

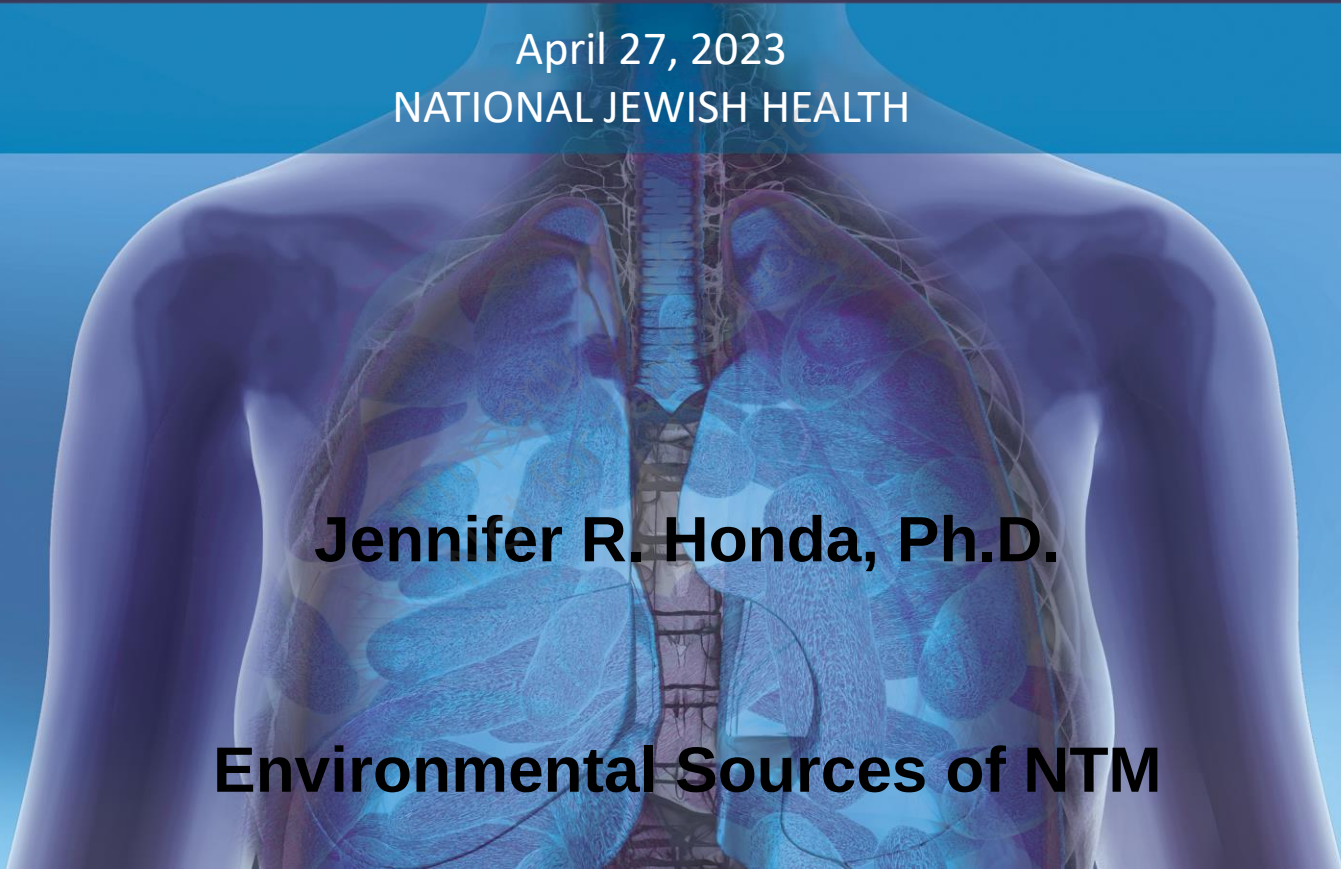
NTM Lecture Series for Providers

April 27, 2023

NATIONAL JEWISH HEALTH

Jennifer R. Honda, Ph.D.

Environmental Sources of NTM



No disclosures.

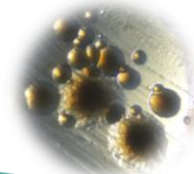
Property of Presenter
Not for reproduction

Honda Lab Research Program

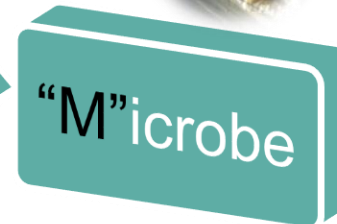


i.e., geography, water associated surface biofilms, soil, dust, animal reservoirs, climate, humidity, temperature, ash, etc.....

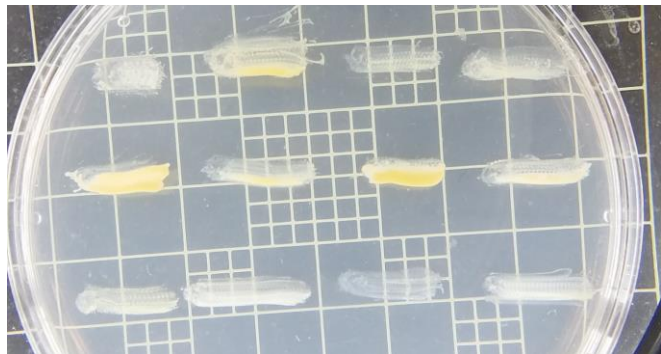
i.e., immune responses of macrophages and airway epithelial cells (humans, mice) to NTM infection



i.e., species of NTM matters, biofilm forming capacity, interaction with host and environment, pigment, virulence factors, etc...



What are *mycobacteria*?



