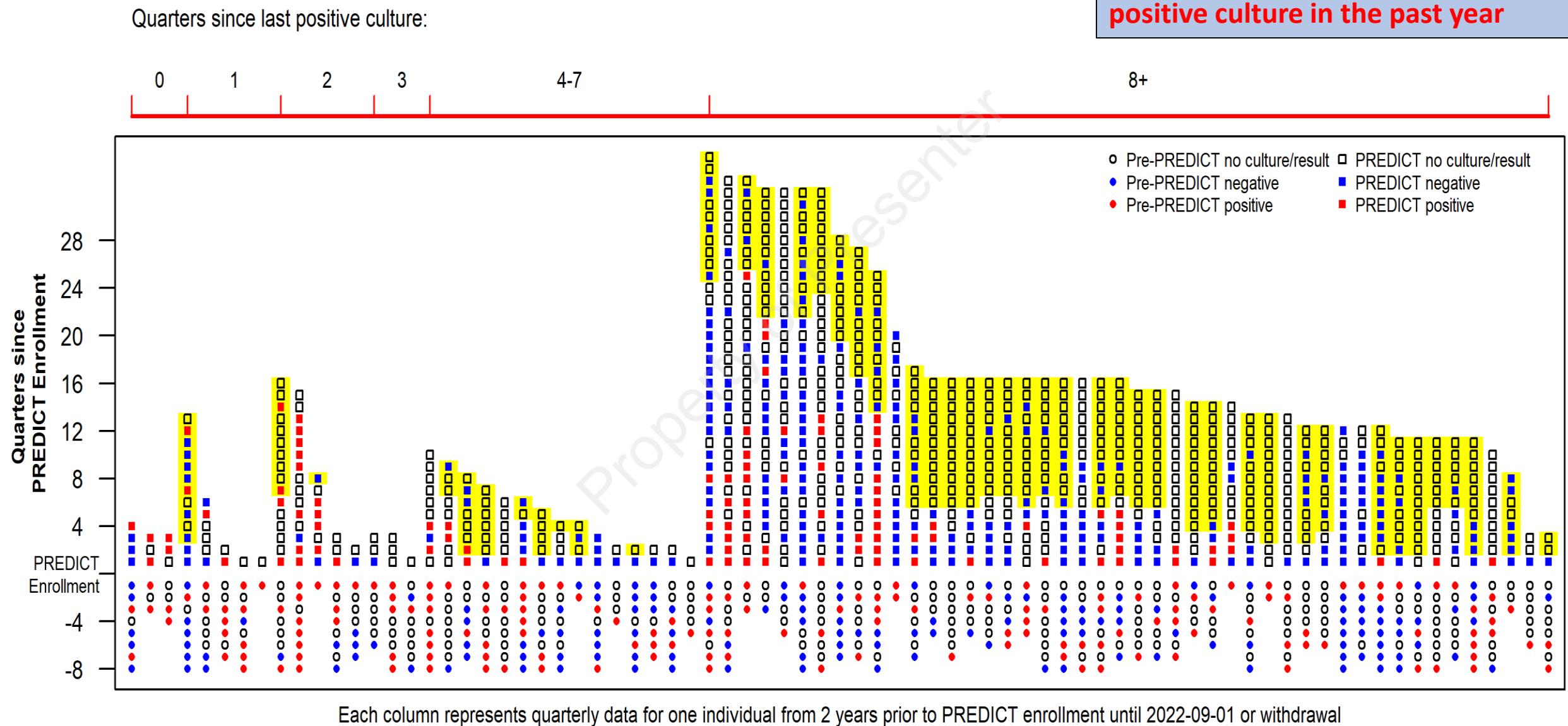
Quarters since last positive culture:



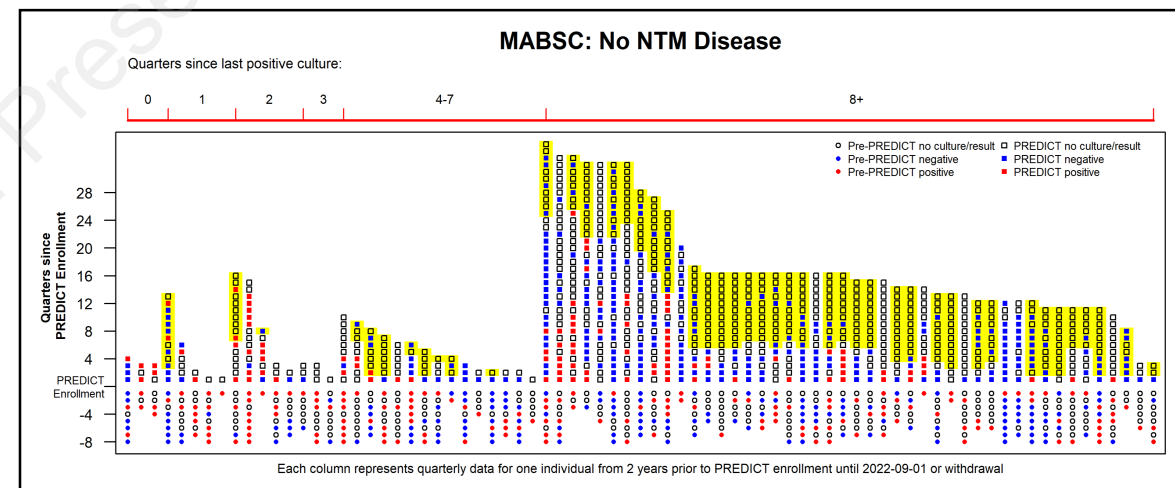
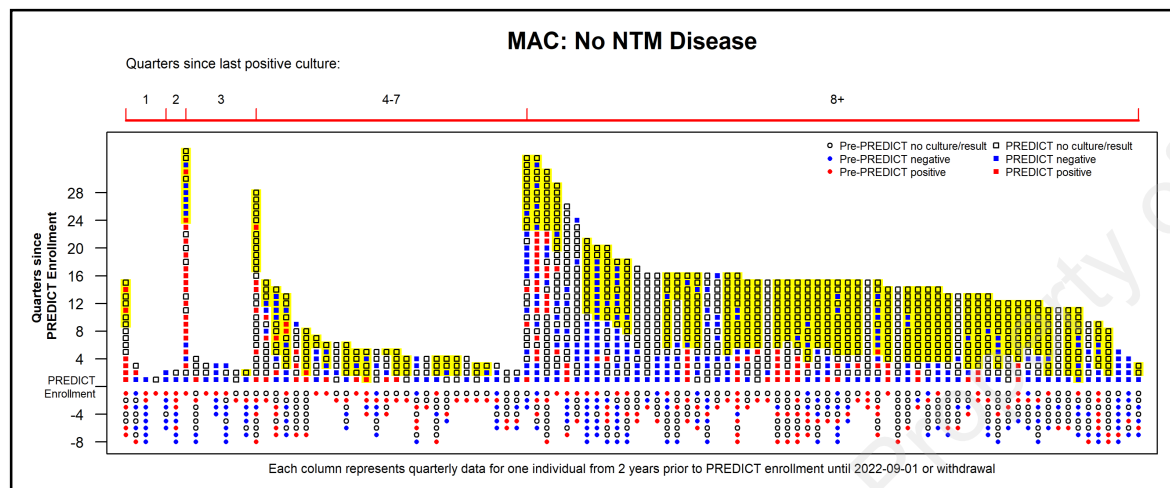
Culture results: PREDICT subjects without NTM Disease

MABSC: No NTM Disease

Only 21% of participants have had a positive culture in the past year



Culture Data among PREDICT Subjects without NTM Disease

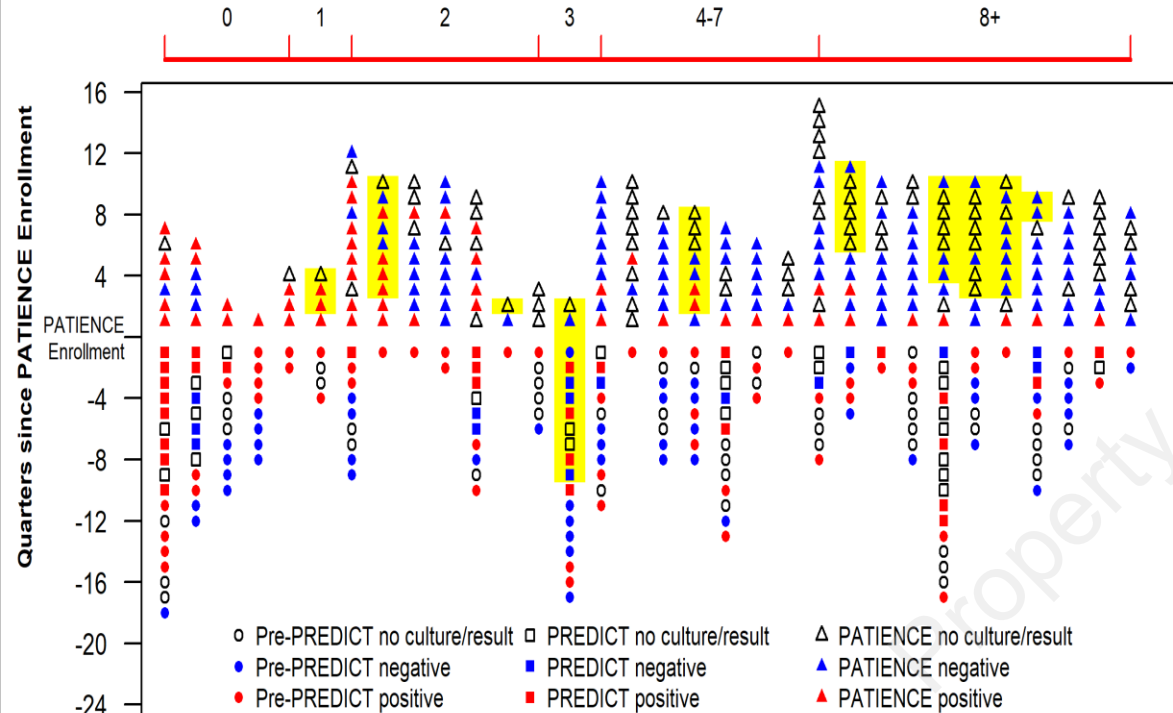


- 23% of participants have had NTM culture obtained in the last 6 months
- 44% of participants have had NTM culture obtained in the last year
- Most missing cultures likely due to elexacaftor/tezacaftor/ivacaftor (Trikafta)

Culture Data: Treatment Response

MAC: PATIENCE

Quarters since last positive culture:

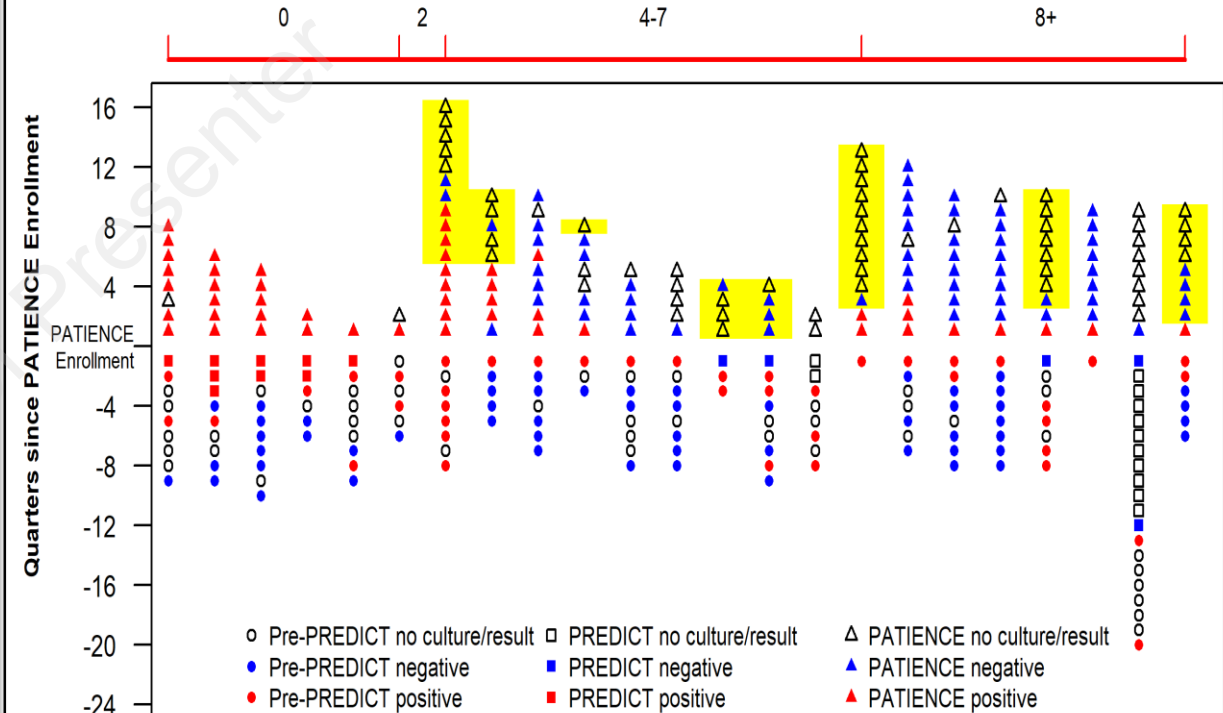


Each column represents quarterly data for one individual
from 2 years prior to PREDICT enrollment until 2022-09-01, 1 year after treatment end date, or withdrawal

MAC (n=33)
 34% culture negative > 2 years
 69% most recent culture negative
 13% remain culture positive
 6% inconclusive due to missing cultures

MABSC: PATIENCE

Quarters since last positive culture:



Each column represents quarterly data for one individual
from 2 years prior to PREDICT enrollment until 2022-09-01, 1 year after treatment end date, or withdrawal

***M. abscessus* (n=22)**
 35% culture negative > 2 years
 70% most recent culture negative
 22% remain culture positive
 4% inconclusive due to missing cultures

So what's wrong with NTM airway cultures?

(context of CF lung disease)

Low sensitivity due to low NTM burden

- Reduced yield due to sputum decontamination
- NTM disease (PATIENCE Trial) 65% positive
- Indolent infection (PREDICT Trial) 38% positive
- Following E/T/I (PREDICT Trial) 18% positive

Lack of sputum samples

- Expecterated sputum rare in children and adults with mild disease
- Dramatic reduction due to E/T/I
- BAL not recommended for asymptomatic screening

The reasons we need biomarkers are also why validation is so challenging!



Colorado NTM Core Clinical Research Service

- Analysis of **unprocessed airway samples**
- **Culture** and **molecular identification** of isolates
- Expanded and custom **drug susceptibility testing**
- SOP for **remote sputum sample collection**
- NTM **biorepository**
- Custom **PK/PD** testing
- Custom **whole genome sequencing (WGS)** of isolates
- WGS **comparisons** to extensive CF and non-CF isolates
- **Database coordination** with WGS and phenotypes
- Clinical trial **design and outcome** analysis

Clinical Trials Assisted by the NTM NRCs

PREDICT Trial (NCT02073409) and **PATIENCE** Trial (NCT02419989) (Nick, Martiniano)

PAINLESS Trial: (NCT04324088)(Nick)

PIVOT Trial: **P**rospective evaluation of saliva **c**ompared to sputum.

ABATE Trial: A Phase 1b, Multi-center Study of IV Gallium Nitrate in Patients with Cystic Fibrosis who are Colonized with Nontuberculous Mycobacteria (CFF and FDA) (Goss, Nick, Singh)

HALT NTM Study: (NCT04024423)(Gross)

ENCORE Trial: (NCT03597347) (Nick)

FORMaT Trial: (ACTRN12618001831279p) (Wainwright)

Improving Treatment of NTM Infection in CF (NCT02372383) (Martiniano)

Protocol For **Bacteriophage Treatment** of *Mycobacterium abscessus* through An Investigational New Drug Application (IND)

27 additional planned or pre-clinical trials also receiving NTM NRC assistance

NTM Outcome Measure Advancement Core

- Custom **PK/PD assays** from sputum, tissue, other biosamples
- Gene or isolate testing for **pre- and post-trial changes** in DST
- **Consulting** on preclinical studies (industry & academics)
- **Phylogenetic analysis** of NTM isolates to distinguish between reactivation, reinfection or polyclonal infections
- Panels of CF and non-CF **NTM isolates** for drug candidate screening.
- Quantitative/ culture independent assessment of **NTM bacterial burden**
- Provide **in vitro** testing of agents against NTM
 - biofilm phenotype, intracellular systems, mouse models