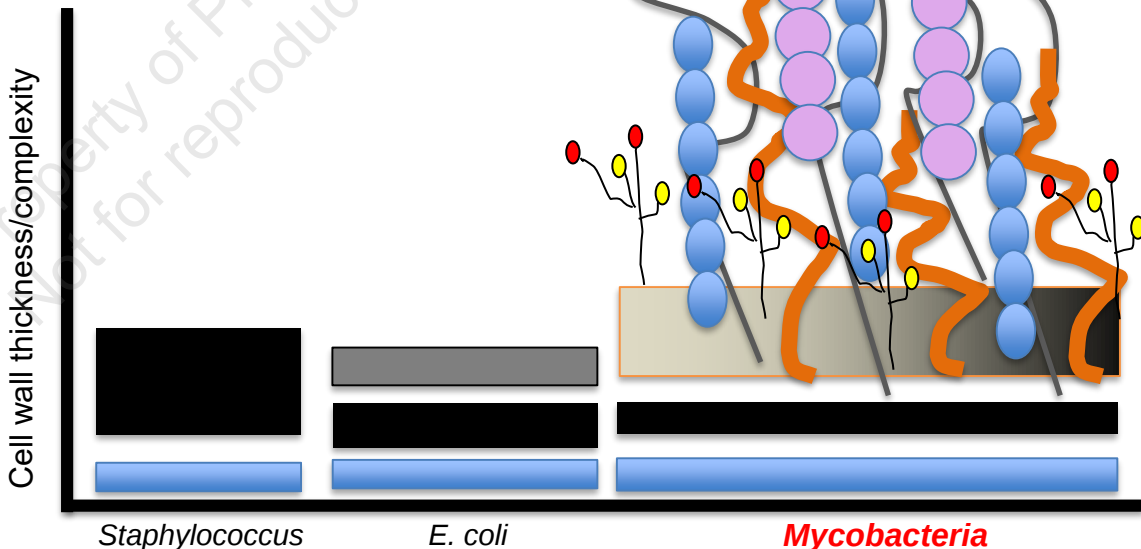
- > 190 different species

<http://www.bacterio.net/mycobacterium.html>

Acid-fast bacteria

- 1) *Mycobacterium tuberculosis* (TB)
- 2) *Mycobacterium leprae* (leprosy)
- 3) **NTM.**

- Thicker & more complex cell wall compared to other bacteria.



Do all mycobacteria cause lung disease?



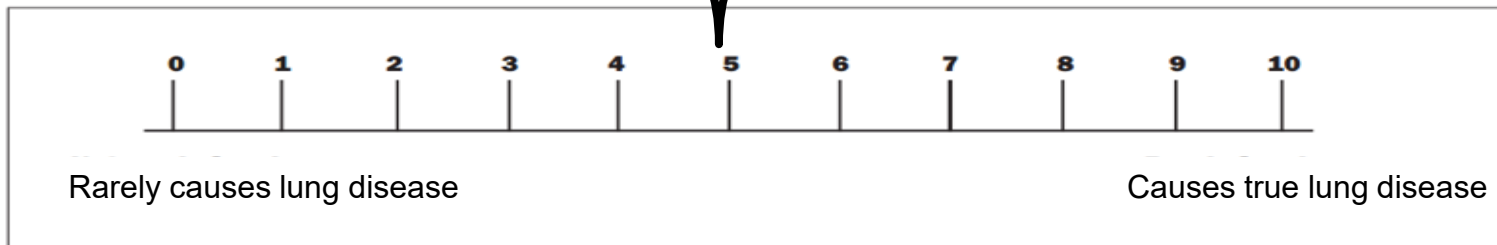
Mycobacterium gordonae
M. gilvum
M. smegmatis



Mycobacterium abscessus
Mycobacterium avium complex (MAC)
 (*M. avium*, *M. intracellulare*, *M. chimaera*)



Mycobacterium tuberculosis (TB)



NTM are one of two “flavors”

“**Rapid**-growing mycobacteria” (RGM)

Visible growth is typically
observed between
3-7 days.

Mycobacterium abscessus (MABS)

- *M. abscessus* subsp. *abscessus*
- *M. abscessus* subsp. *bolletii*
- *M. abscessus* subsp. *massilense*

“**Slow**-growing mycobacteria” (SGM)

Visible growth is typically
observed between
7-60 days.

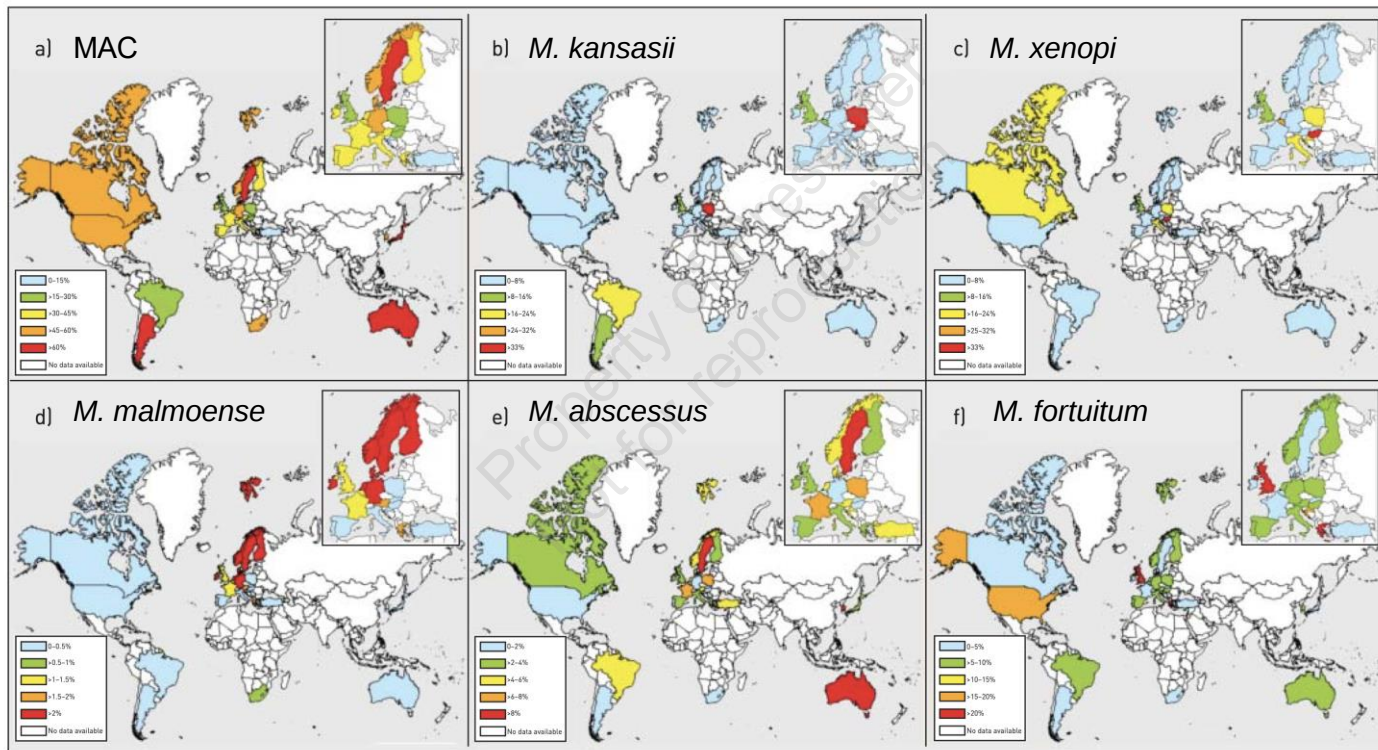
Mycobacterium avium complex (MAC)