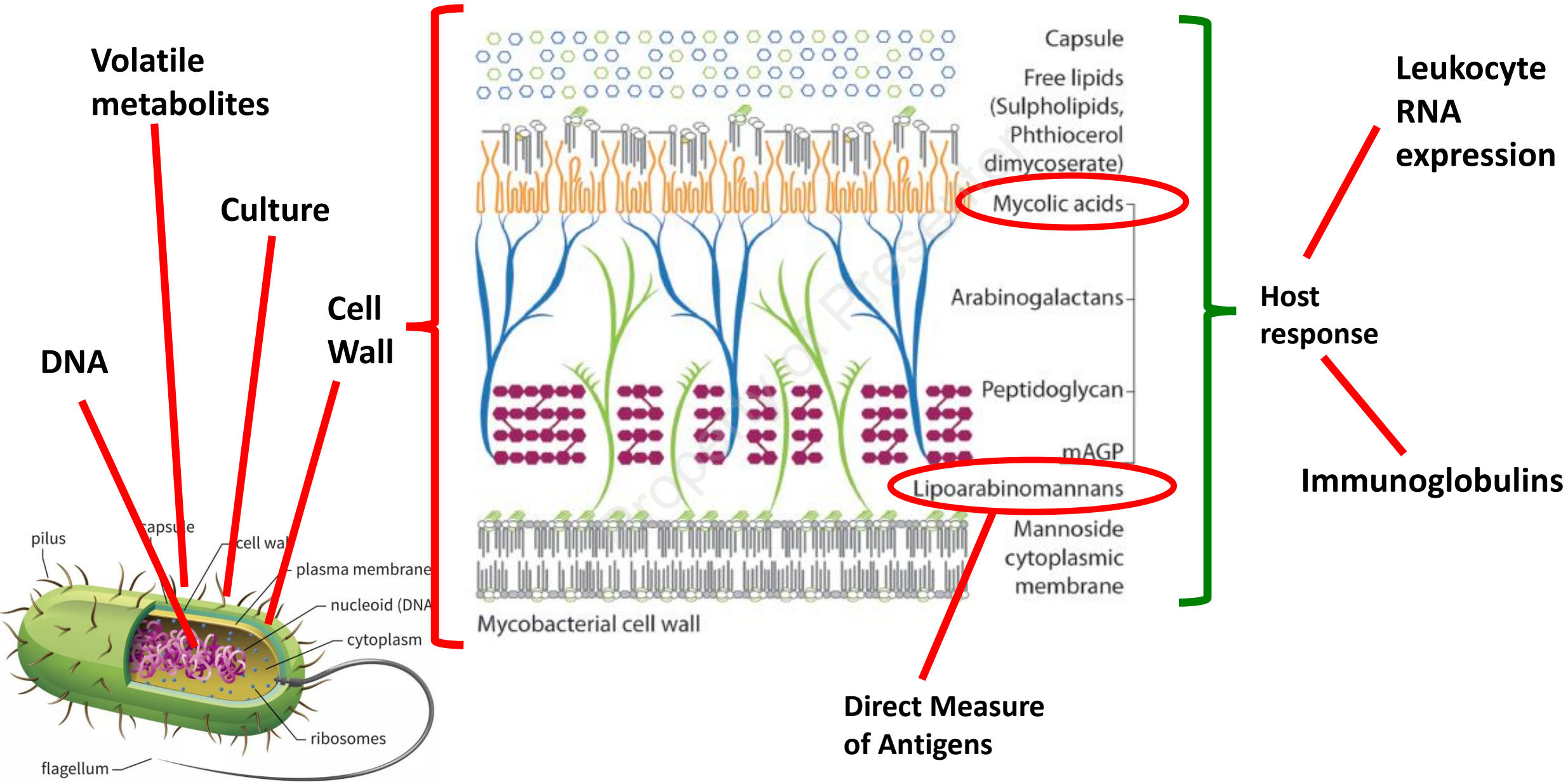
J Nick, MD

C Daley, MD

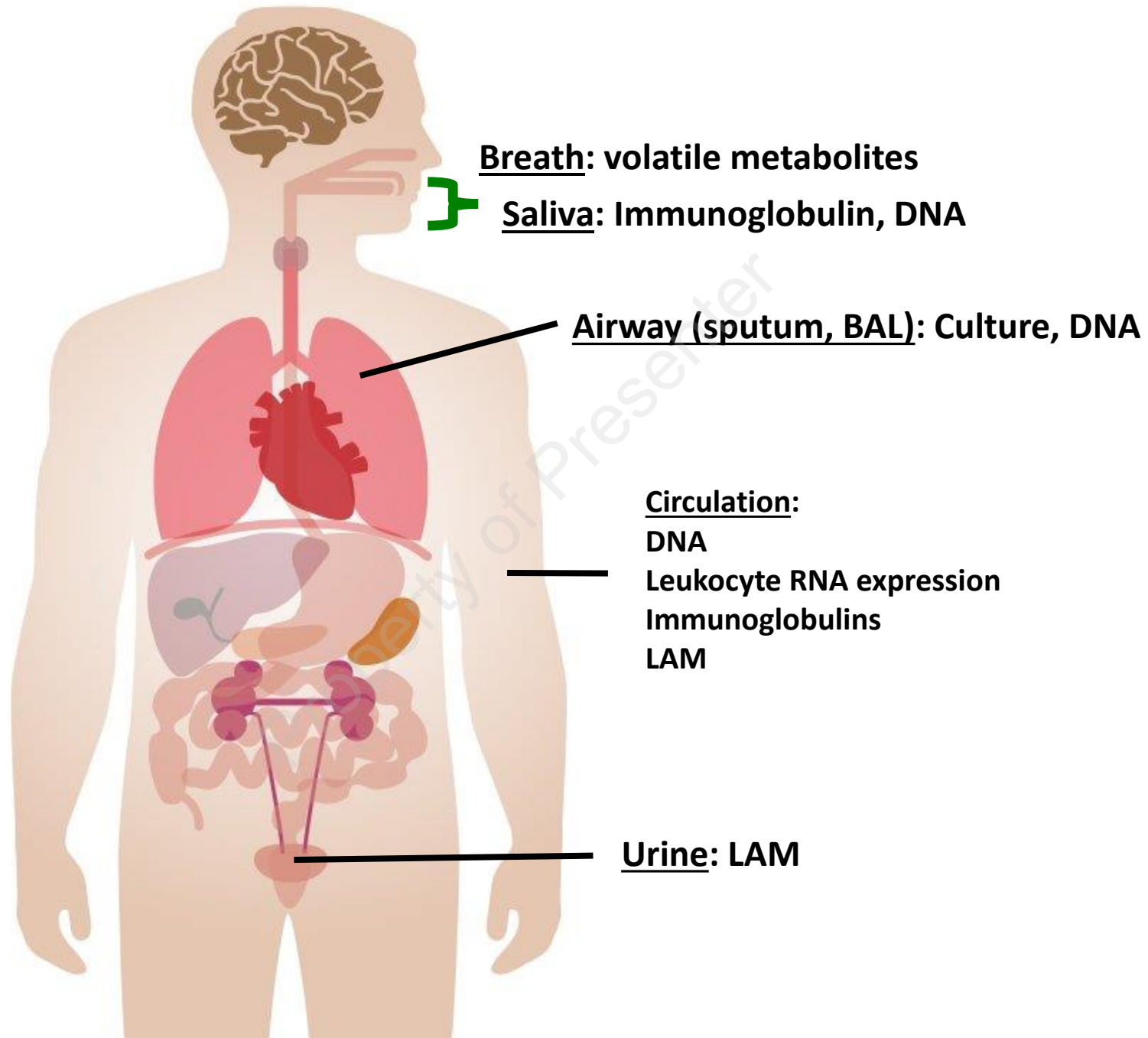
M Strong, PhD

R Davidson, PhD

Targets for NTM Detection



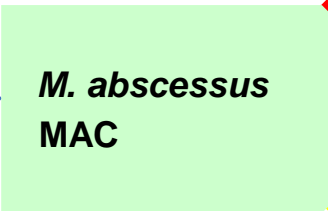
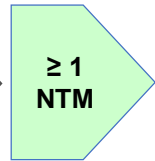
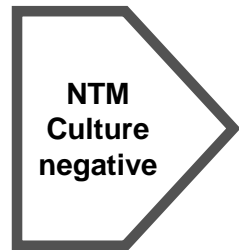
Sites and Specimens for NTM Detection



Utilizing prospectively identified NTM cohorts to replace single sputum cultures in marker studies

PIVOT Trial

Saliva collection from
PAINLESS, PREDICT & PATIENCE

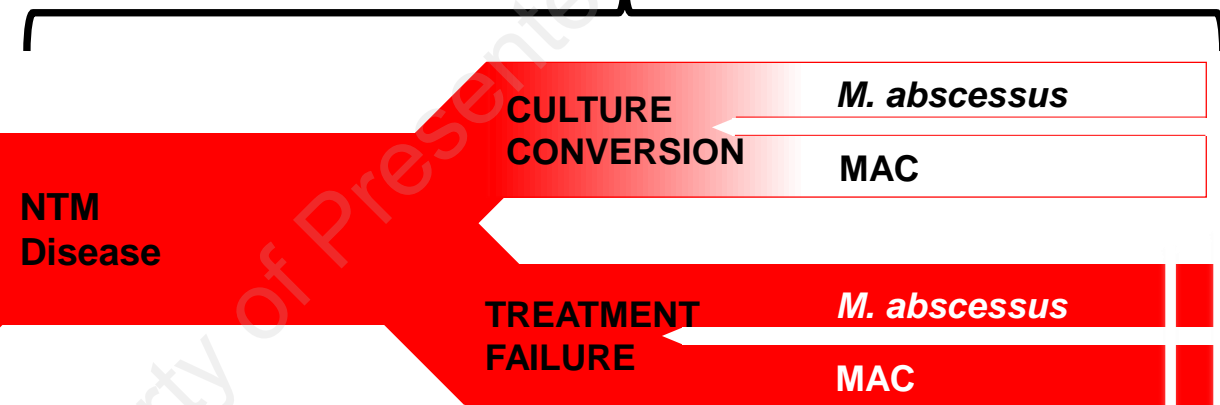


PREDICT
Trial

PAINLESS Trial: Prospective
Analysis of urine LAM to
Eliminate NTM Sputum
Screening (CF) (NCT04324088)

PATIENCE Trial

NTM Disease and Response to Treatment



Compassionate
use of
bacteriophage
treatment of *M.*
abscessus
through an IND

Indolent or
Transient

PREDICT Trial
Indolent Infection



Biomarker studies through Colorado NTM Outcome Measure Advancement Core NRC

NTM Markers Currently Being Tested in Trials

NTM genome: Colorado Adult P&P Award

- **WGS** (Michael Strong, PhD, Rebecca Davidson, PhD NJH).

Radiographic predictors: CFF CRSP Award

- **HRCT** (Stacey Martiniano, MD, CHCO, David Lynch, MD, NJH).



Sputum: NIH-funded ancillary study (R01 HL146228)

- **Microbiome** (Rebecca Davidson, PhD, NJH).
- **Volatile sputum metabolites** (Jane Hill, PhD, University of British Columbia).

Urine: CFF IRI Clinical Trial Award (PAINLESS Trial)

- **Urine lipoarabinomannan** (Delphi Chatterjee, PhD, CSU).

Saliva: CFF Clinical Trial Award (pending)(PIVOT Trial)

- **Targeted amplicon** (Rebecca Davidson, PhD, NJH).
- **Antibodies** (Kara Calhoun, MD, UCD).

NTM Markers Under Evaluation to be Added to Trials

Breath: CFF Clinical Pilot Award

- **Volatile breath metabolites** (Jane Hill, PhD, University of British Columbia).

Whole Blood: CFF Clinical Pilot Award

- **Circulating leukocyte RNA signatures** (Mimi Saavedra, MD, NJH).

Plasma: CFF Clinical Pilot Award



- **Circulating DNA signatures** (Pradeep Singh, MD, Steve Salipante, MD, PhD, University of Washington).

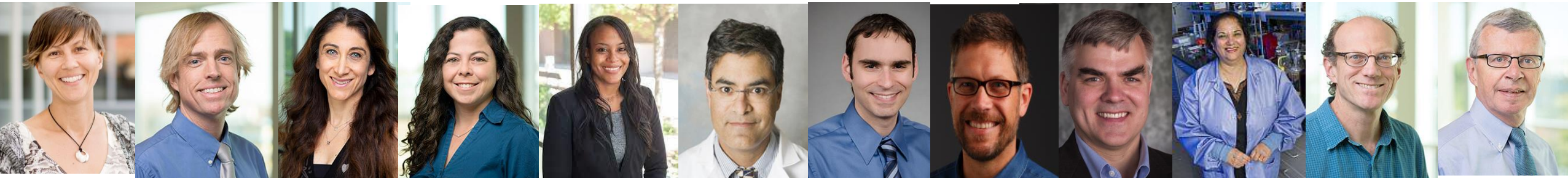
Serum: CFF Clinical Pilot Award

- **Antibodies and inflammatory markers** (Ken Malcolm, PhD, NJH).
- **Mycolic Acid Antibodies** (Diagnostig, UK).
- **Cholesterol metabolites** (Jen Philips, MD, PhD, Washington Univ).

Sputum



- **Bacterial RNA (R/S ratio)** (Nick Walter, MD, PhD, Martin Voskuil, PhD, University of Colorado)



Jane Hill, PhD M. Strong, PhD M. Saavedra, MD R. Davidson, PhD K. Calhoun, MD P. Singh, MD S. Salipante, MD, PhD Walter, MD, PhD Voskuil PhD D. Chatterjee, PhD Malcolm, PhD Lynch, MD

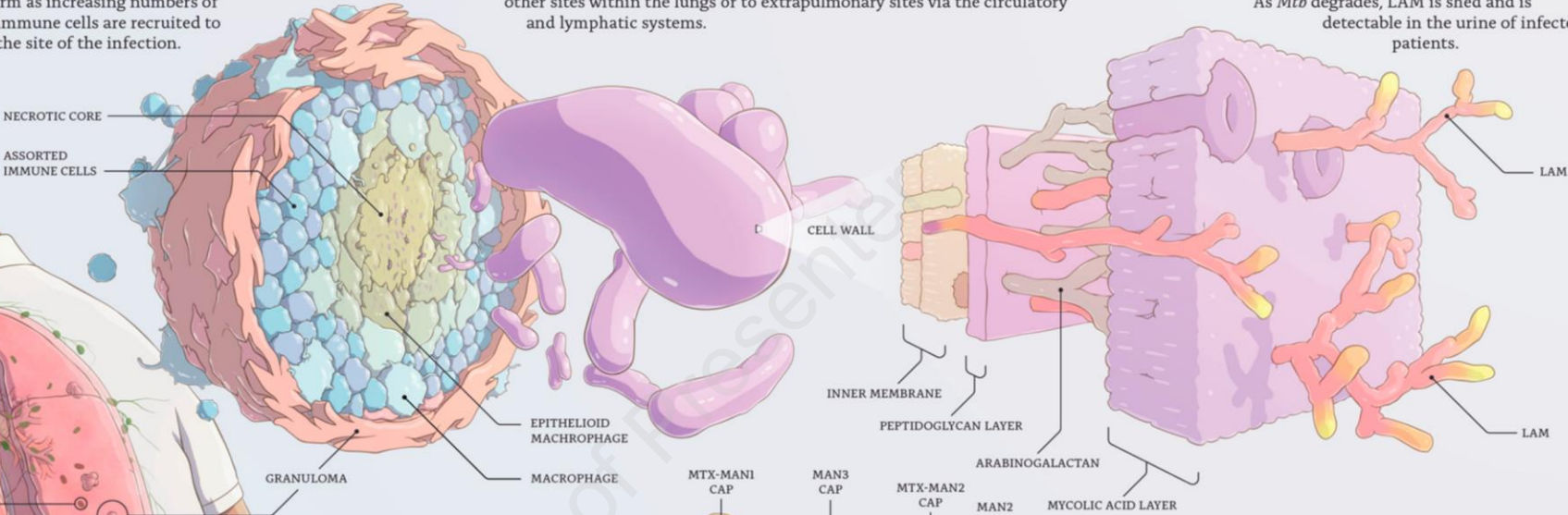
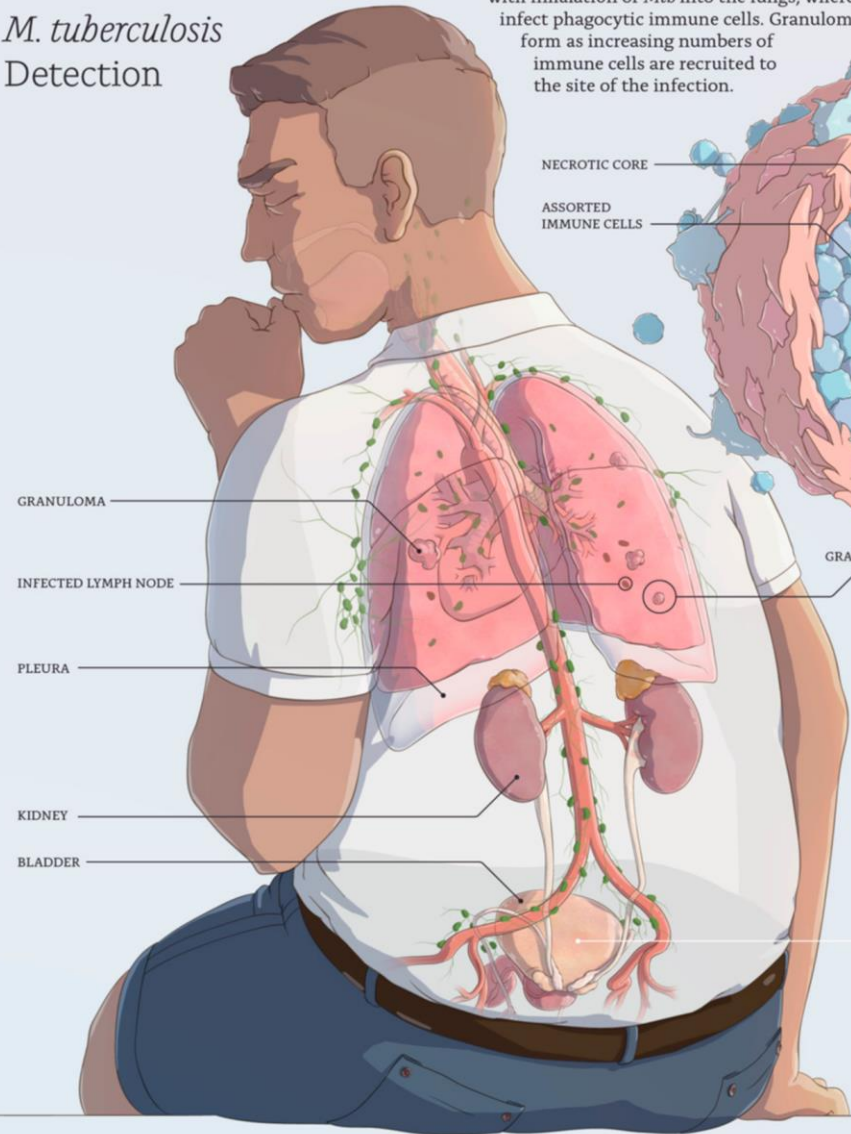
Urine Lipoarabinomannan (LAM)

Urine LAM Tests for *M. tuberculosis* Detection

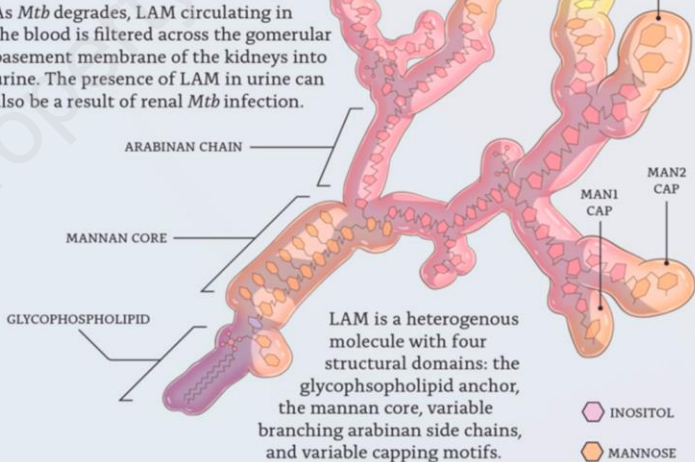
Mycobacterium tuberculosis (*Mtb*) infection begins with inhalation of *Mtb* into the lungs, where they infect phagocytic immune cells. Granulomas form as increasing numbers of immune cells are recruited to the site of the infection.