- *M. avium*
- *M. intracellulare*
- *M. chimaera*
- others

Learning Objectives

- 1). Become familiar with examples of environmental niches for NTM.
- 2). Recognize features that may promote NTM in the environment – showcasing Hawai'i.
- 3). Identify possible methods to reduce environmental NTM exposures.

Where you live may matter

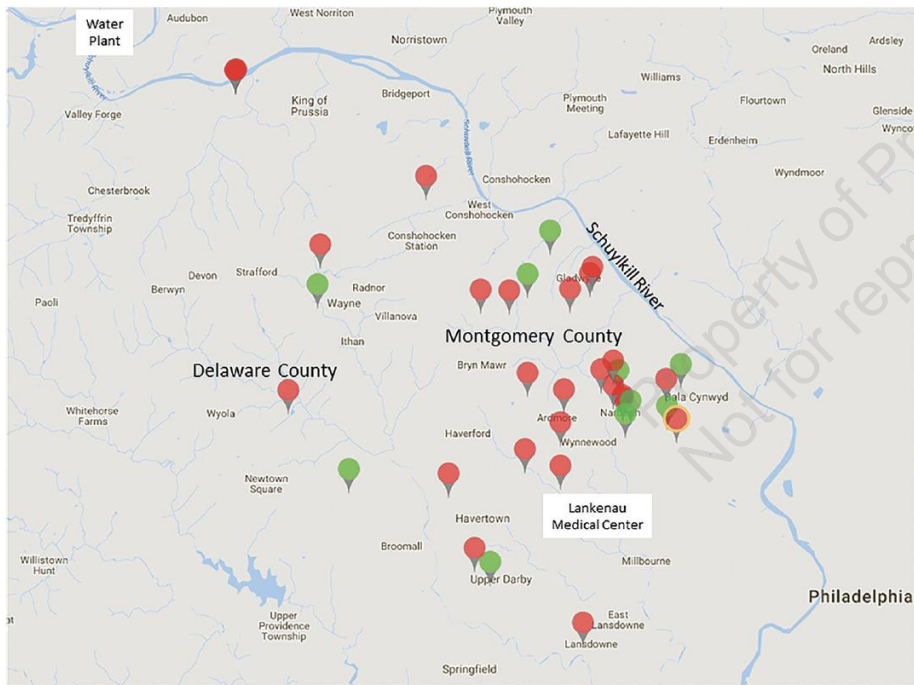


NTM identified from drinking water systems globally

Location:	% NTM recovery:	Species:	Reference:
Mexico	16% (19/120)	<i>M. mucogenicum</i> most common; <i>M. avium</i> , no <i>M. abscessus</i>	Perez, <i>et al.</i> , BMC Res Notes, 2013
Greece	22% (42/197)	<i>M. chelonae</i> most common; no <i>M. abscessus</i>	Tsintzou, <i>et al.</i> , Water, Air, Soil Poll, 2000
USA	33% (46/139)	9 species, <i>M. mucogenicum</i> most common; no <i>M. abscessus</i>	Covert, <i>et al.</i> , AEM 1999
Colombia	50% (9/18)	<i>M. mucogenicum</i> most common; no <i>M. abscessus</i>	Dávalos, <i>et al.</i> , Env Res & Public Health, 2021
Argentina	52% (64/124)	<i>M. gordonae</i> most common	Oriani, <i>et al.</i> , Int J. Mycobacter, 2019
Australia	62% (236/384)	<i>M. gordonae</i> most common; <i>M. abscessus</i> identified	Thomson, <i>et al.</i> , BMC Microb, 2013
Paris, France	72% (104/144)	<i>M. gordonae</i> and <i>M. nonchromogenicum</i> most common; Potentially pathogenic, 16%; no <i>M. abscessus</i> .	Le Dantec, <i>et al.</i> , AEM, 2002

Which species of NTM is found, matters.

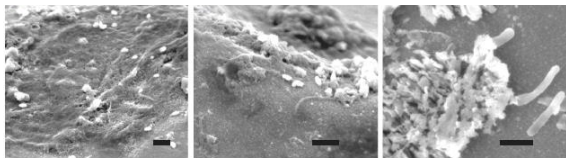
Genetically similar *M. avium* identified from participant lungs and households



Matched Study (Pennsylvania)	
	% NTM recovery:
Recovery from non-participant households	81% (30/37)
Recovery from <i>M. avium</i> participant households	91% (19/21)
Of the <i>M. avium</i> participant households	52% (11/21) matched to environment.

NTM and shower biofilms

Showerhead biofilm



Showerhose biofilm

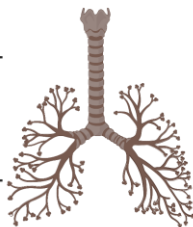


**** MATCH ****

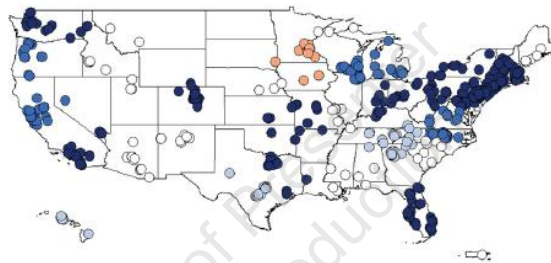
NTM DNA



NTM DNA

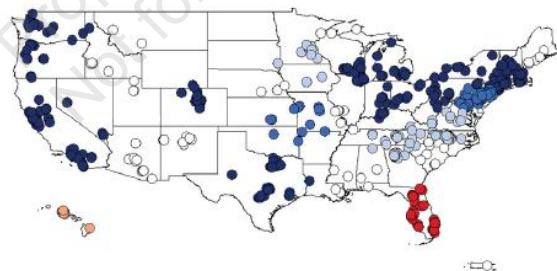


M. fortuitum Complex



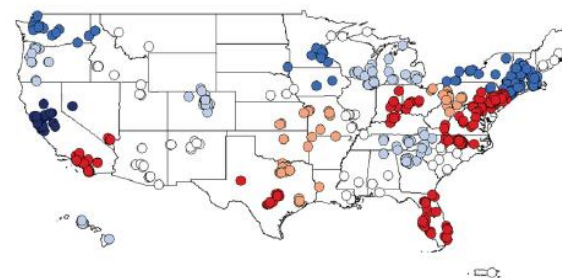
% 0.0-0.02 0.02-0.2 0.2-1.1 1.1-2.3 2.3-9.7

M. abscessus Complex



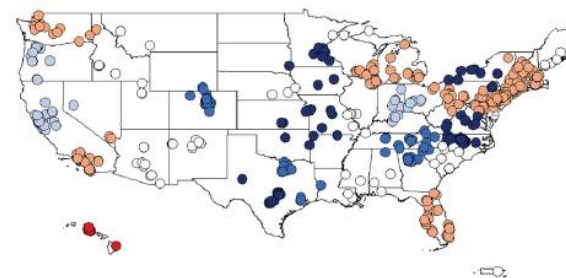
% 0.0-0.01 0.01-0.06 0.06-1.1 1.1-3.2 3.2-6.6

M. mucogenicum/phocaicum



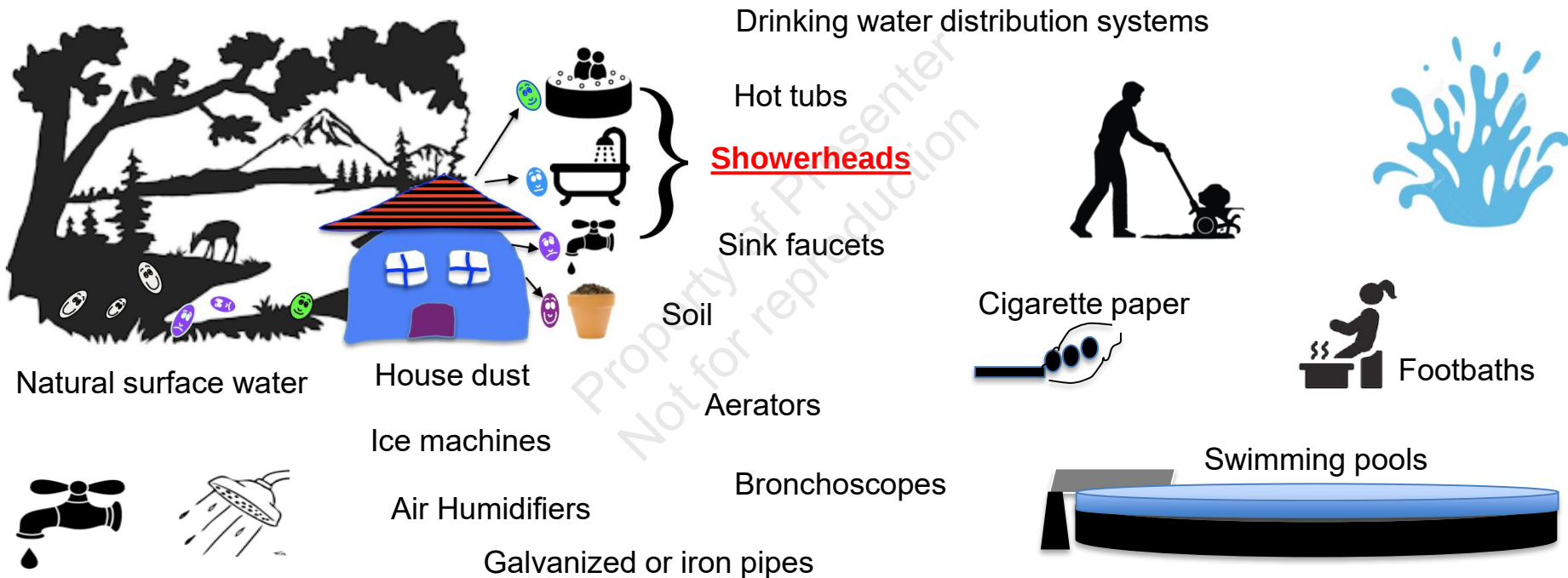
% 0.3-2.2 2.2-5.5 5.5-11.1 11.1-15.0 15.0-33.3