Granulomas form a barrier that may prevent the further spread of *Mtb*. However, they may also allow *Mtb* to continue infecting recruited phagocytes and thus continue replicating. If the bacterial loads become too great, *Mtb* can break out and spread to other sites within the lungs or to extrapulmonary sites via the circulatory and lymphatic systems.

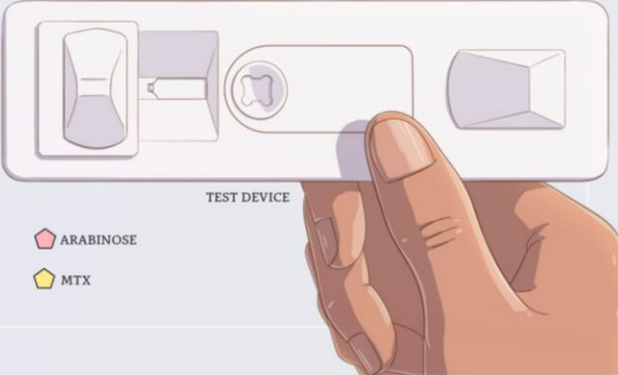
Mtb has a unique cell wall that contains lipoarabinomannan (LAM). LAM is firmly but non-covalently attached to the membrane. As *Mtb* degrades, LAM is shed and is detectable in the urine of infected patients.



As *Mtb* degrades, LAM circulating in the blood is filtered across the glomerular basement membrane of the kidneys into urine. The presence of LAM in urine can also be a result of renal *Mtb* infection.



LAM is detected using immunoassay-based rapid diagnostic tests. Next-generation tests use signal amplification technology to detect very low concentrations of LAM with increased diagnostic sensitivity. To maintain high specificity, tests need to target *Mtb*-specific structural LAM-domains like MTX-mannose caps.



Urine LAM

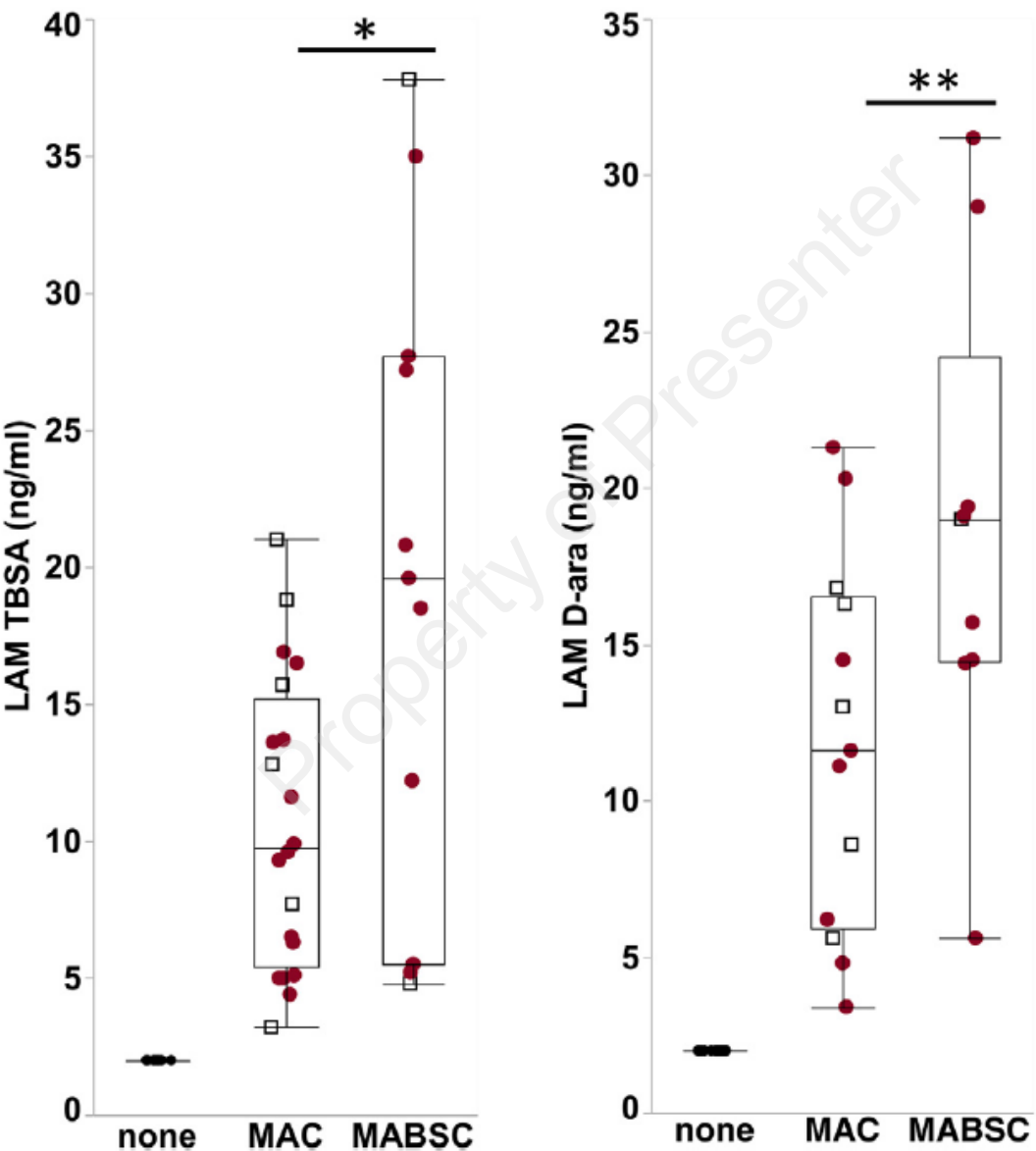
Advantages

- Sample is readily available from all subjects
- Test is well validated in TB among individuals coinfecting with HIV
- May reflect global infectious burden (all lobes)

Challenges

- Very low burden of infection in CF versus TB
- LAM from NTM differs from TB
- Generally below LOD for approved immunoassays
- Nonspecific for species or subspecies of NTM

Urine lipoarabinomannan (D-ara or TBSA) analyzed by GC/MS correlates with past NTM culture history



D. Chatterjee (CSU)

PAINLESS Trial: Prospective Analysis of urine LAM to Eliminate NTM Sputum Screening (CF) (NCT04324088)

- Never culture positive of NTM by chart review **and** minimum of 3 cultures in last 3 years
- Annual urine LAM compared to sputum cultures during clinical care
- Electronic consent
- Sputum collection by mail (or in person)



E. Armantrout, RN



V. Lovell, RN



PAINLESS Trial: Prospective Analysis of uriNe LAM to Eliminate NTM Sputum Screening (NCT04324088)

Concordance of a positive urine LAM result and with previous airway cultures for NTM

	Urine LAM Pos (+)	Urine LAM Neg (-)
Pos. NTM History	n=4 True Pos.	n=0 False Neg.
Neg. NTM History	n=10 False Pos.	n=96 True Neg.

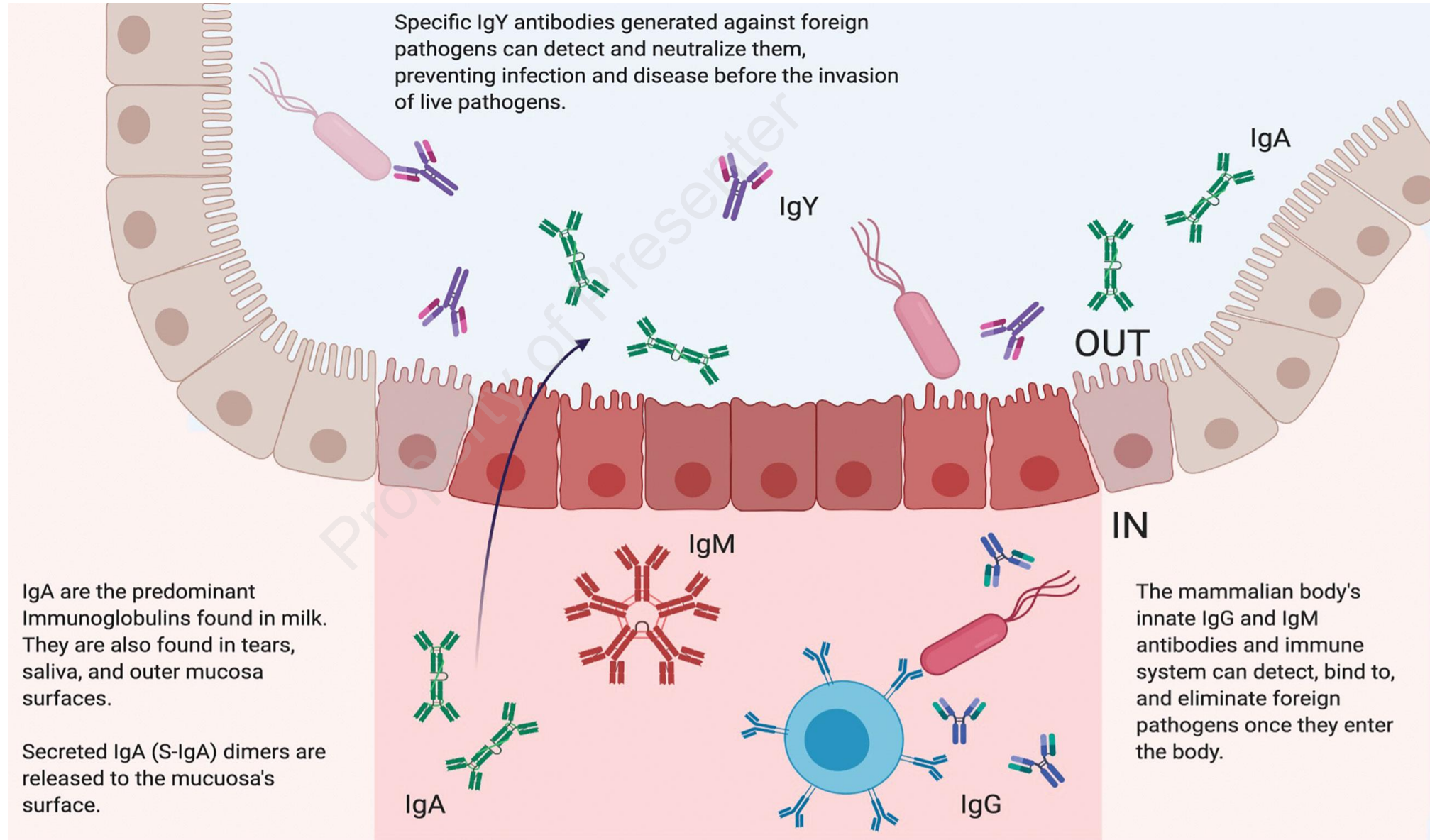
Negative Predictive Power = 100%
Sensitivity = 100%
Specificity = 90.6%
Positive Predictive Power = 28.6%

Next Steps

CFF-NICK20A0 (Nick, PI) Urine
Lipoarabinomannan as a Marker for Low-risk of NTM Airway Infection –**cap at n=100**
Expansion of urine LAM screening to multiple sites

Longitudinal Assessment of Urine Lipoarabinomannan as a Culture-Independent Systemic Marker of Pulmonary Nontuberculous Mycobacteria *Urine LAM analysis from PREDICT and PATIENCE*

Immunoglobulin titers (serum and/or saliva)



Immunoglobulin titers (serum and/or saliva)

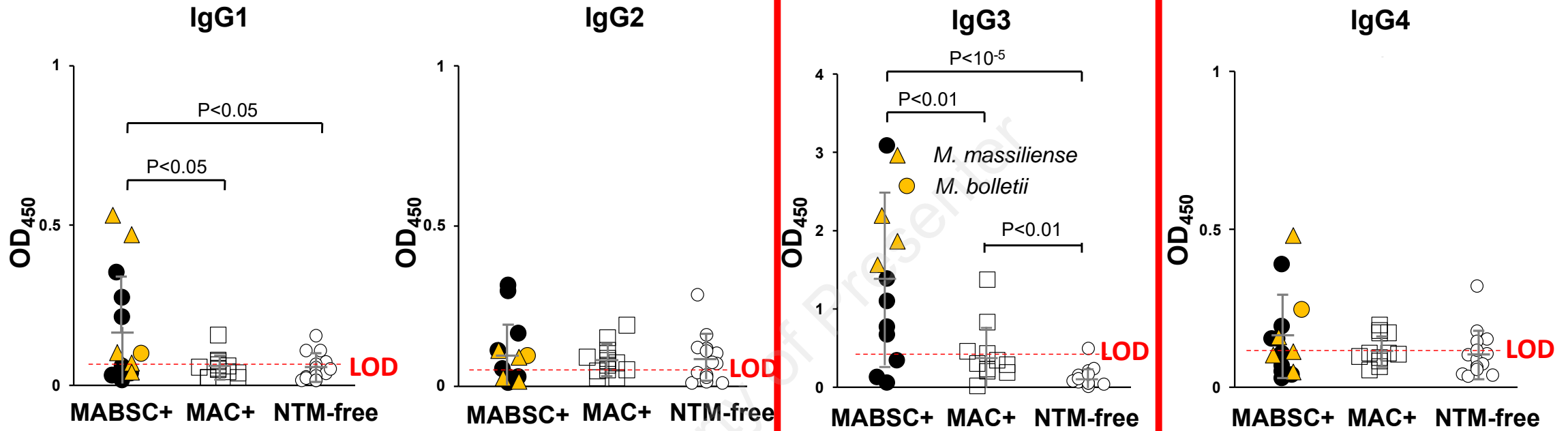
Advantages

- Sample(s) are readily available from all subjects
- Potentially highly sensitive

Challenges

- Depends on functioning immune system
- Nonspecific for species

Serum IgG titers

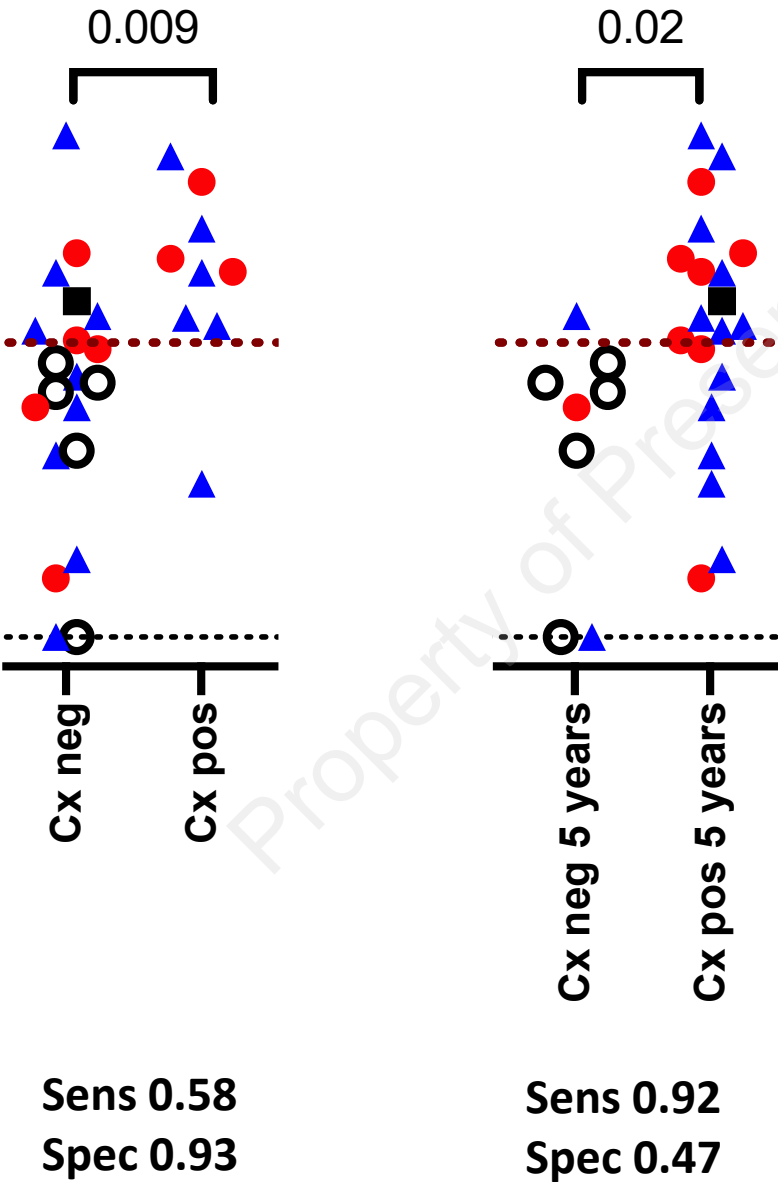


ELISA using NTM whole cell lysates
Culture status over a 5-year period
Dual infections assigned to MABSC



K. Malcolm (NJH)

Saliva: IgG + IgA against NTM cell lysates



K. Calhoun, MD

Molecular Detection of NTM from raw sputum (and saliva)

1. Quantitative real-time PCR (qPCR)

- *M. abscessus* or MAC assays
- Bacterial DNA quantification compared to standard curve

2. Targeted Amplicon Sequencing Panel

- Simultaneous detection of multiple NTM species
- More sensitive than qPCR

Advantages:

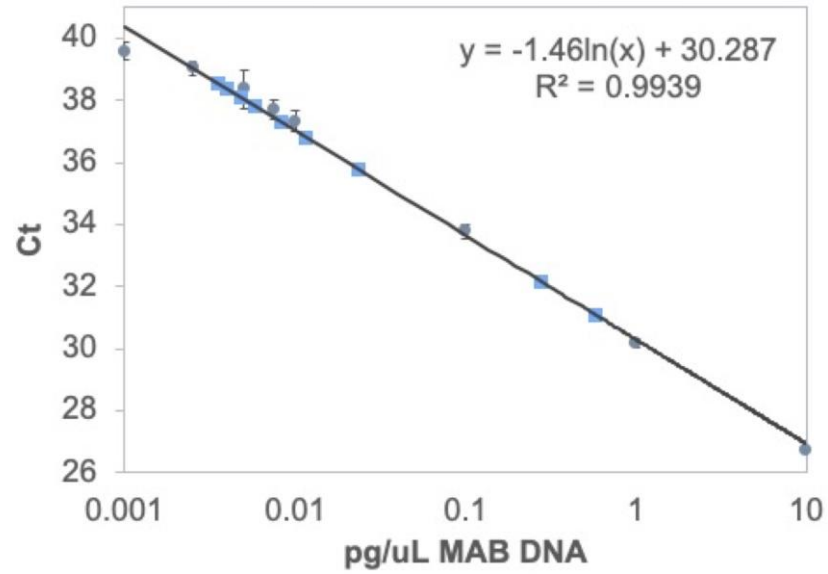
- Minimal processing of samples (no decontamination)
- Low DNA input for assays
- Samples are direct from the site of infection
- Easy to obtain (saliva)
- Highly sensitive for species identification

Challenges:

- Very low burden of infection (saliva)
- Low availability of samples (sputum)
- Unable to distinguish between living/dead bacteria
- Water contaminants in saliva

M. abscessus qPCR tested in 120 sputum samples

- from 86 subjects in the PREDICT and PATIENCE trials



• STD Curve ■ Sputum Sample — Log Curve

High cycle threshold but
Excellent fit within standard curve

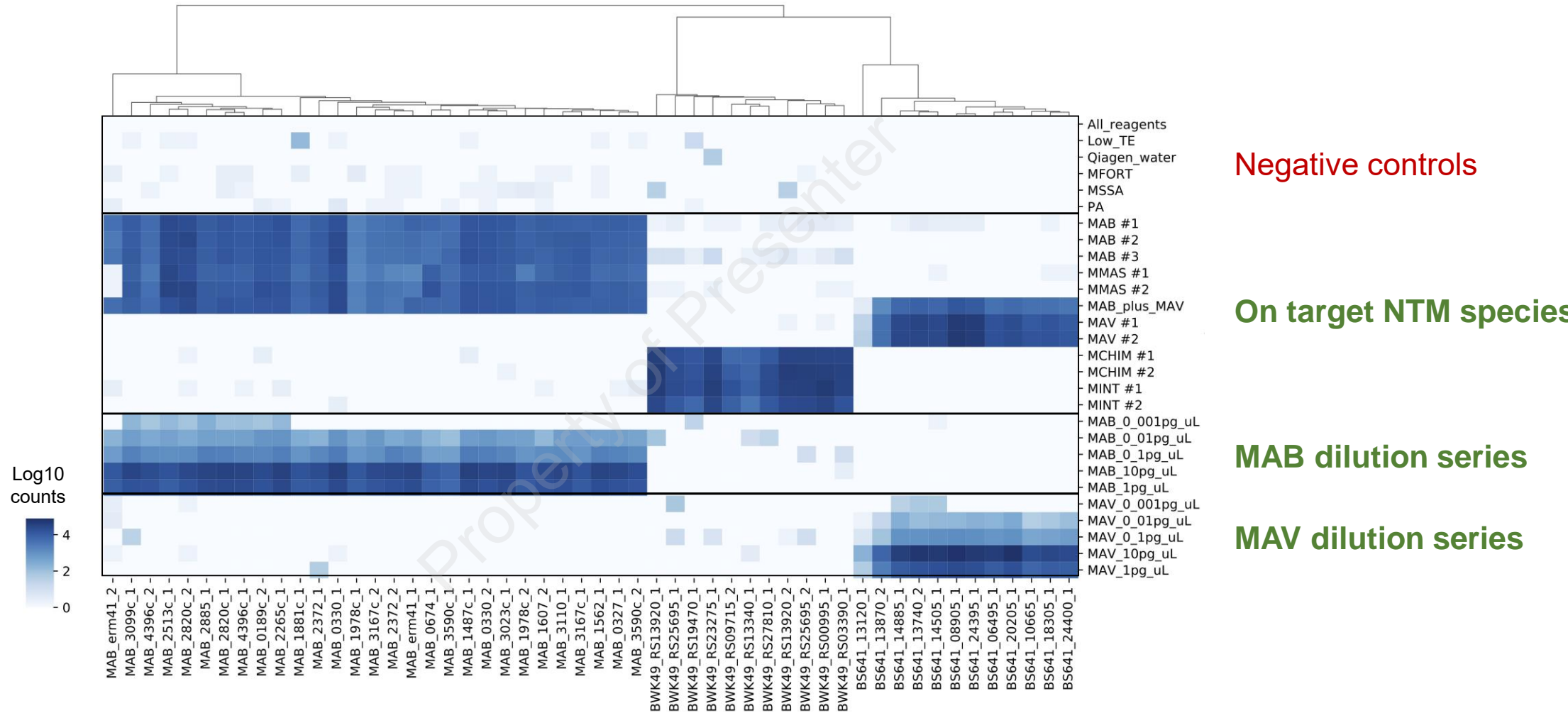
	Sens	Spec	PPV	NPV
Matched culture (n=90)	0.79	0.91	0.61	0.96



R. Davidson (NJH)

Targeted Amplicon Sequencing for NTM

59 amplicons in sequencing panel



Negative controls

On target NTM species

MAB dilution series

MAV dilution series

Columns = amplicons
Rows = QI samples



R. Davidson (NJH)

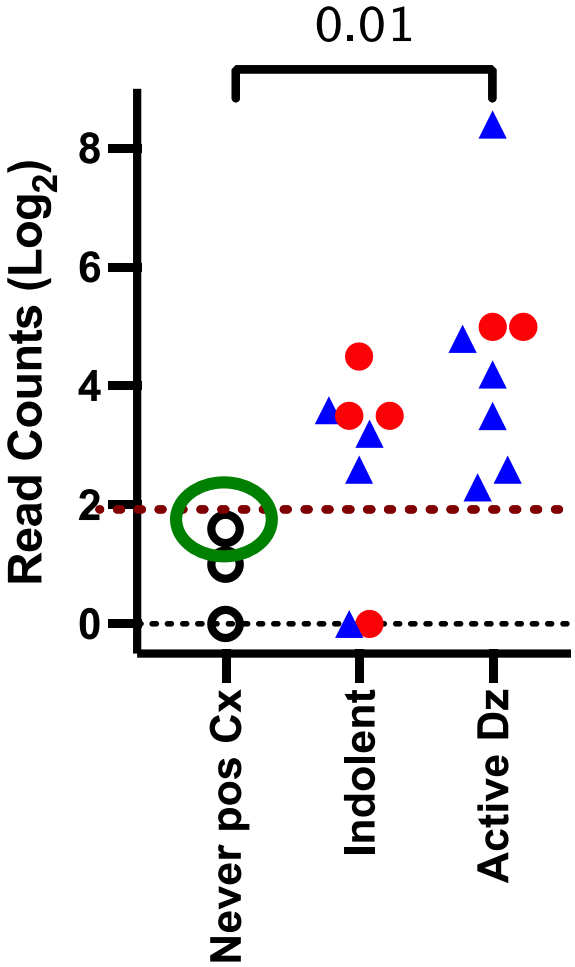
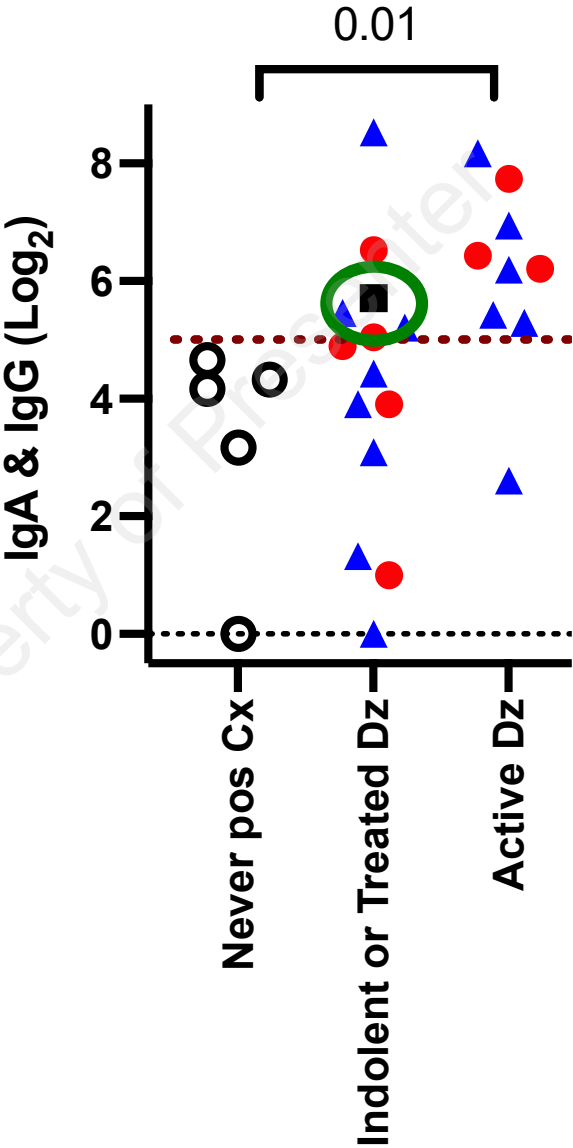
Combined Immunoglobulin and Molecular markers for NTM in saliva

Sum of Ig and Mol positive
Active Dz Sensitivity 0.80

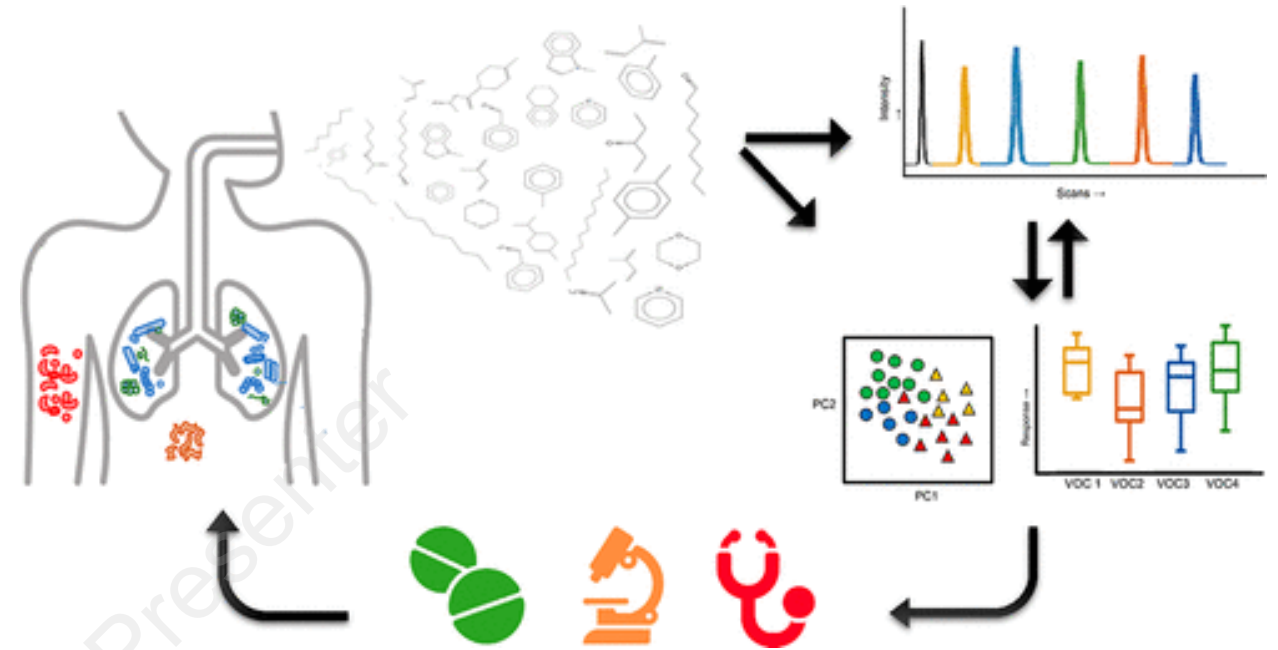
Either Ig or Mol positive
Specificity 1.0



Subject with *M. lentiflavum* correctly identified as having a positive NTM culture by Immunoglobulin assay, but neither MAC or *M. abscessus* by molecular assay.



Volatile Molecules (breath or sputum)



Advantages

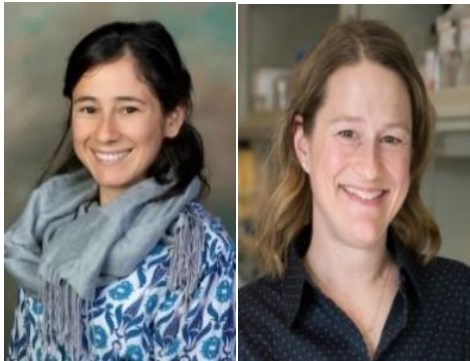
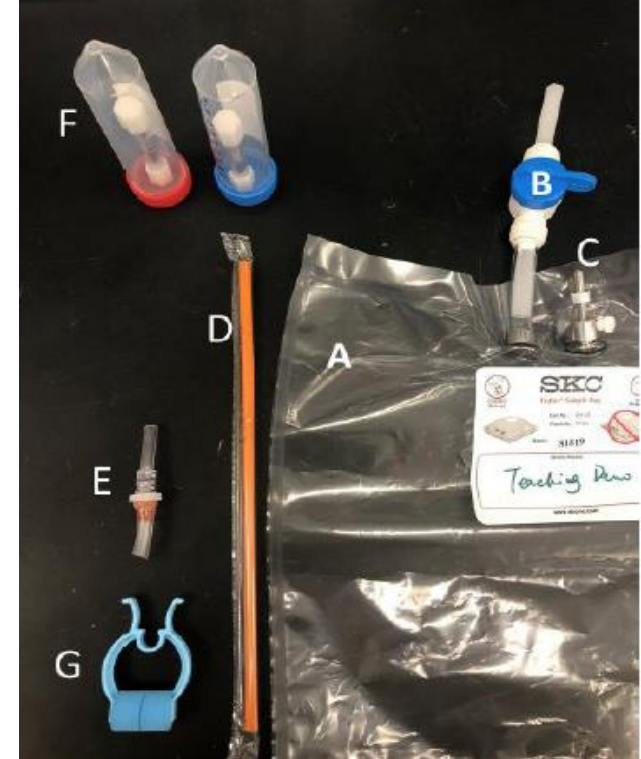
- Non-invasive sample from airway
- Can be applied to sputum samples
- Potentially highly sensitive
- Successful reports in TB and a variety of airway infections

Challenges

- Method and interpretation not standardized or widely available

Volatile Molecules (breath or sputum)

- Subjects breath into 1.5L Tedlar bags
- Breath is drawn through a filter onto 3-bed thermal desorption tubes (TDT) via a vacuum pump
- TDT contents are desorbed at 330 °C into a cryogenically cooled (-120 °C) inlet liner
- Analyzed by 2-D gas chromatography time of flight mass spectrometer (GC×GC-TOFMS)
- Method can be modified to sample “headspace” of sputum in a tube

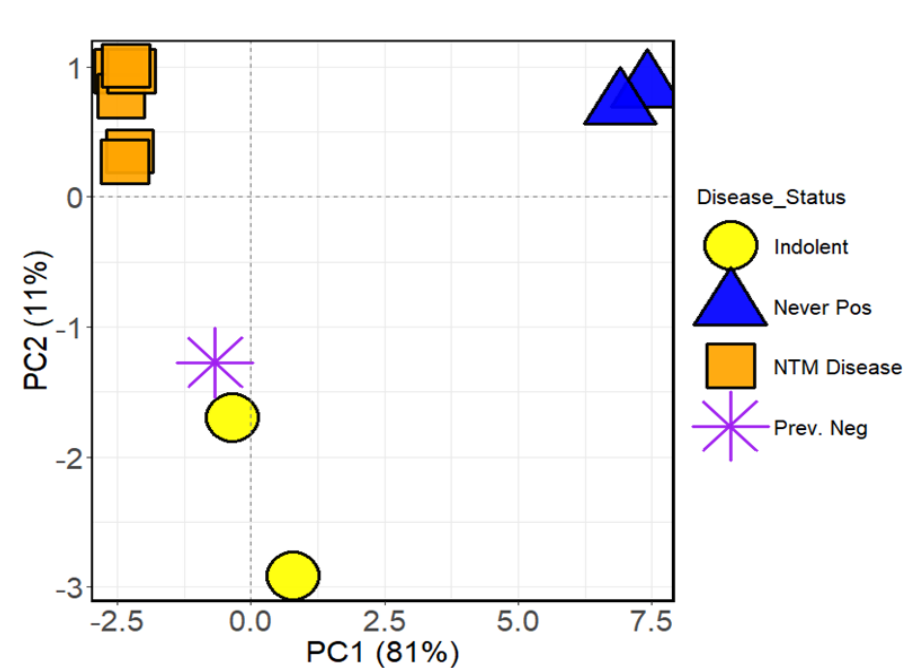


S. Caceres, MS

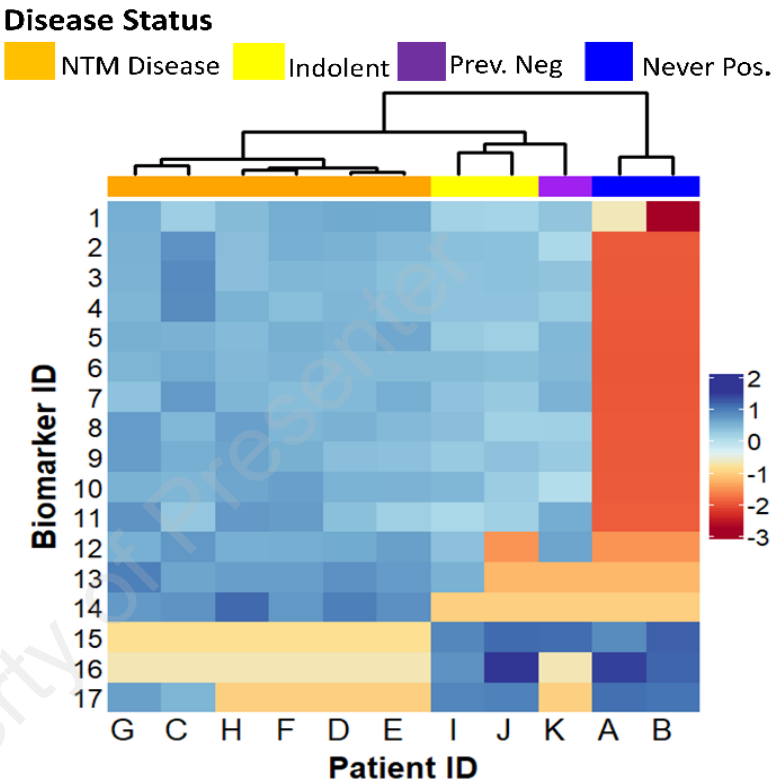


K. Poch, BS

Volatile molecules in exhaled breath distinguished between disease states



Principal component score plot constructed using the normalized, transformed and scaled expression profile of 17 features selected by the Boruta method



Dendrogram and heatmap using the 17-compound breathprint

Analysis of stored sputum from PREDICT and PATIENCE Trials
 R01HL146228



Jane Hill, (UBC)

Markers of Treatment Response?