M. avium Complex



% 0.0-0.2 0.2-0.8 0.8-2.2 2.2-7.8 7.8-23.0

Other environmental niches for NTM



Chan, Breslawsky, NTMIR, 2019
 Honda, *et al*, PLOS Neg Trop, 2015
 De Groote, *et al*, AEM, 2006

Guimaraes, *et al*, Am J Infect Con, 2016
 Martin, *et al*, Am Rev Resp Dis 1987
 Briancesco, *et al*, Microchem J, 2014

Falkinham, *et al*, J Clin Micro, 1995
 Falkinham, *et al*, Sem Resp Crit Care Med, 2013
 Falkinham, *et al*, J Water Health, 2007

Learning Objectives

- 1). Become familiar with examples of environmental niches for NTM.
- 2). Recognize features that may promote NTM in the environment – showcasing Hawai'i.
- 3). Identify possible methods to reduce environmental NTM exposures.

Certain metals as NTM predictors – location dependent

Oregon

As molybdenum increases, MAC infections increase by 45%.
 ((Molybdenum associated with disease risk in Colorado)).

As vanadium increases, *M. abscessus* infections increase by 41%.

Figure E1

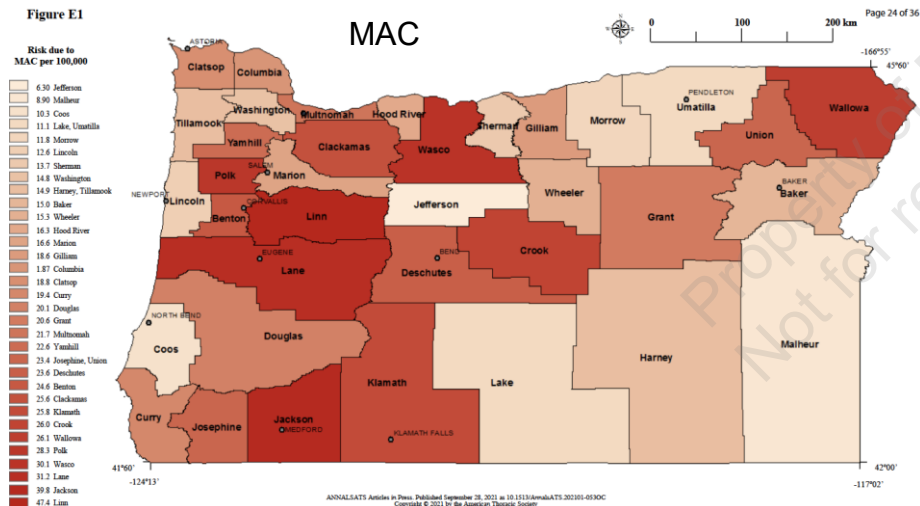
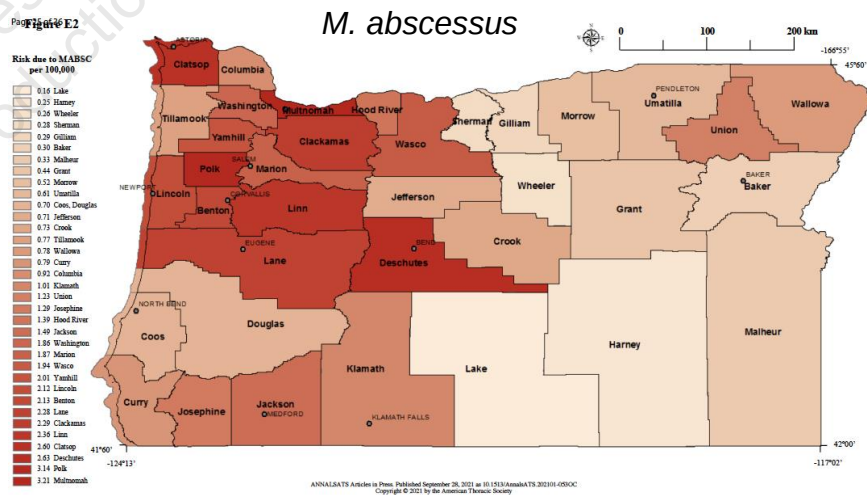
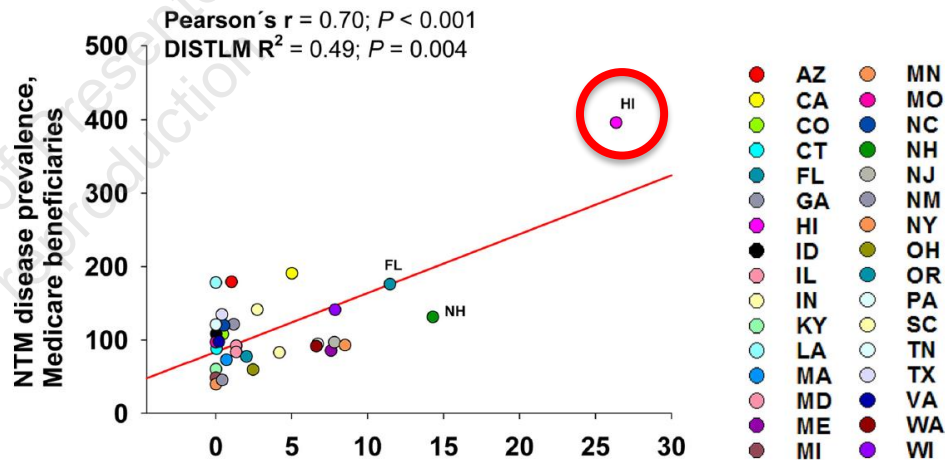
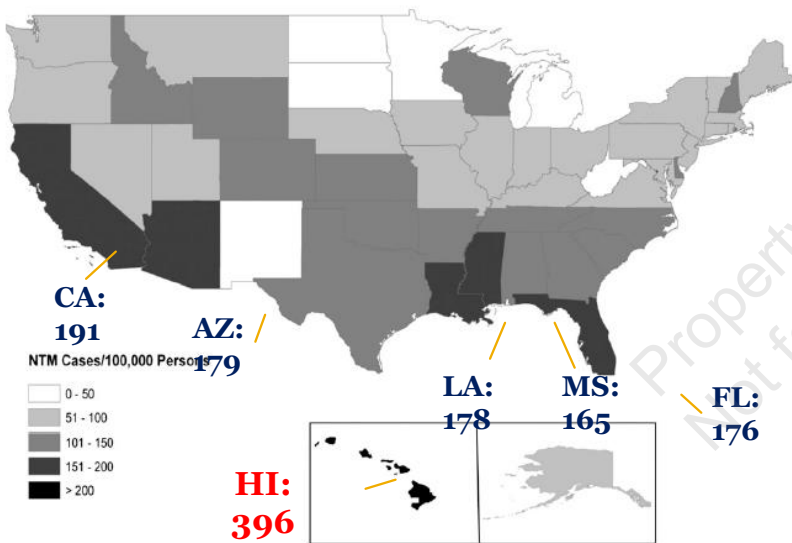


Figure E2



Hawai'i is a hot spot for NTM lung disease

Highest national prevalence - 396 cases/100,000 population among persons > 65 years-old (1997-2007).

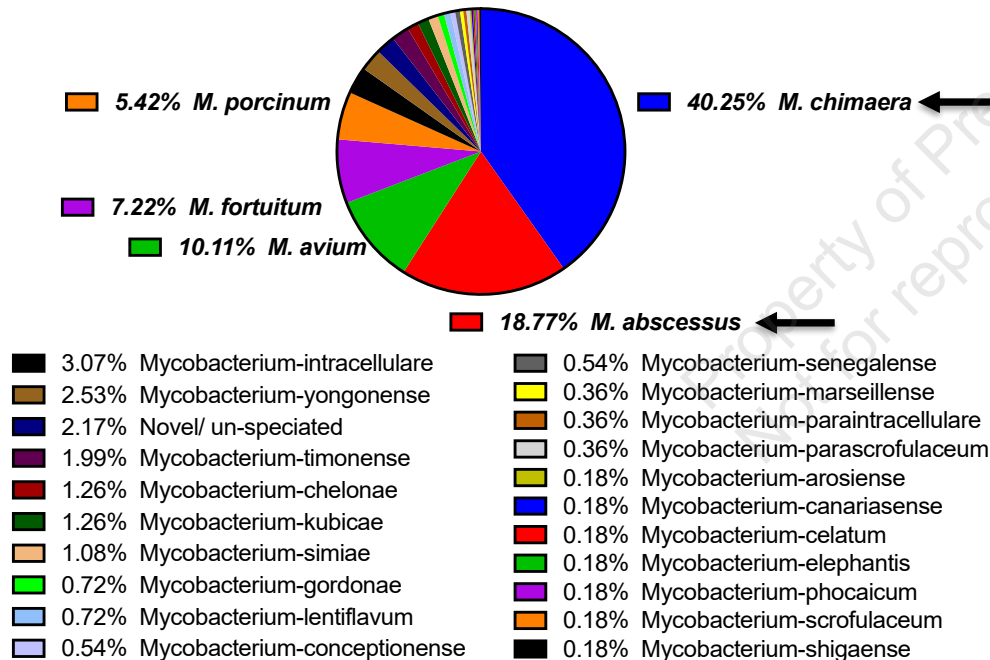


Hawai'i shows the highest, national age-adjusted mortality rates from NTM lung disease.

Respiratory NTM isolate diversity

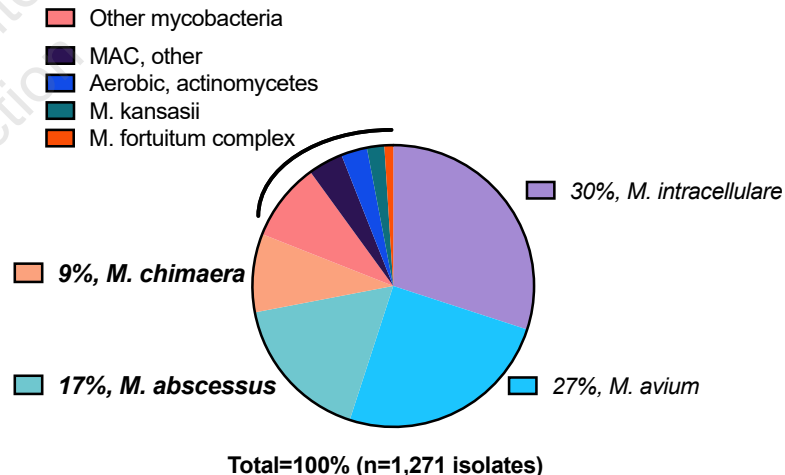
Diagnostic Laboratory Services, Hawai'i

Total: 554 clinical isolates



Unpublished

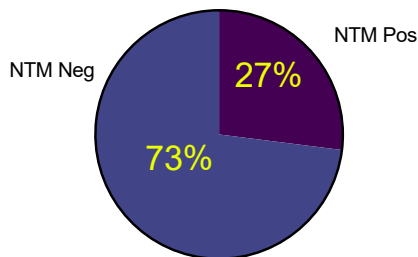
National Jewish Health



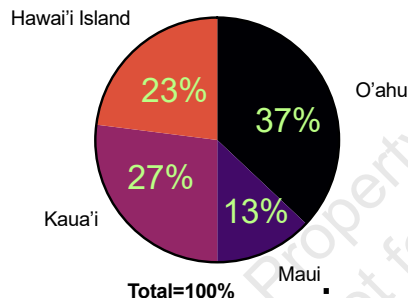
Acknowledgement: Dr. Reeti Khare; Director
NJH Mycobacterial Clinical Labs
Jan-Aug 2021