26 y/o male with Cystic Fibrosis

- *M. abscessus* in 2016
- Enrolled in the PREDICT Trial
- Met diagnostic criteria in early 2017
- Enrolled in the PATIENCE Trial
- Refractory to antibiotic treatment
- Two effect phages identified
- Initiated on phage therapy Sept 2020
- Culture conversion January 2021
- Lung Transplant October 2021
- Completed phage (and antibiotics) March 2022

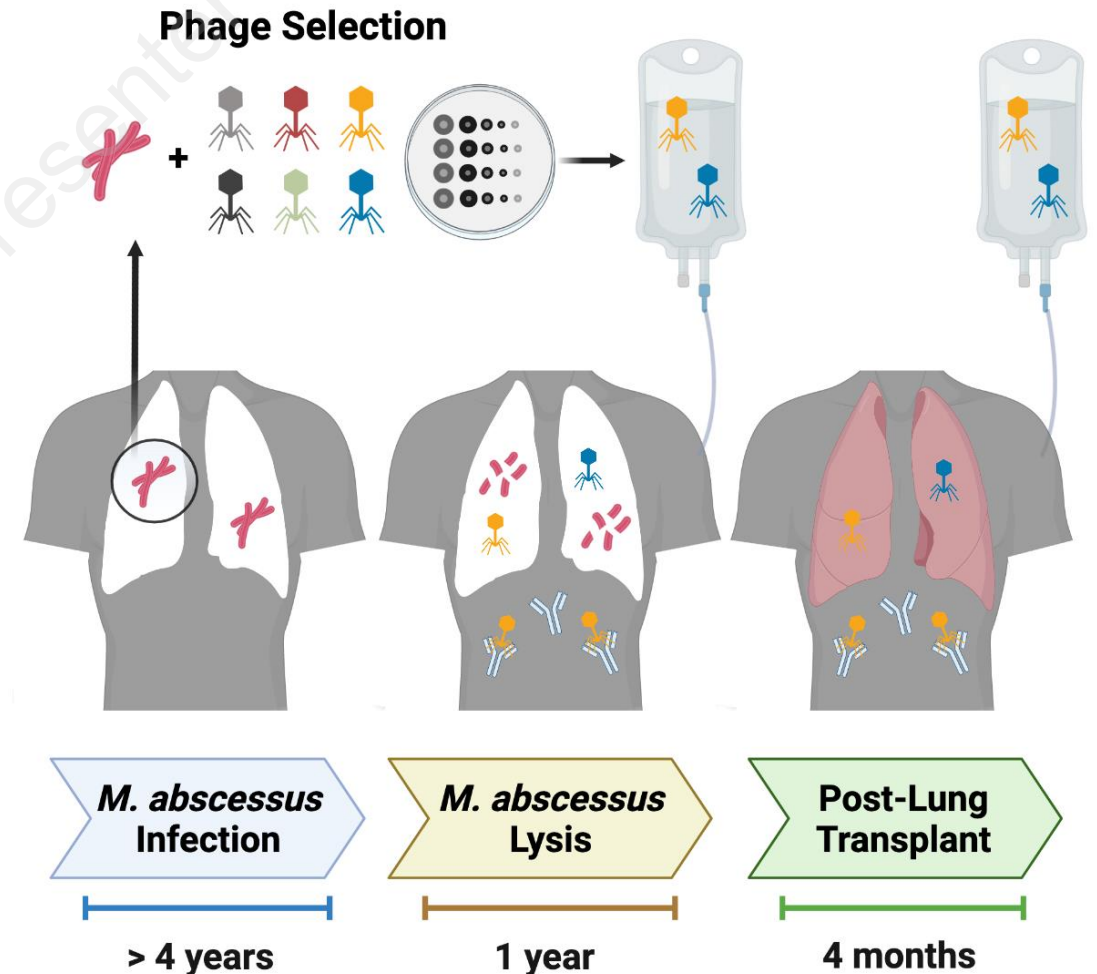


G. Hatfull (Pitt)



R. Dedrick (Pitt)

Host and pathogen response to bacteriophage engineered against *Mycobacterium abscessus* lung infection



Bacteriophages

- Natural predators of bacteria
- Most abundant organism on earth (10^{31})
- 10^{23} phage infections per second
- Highly specific host range-species/genus
- Incapable of infecting human cells

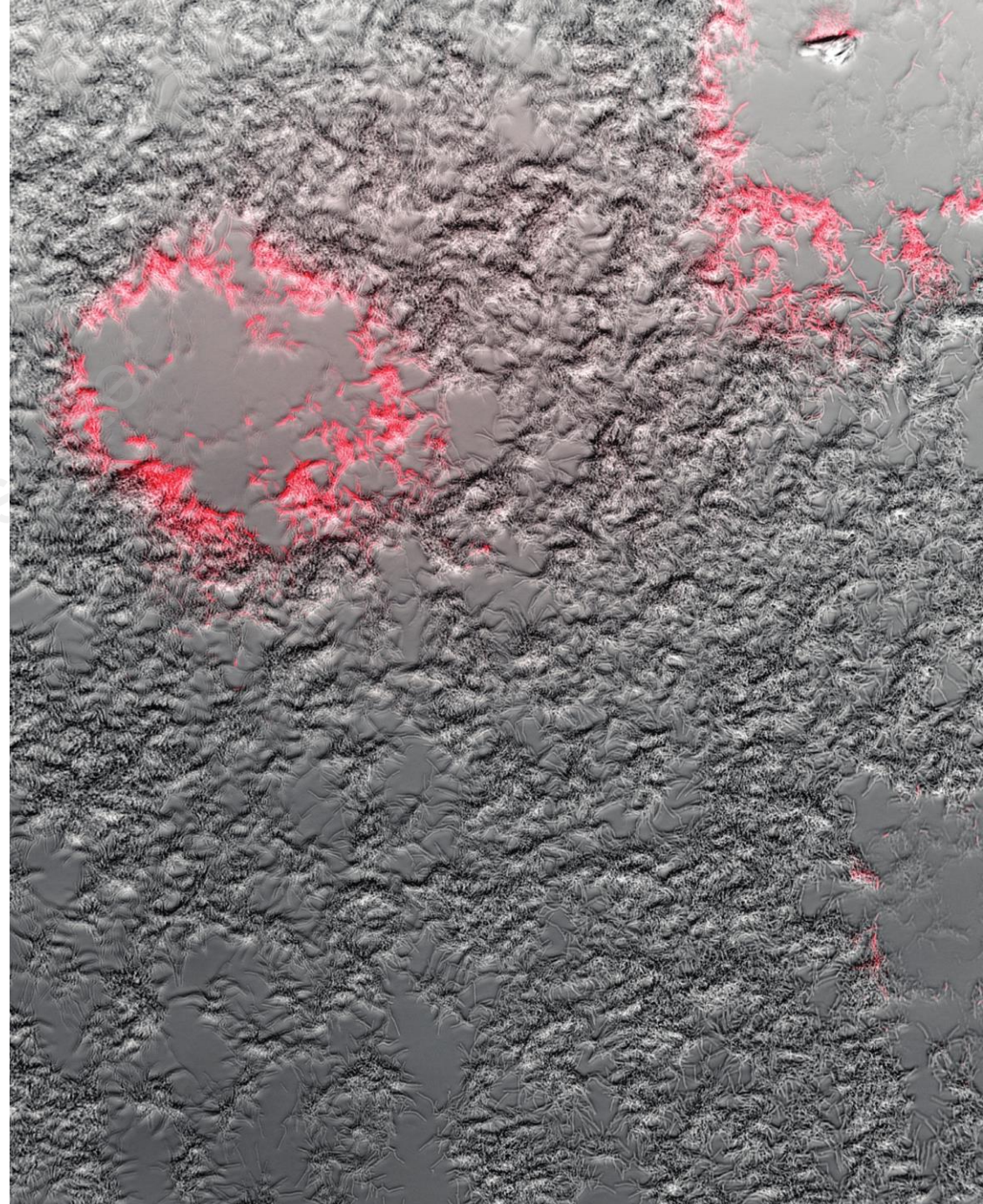
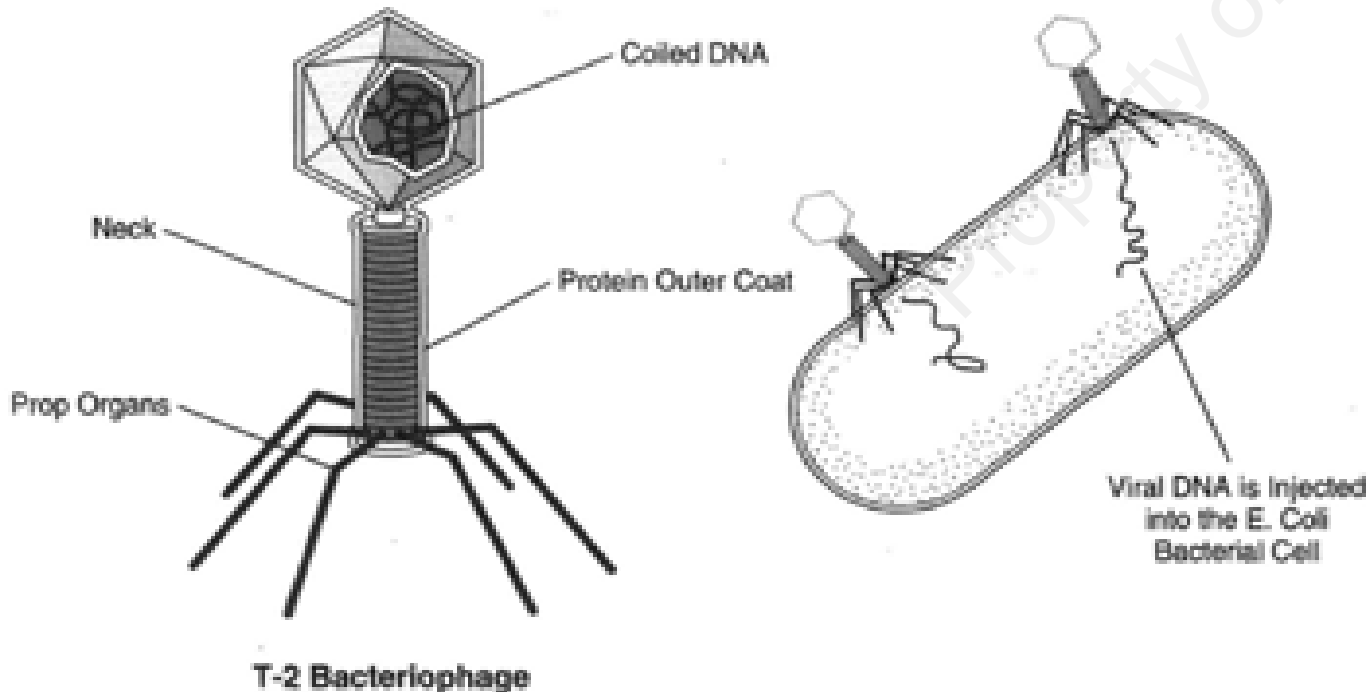
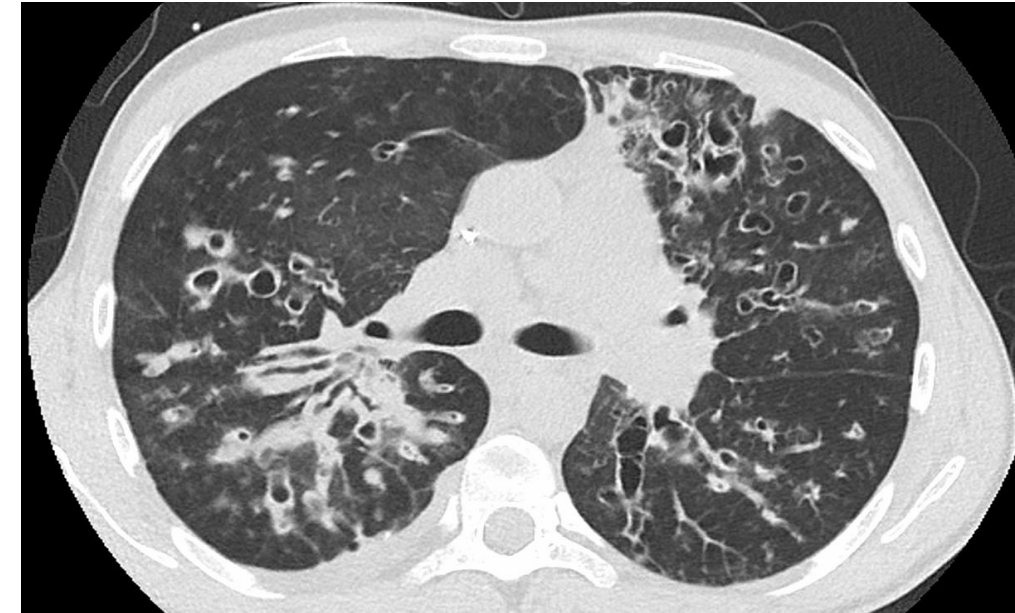


Image courtesy of Charles Dulberger and Carlos Guerrero-Bustamante

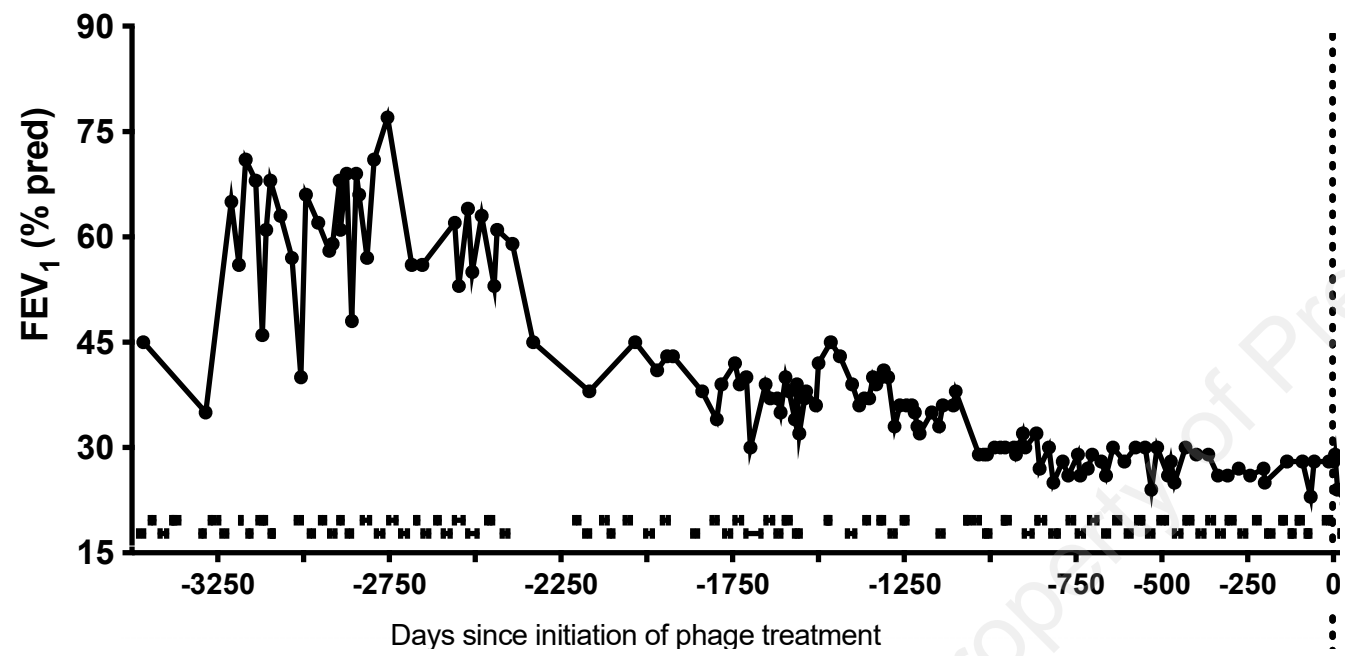
Case

JJ is 25-year-old man with cystic fibrosis (CF) and treatment refractory *Mycobacterium abscessus* lung disease

- Genotype H199Y/2184insA- no approved CFTR modulator
- Chronic infection with multi-drug resistant *P. aeruginosa* and methicillin-resistant *S. aureus*.
- Pancreatic insufficiency
- CF-related diabetes
- CF-related sinus disease
- Malnutrition with nighttime feeding via PEG
- Not currently listed for lung transplant



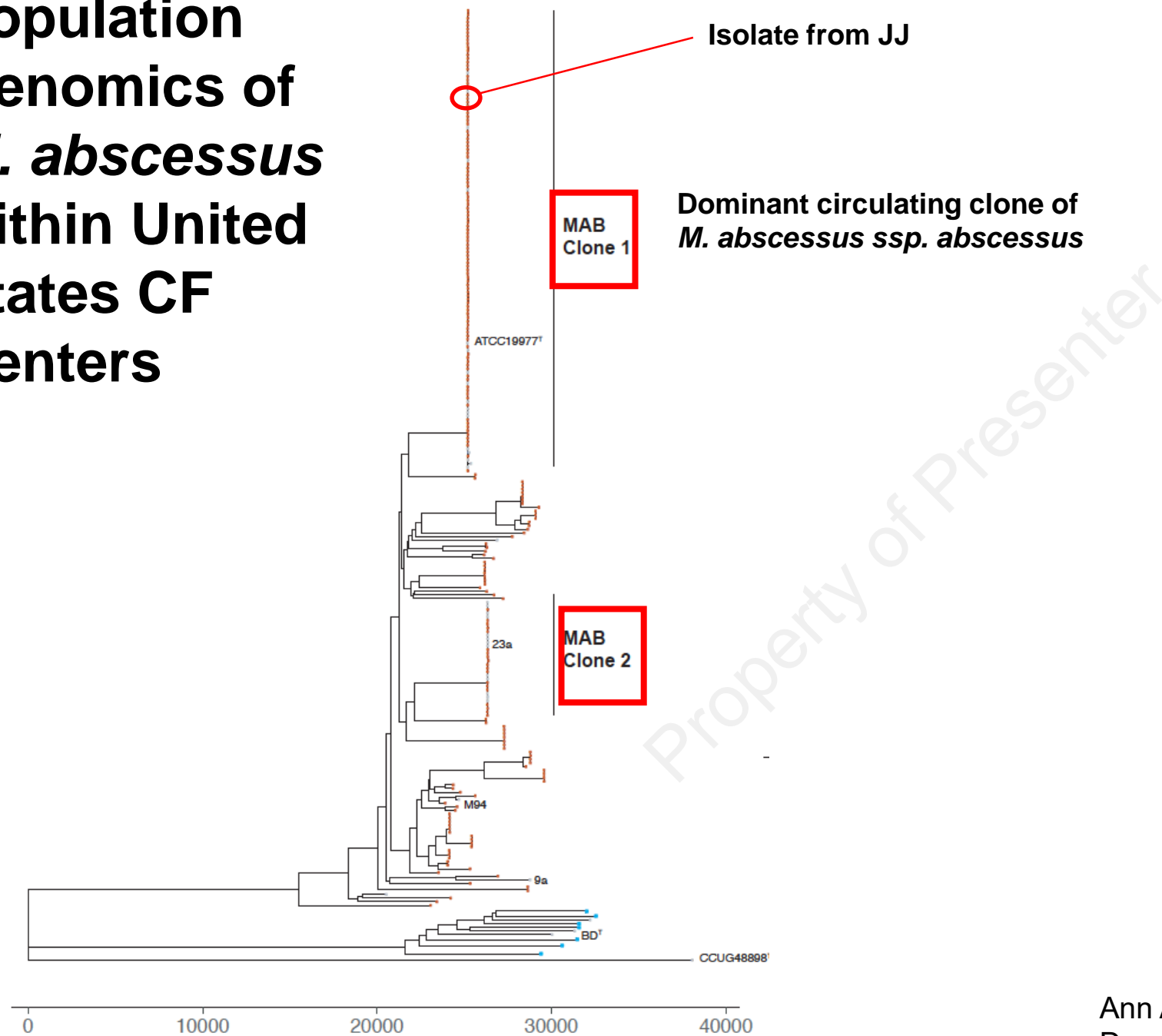
Clinical features



Lung Function
Forced Expiratory Volume in 1 second (FEV₁) (%predicted)

Hospitalizations

Population Genomics of *M. abscessus* within United States CF Centers

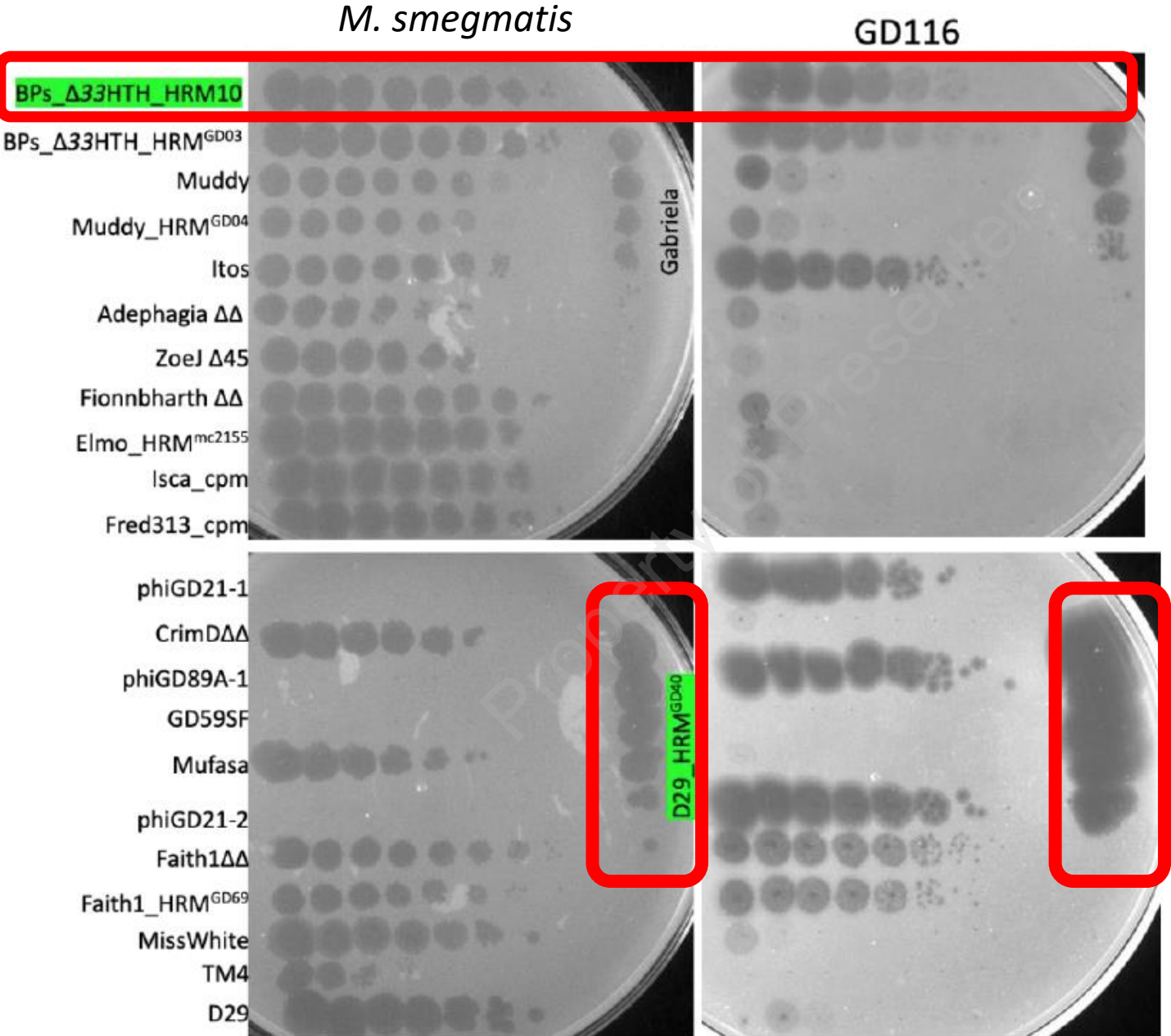


Ann Am Thorac Soc. 2021
Dec;18(12):1960-1969



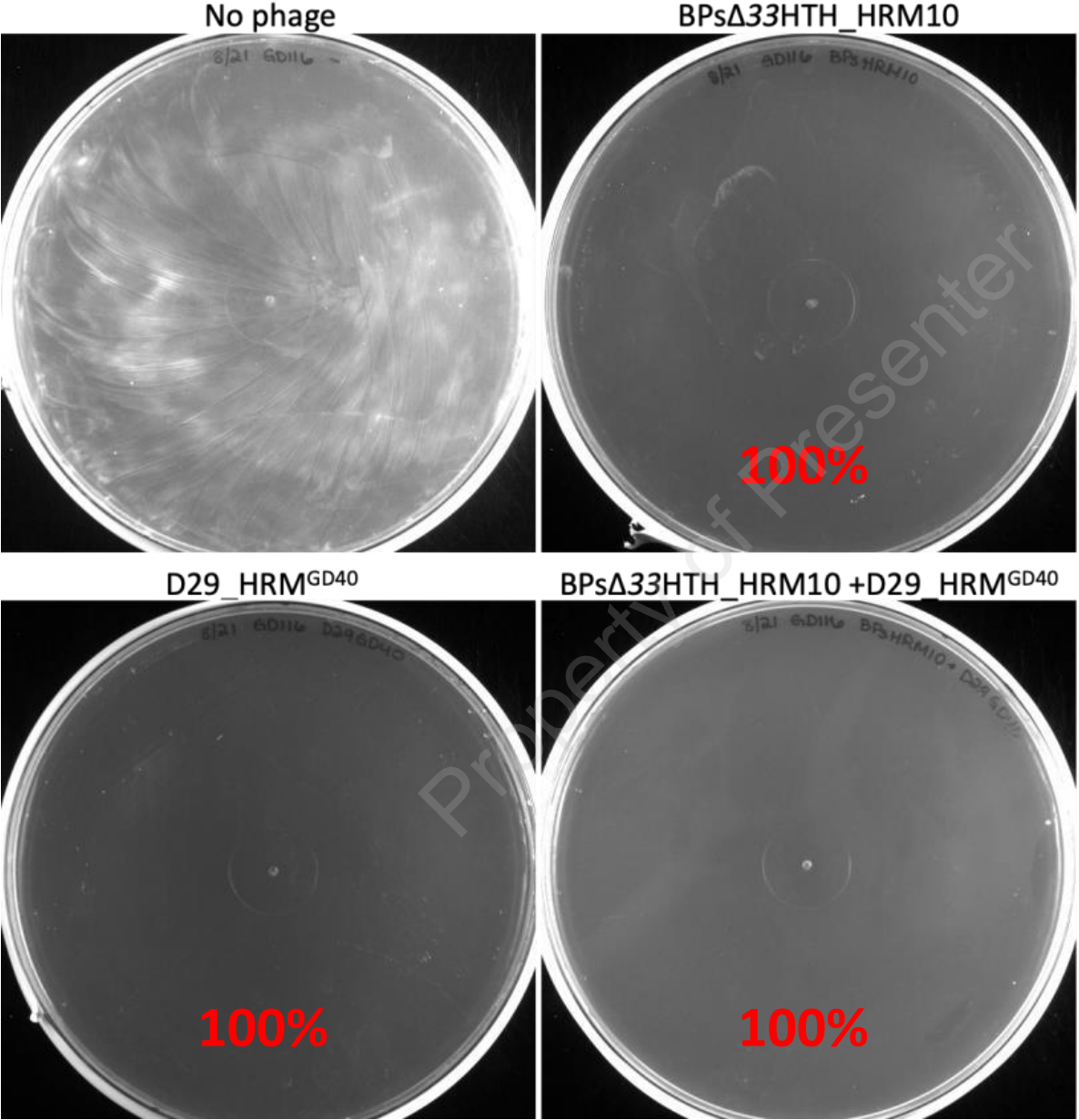
R. Davidson, PhD N. Hasan, PhD

Phage-selection



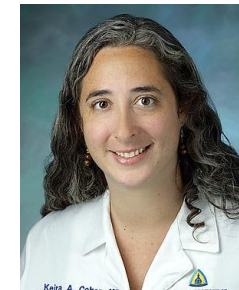
G. Hatfull (Pitt) R. Dedrick (Pitt)

Efficiencies of phage killing of *M. abscessus* GD116 *in vitro*



Regulatory Requirements for Bacteriophage Treatment of *M. abscessus*

- Material Transfer Agreement (MTA) between Univ. Pittsburgh and NJH
- Write a treatment protocol
- Investigational New Drug (IND) approval from the FDA
- IRB approval for phage administration
- Informed consent of the patient
- Protocol with St. Joseph Pharmacy to dilute the stock in PBS and package the phage in syringes
- IRB approval for banking of isolates (PREDICT and PATIENCE Trials)
- IRB approval for specimen collection
- Protocol for explanted lung analysis
- IRB approval for analysis of explanted lungs
- Biohazard protocol for phage administration at St. Joes and UCH



Keira A Cohen, M.D.



M. Jones, RN



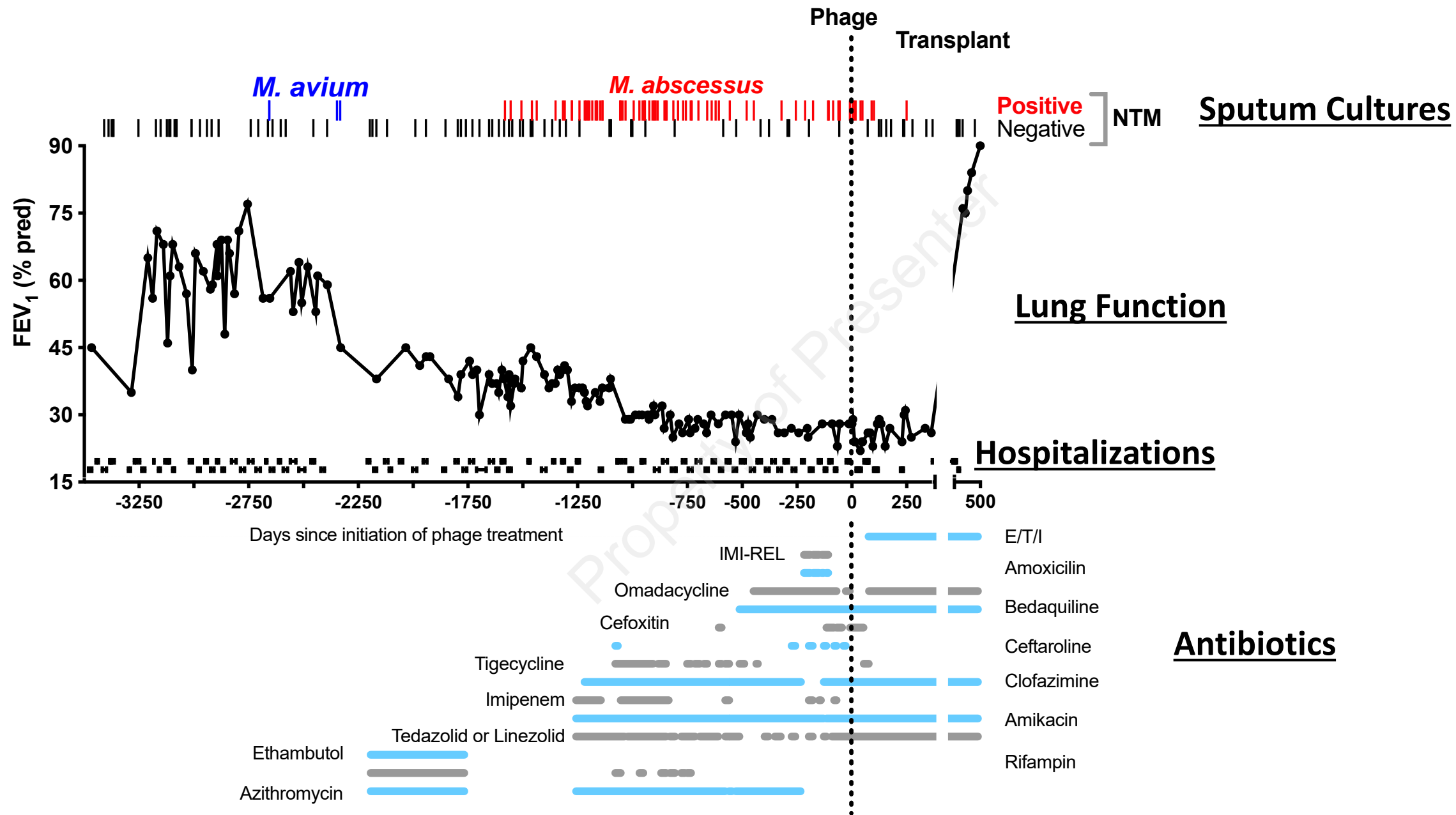
V. Lovell, RN

Protocol For Bacteriophage Treatment of *M. abscessus*

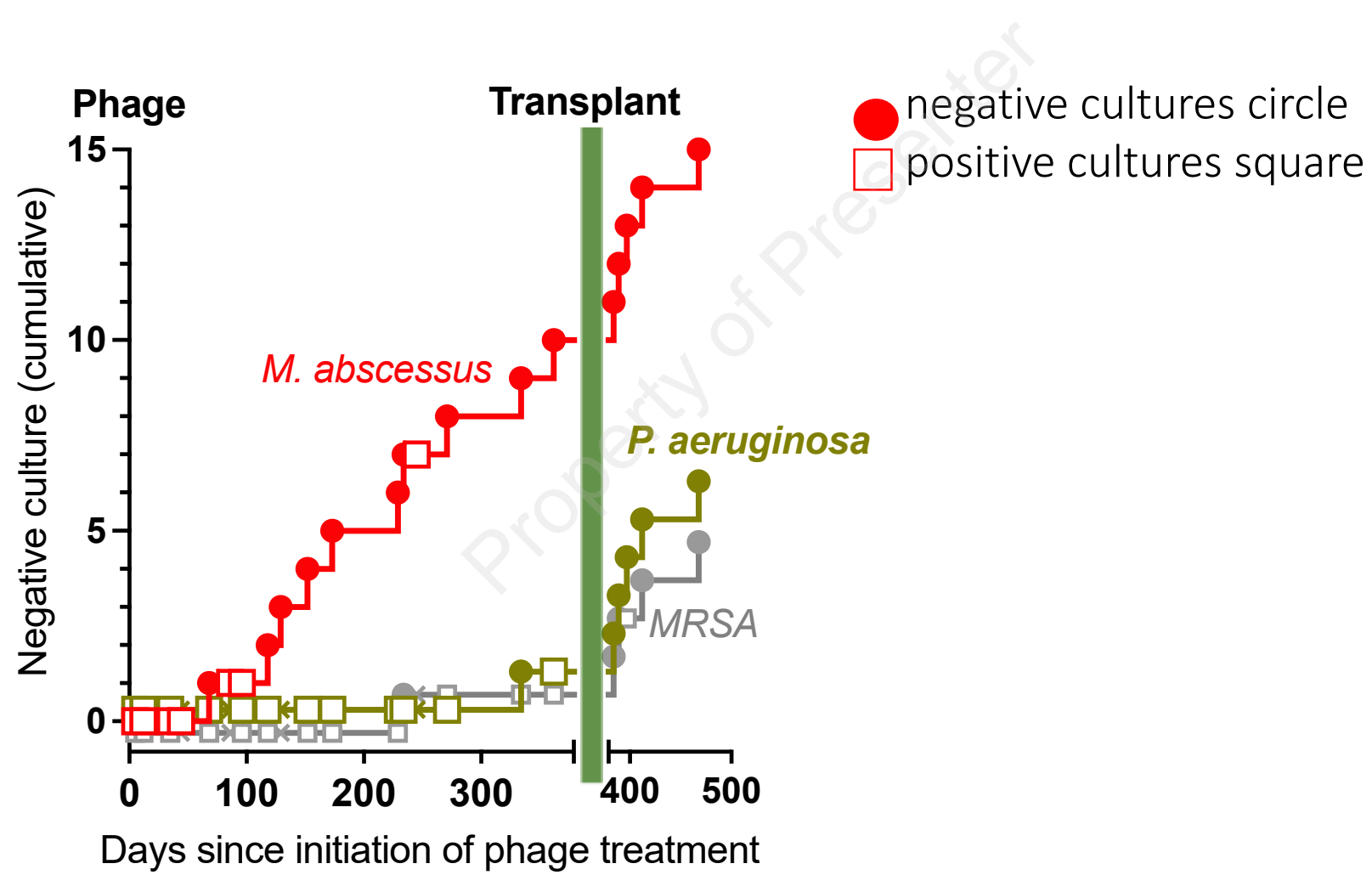
- Initial doses while monitored (St. Joseph)
- 5 ml (10^8 to 10^9 PFU) solution of bacteriophage cocktail (BPs Δ 33HTH_HRM10 and D29_HRM_{GD40}) IV via slow push twice daily via mediport.
- Continuation of intensive antibiotic treatment
- Phage administered by patient when home
- Weekly telehealth or clinic, and visiting nurse and labs.
- Initial approval requested for 24 weeks- extended indefinitely
- Monitoring of cultures and radiographs per standard level of care.