Large scale environmental NTM sampling, Hawai'i

All state (2,831 samples)



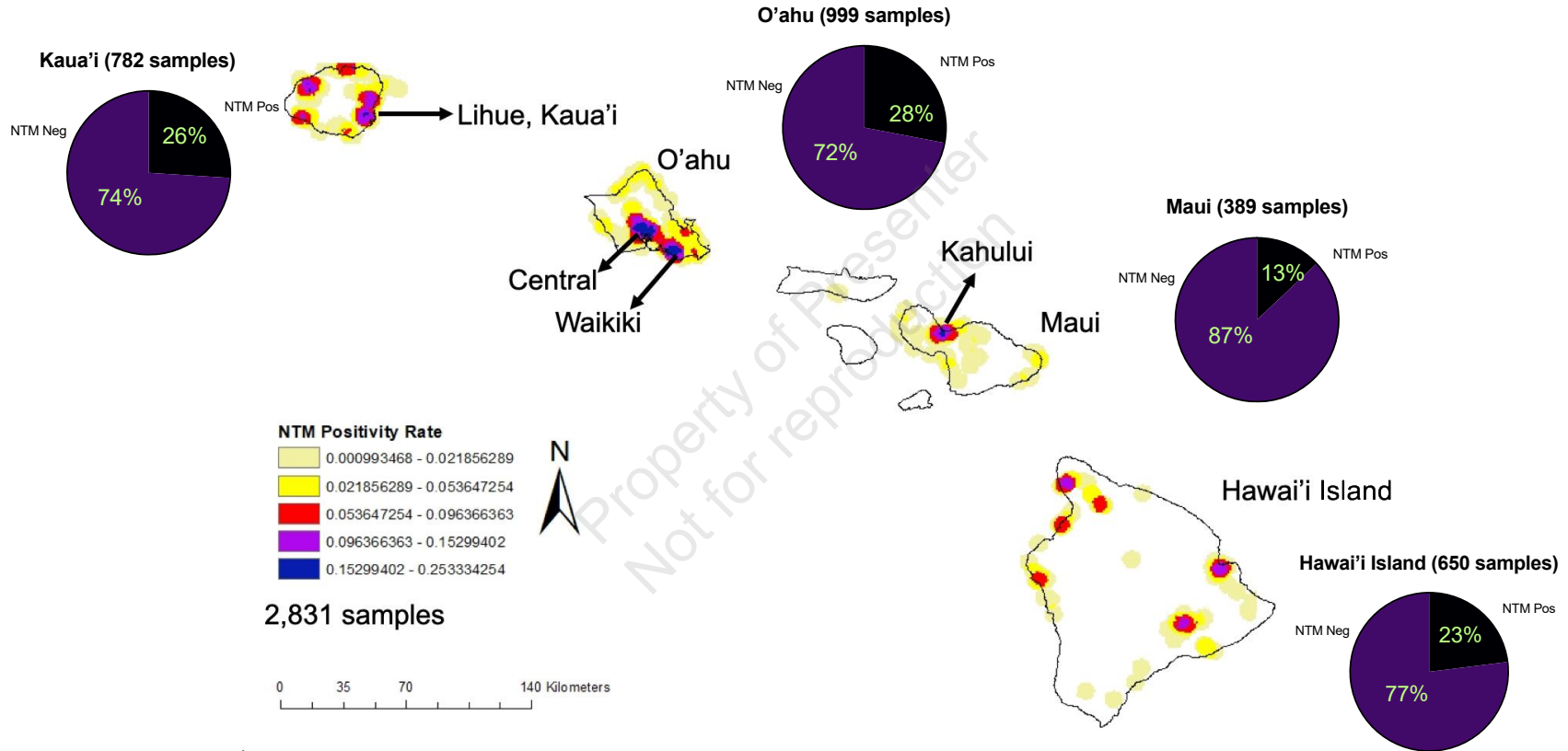
Island distribution
766 total NTM+ samples



Sample Type: NTM positive samples

	Biofilm	Soil	Dust
Entire State	458/1779 (26%)	251/728 (35%)	34/279 (12%)
O'ahu	175/642 (27%)	79/213 (37%)	14/110 (13%)
Hawai'i Island	108/362 (30%)	59/230 (27%)	5/53 (9%)
Maui	50/265 (19%)	38/76 (50%)	9/48 (19%)
Kaua'i	123/499 (25%)	75/209 (35%)	6/68 (9%)