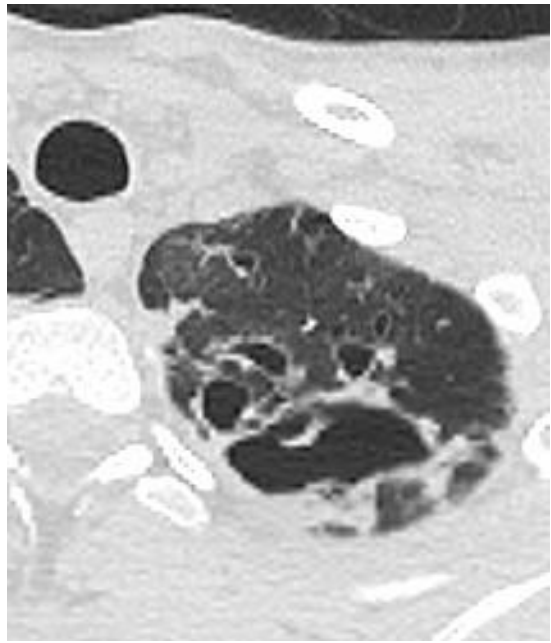
Clinical features



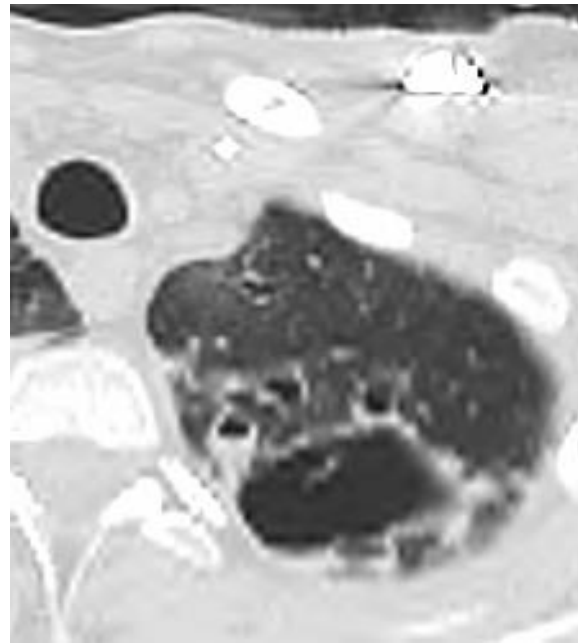
Cumulative negative airway cultures following initiation of phage therapy



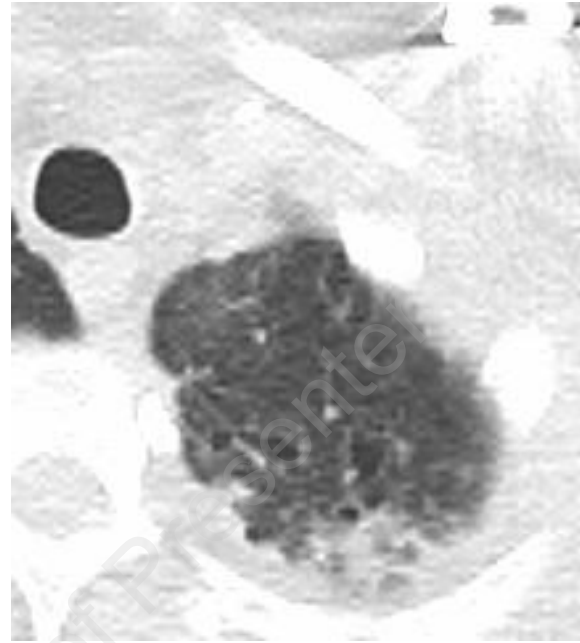
Pre phage day -2



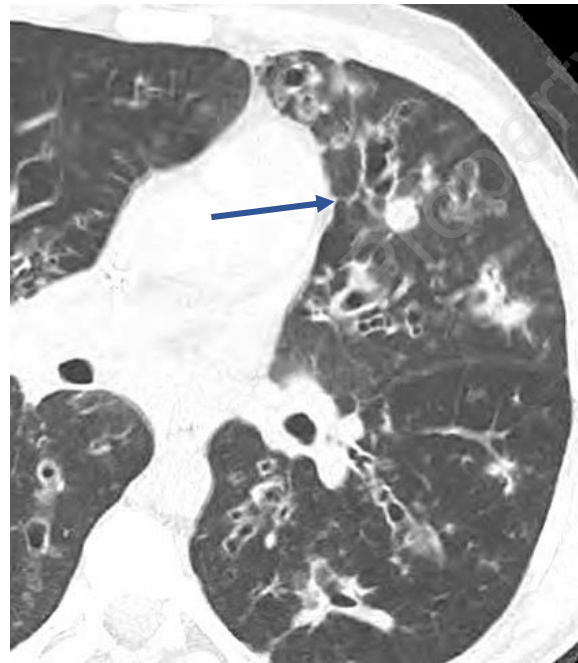
Post phage day 81



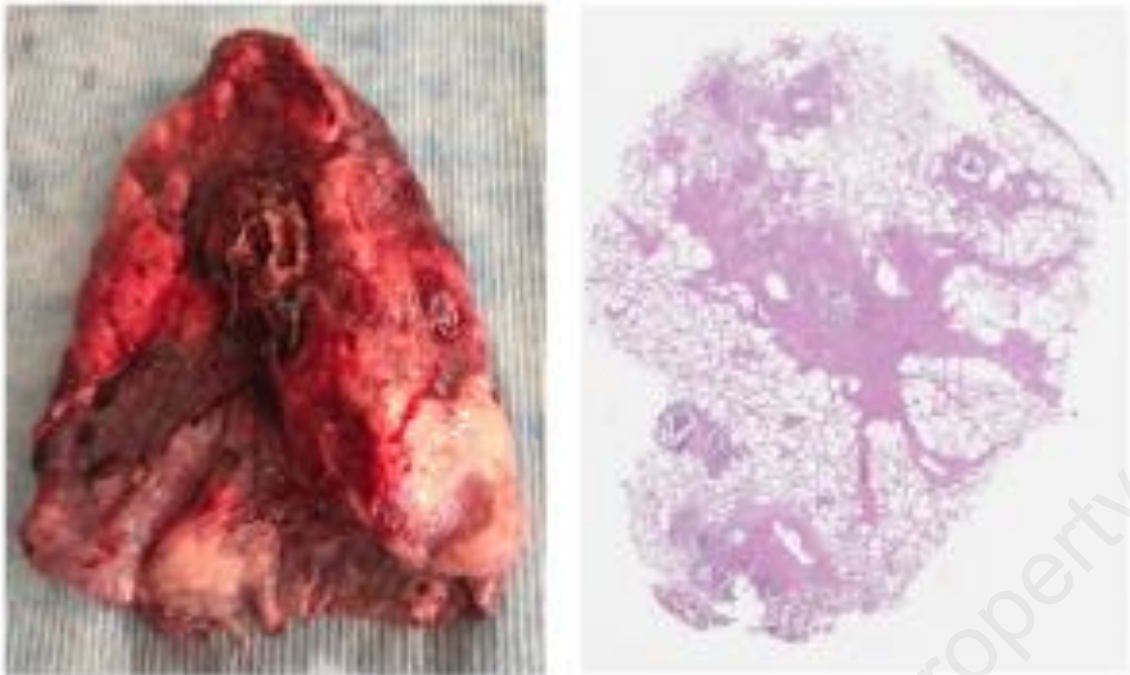
Post phage day 229



Radiologic Changes During Phage Therapy



Systematic analysis of explanted lung tissue for *M. abscessus*



	LUL	LLL	RUL	RML	RLL	RMB	LMB	
Endobronchial Secretions								Bacterial Cx
								AFB stain
								NTM Cx
								MAB qPCR
BAL								AFB stain
								NTM Cx
								MAB qPCR
Tissue Homogenate								AFB stain
								NTM Cx
Tissue								AFB stain

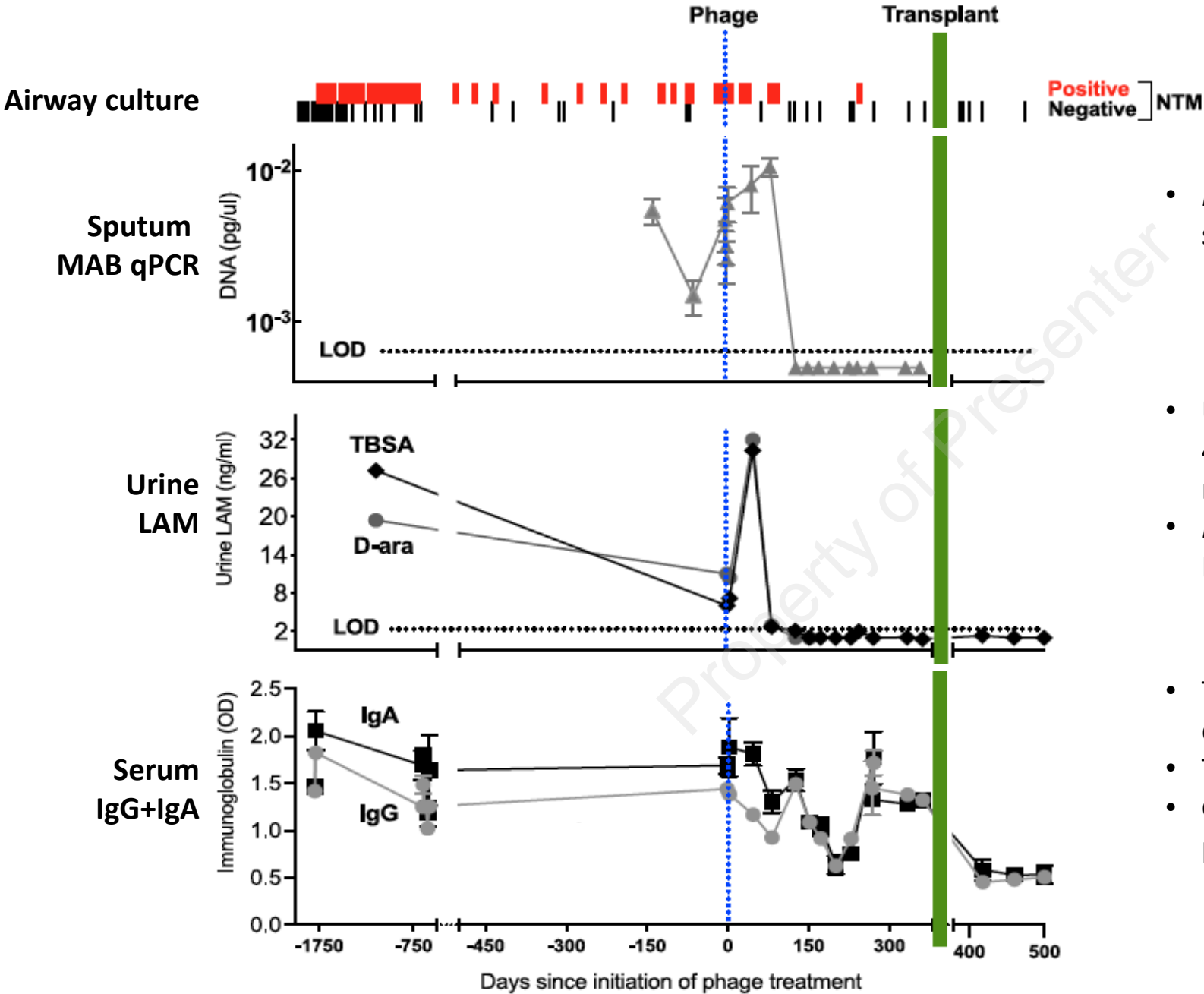
P. aeruginosa
 M. avium
 M. abscessus

M. avium grown from endobronchial secretions.

M. avium last recovered from airway cultures 6.4 years pre-transplant
(1 year of successful treatment)

139 cultures negative for *M. avium*

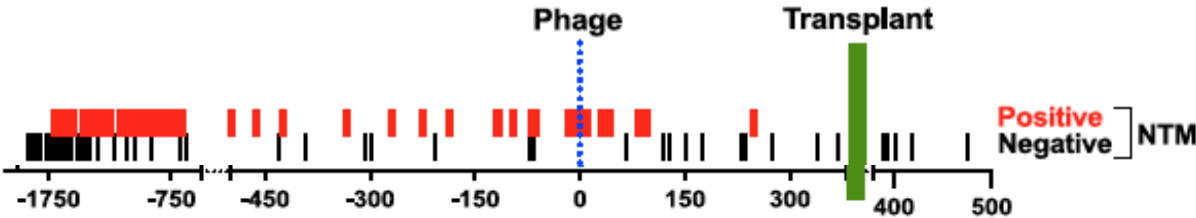
Markers of Treatment Response



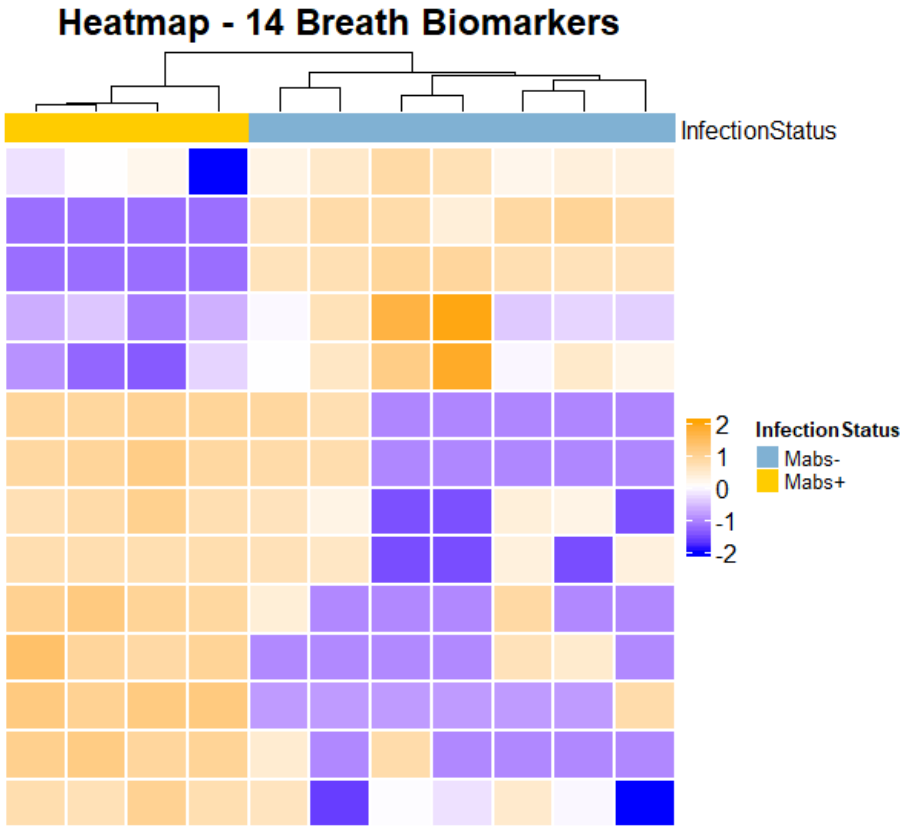
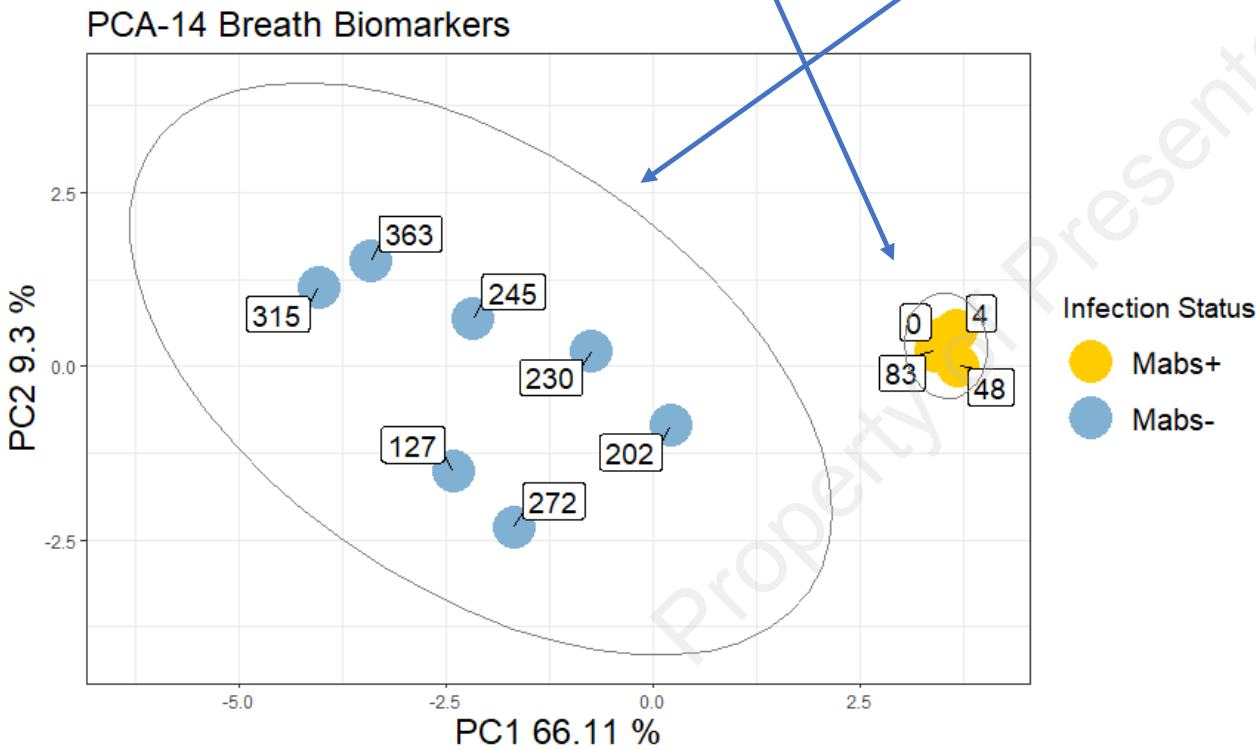
- *M. abscessus*-specific qPCR closely reflected sputum culture results
- Urine LAM signaled dramatic NTM lysis at day 44, and fell below LOD when cultures turned negative.
- *M. avium* infection apparently below urine LAM LOD
- Titers fell rapidly with initial treatment, possibly due to binding of LAM?
- Titers rose at time of late positive culture
- Greatest decrease seen post-transplant, possibly due to elimination of *M. avium*

Markers of Treatment Response?

Airway culture

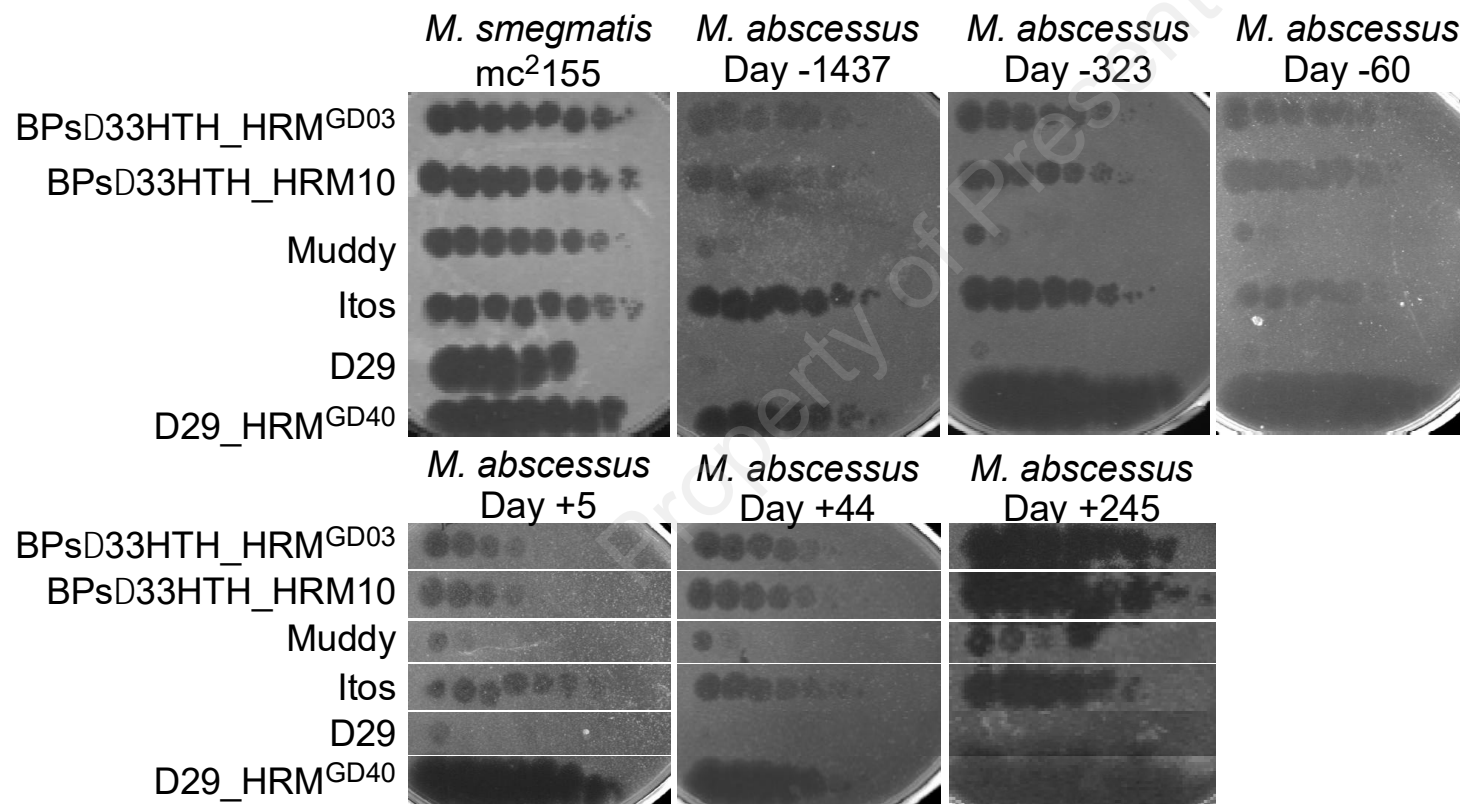


- *Volatile metabolites from breath* closely reflected sputum culture results



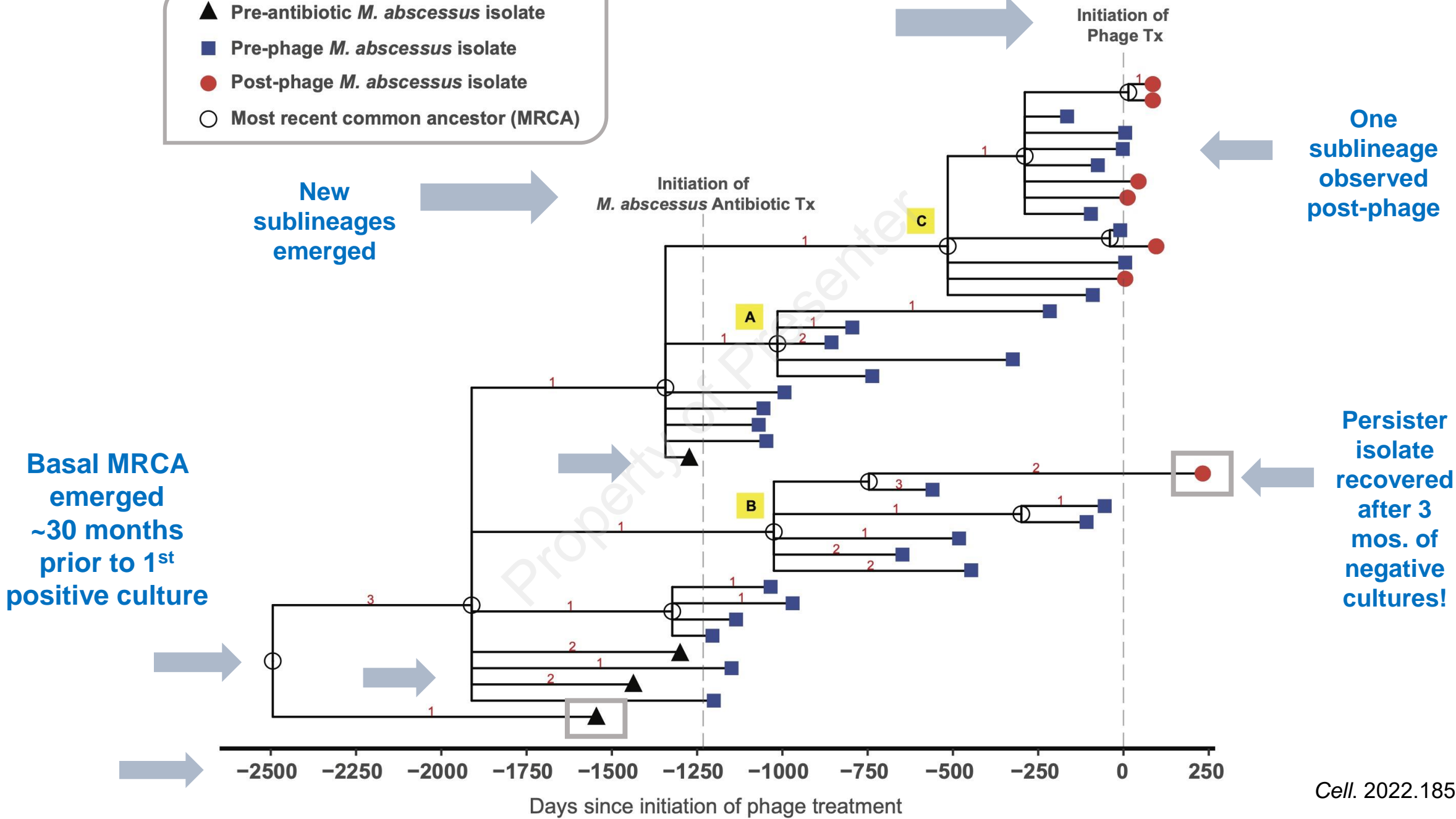
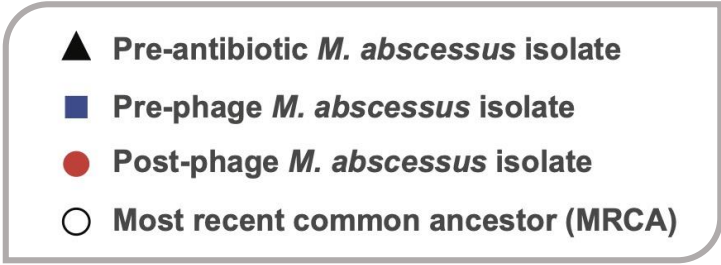
Jane Hill, (UBC)

Phage Therapy Did Not Select for Phage-resistant Isolates

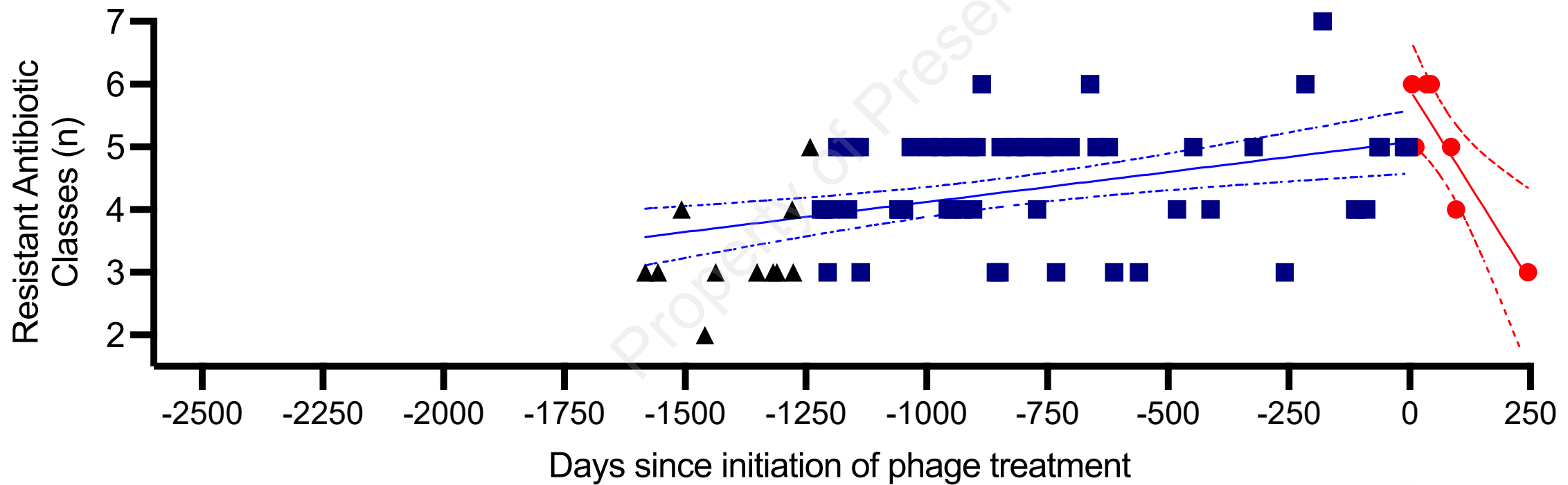


G. Hatfull, PhD

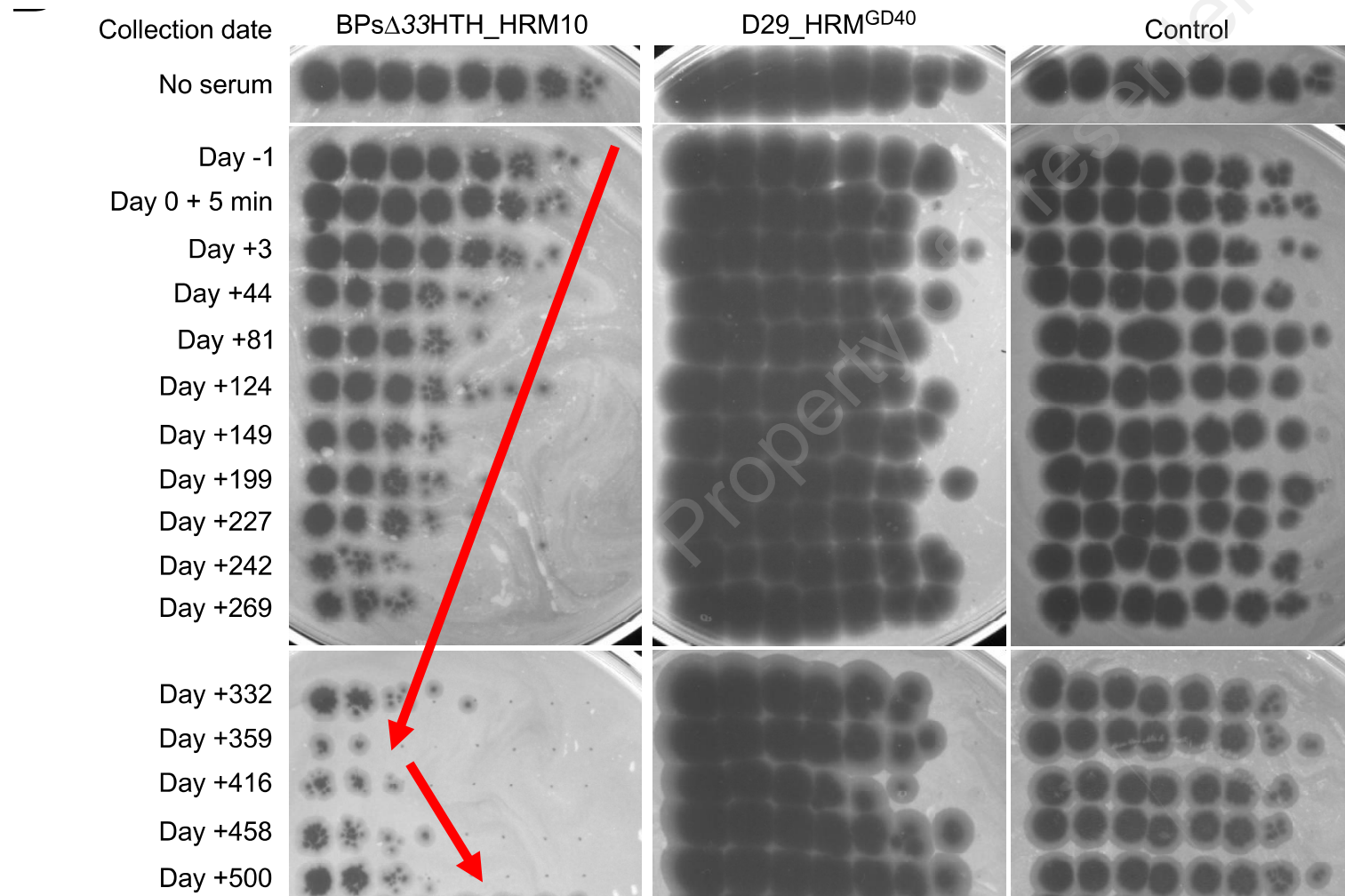
R. Dedrick, PhD



Antibiotic Sensitivity of *M. abscessus* Isolates Following Phage Therapy



Antibody Binding to Phages Pre- and Post- Phage Treatment Demonstrates Late Development of Anti-phage Neutralizing Antibodies.



G. Hatfull, PhD

R. Dedrick, PhD



Conclusions

- People with CF are at high risk for NTM infection
- Unique aspects of CF care has led to a high proportion of individuals with apparently indolent infection, making identification of who will benefit from treatment more challenging.
- Low sensitivity of sputum cultures combined with CFTR-modulator therapy have significantly reduced our ability to screen for NTM or monitor any aspect of disease or treatment.
- Culture-independent markers are urgently needed for clinical care and clinical trials in this population.
- Phage combined with antibiotic treatment resulted in apparent eradication of *M. abscessus*, allowing for a successful lung transplant, without evidence of post-transplant infection.

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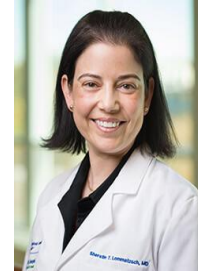
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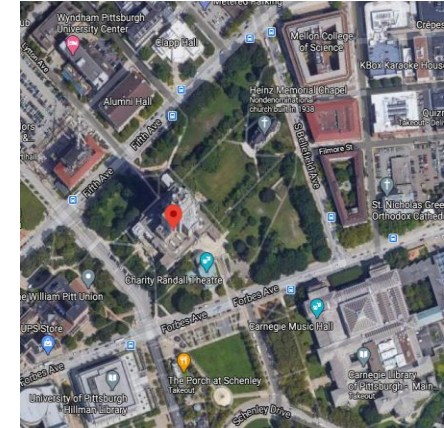
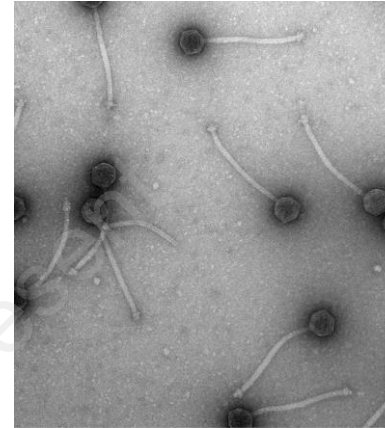
C. Barboa, BS



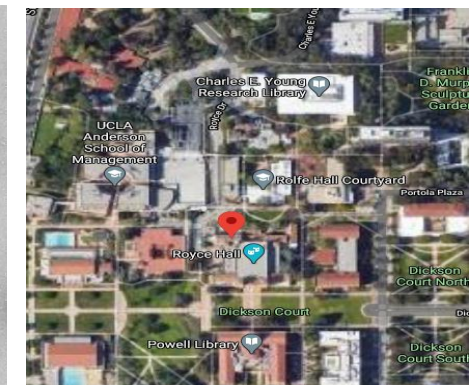
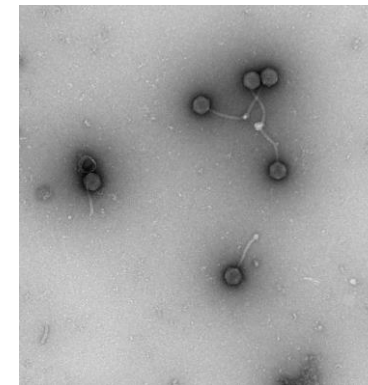
S. Vahling, MEd

Phage engineering

- BPs was isolated using *Mycobacterium smegmatis* as a host.
 - BPs is temperate
 - BPs Δ 33HTH was constructed in which the helix-turn-helix DNA binding domain of the repressor gene (gene 33) is deleted, rendering the phage lytic for *Mycobacterium smegmatis*
 - Mutant BPs Δ 33HTH_HRM10 was isolated and is able to efficiently infect specific *M. abscessus* strains
 - Not considered a GMO by the European Union
-
- D29 is a lytic phage, was isolated using *Mycobacterium smegmatis* as a host
 - Native truncated, non-functional, repressor gene which does not allow for temperate lifestyle



Pittsburgh, Pennsylvania



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