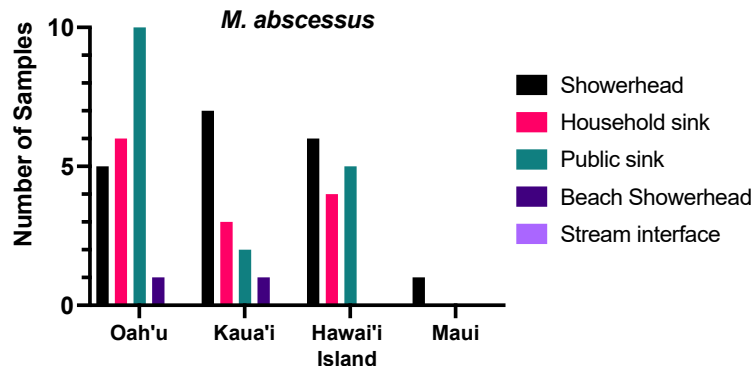
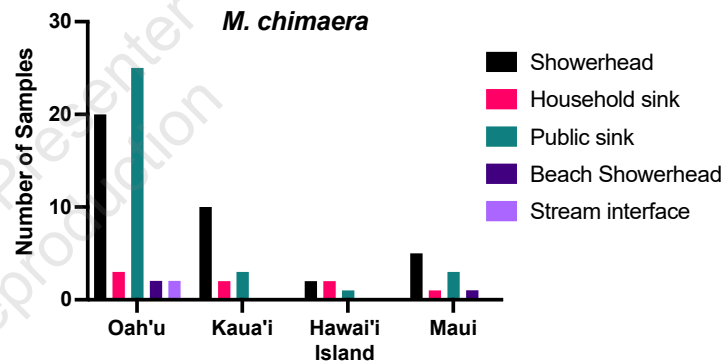
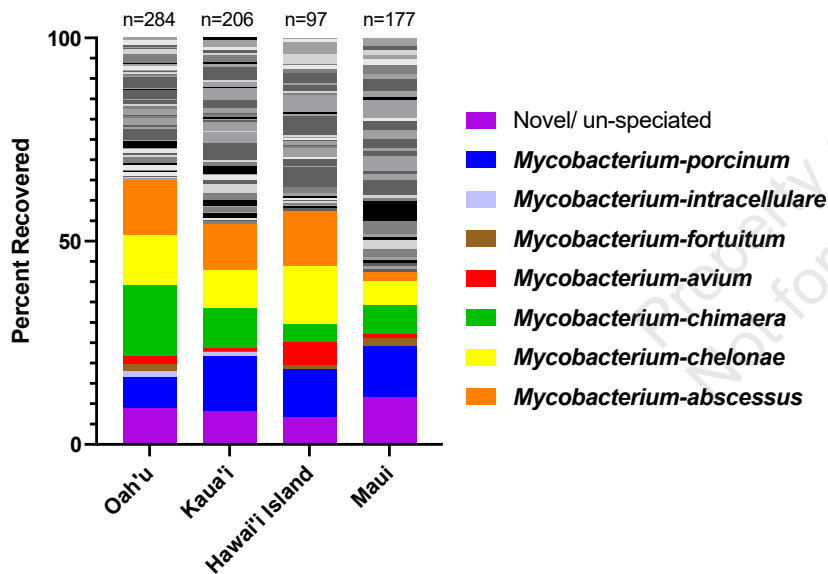
Similar to Falkinham 2011 US samples
28% (109/394)
(biofilm, soil, filters)



Of	8.55%	Novel/ un-specified	0.70%	Mycobacterium-monacense	0.20%	Mycobacterium-senegalense
	3.68%	Mycobacterium-gordonae	0.70%	Mycobacterium-moriokaense	0.20%	Mycobacterium-farcinogenes
	2.68%	Mycobacterium-mucogenicum	0.70%	Mycobacterium-immunogenum	0.20%	Mycobacterium-rutilum
	2.39%	Mycobacterium-avium	0.60%	Mycobacterium-grossiae	0.20%	Mycobacterium-parmense
	2.19%	Mycobacterium-iranicum	0.60%	Mycobacterium-vanbaalenii	0.20%	Mycobacterium-llatzerense
	1.89%	Mycobacterium-gallinarum	0.50%	Mycobacterium-bacteremicum	0.20%	Mycobacterium-goodii
	1.69%	Mycobacterium-paragordonae	0.40%	Mycobacterium-arceuilense	0.20%	Mycobacterium-barrassiae
	1.59%	Mycobacterium-peregrinum	0.40%	Mycobacterium-aurum	0.10%	Mycobacterium-agri
	1.49%	Mycobacterium-neoaurum	0.40%	Mycobacterium-fluoranthenivorans	0.10%	Mycobacterium-simiae
	1.49%	Mycobacterium-phocaicum	0.40%	Mycobacterium-marinum	0.10%	Mycobacterium-longobardum
	1.39%	Mycobacterium-flavescens	0.40%	Mycobacterium-paraffinicum	0.10%	Mycobacterium-mageritense
	1.39%	Mycobacterium-litorale	0.40%	Mycobacterium-lentiflavum	0.10%	Mycobacterium-parascrofulaceum
To	1.29%	Mycobacterium-fortuitum	0.40%	Mycobacterium-chubuense	0.10%	Mycobacterium-psychrotolerans
	1.29%	Mycobacterium-rhodesiae	0.30%	Mycobacterium-poriferae	0.10%	Mycobacterium-scrofulaceum
	1.19%	Mycobacterium-timonense	0.30%	Mycobacterium-canariense	0.10%	Mycobacterium-indicus
	0.99%	Mycobacterium-septicum	0.30%	Mycobacterium-vaccae	0.10%	Mycobacterium-murale
	0.89%	Mycobacterium-florentinum	0.30%	Mycobacterium-genavense	0.10%	Mycobacterium-kansasii
	0.89%	Mycobacterium-triplex	0.30%	Mycobacterium-marseillense	0.10%	Mycobacterium-chitae
	0.89%	Mycobacterium-stomatipteriae	0.20%	Mycobacterium-alvei	0.10%	Mycobacterium-asiaticum
	0.80%	Mycobacterium-conceptionense	0.20%	Mycobacterium-malmesburyense	0.10%	Mycobacterium-tokaiense
	0.80%	Mycobacterium-gilvum	0.20%	Mycobacterium-gadium	0.10%	Mycobacterium-arabiense
	0.80%	Mycobacterium-wolinskyi	0.20%	Mycobacterium-franklinii	0.10%	Mycobacterium-colombiense
	0.70%	Mycobacterium-boenickei	0.20%	Mycobacterium-smegmatis	0.10%	Mycobacterium-insubricum
	0.70%	Mycobacterium-frederiksborgense	0.20%	Mycobacterium-jacuzzi	0.10%	Mycobacterium-sediminis
	0.70%	Mycobacterium-intracellulare	0.20%	Mycobacterium-yongonense	0.10%	Mycobacterium-arupense
			0.20%	Mycobacterium-ratisbonense	0.10%	Mycobacterium-setense
					0.10%	Mycobacterium-madagascariense
					0.10%	Mycobacterium-phlei

t,

Environmental NTM distribution per island



NTM in indoor vs outdoor Hawai'i environments

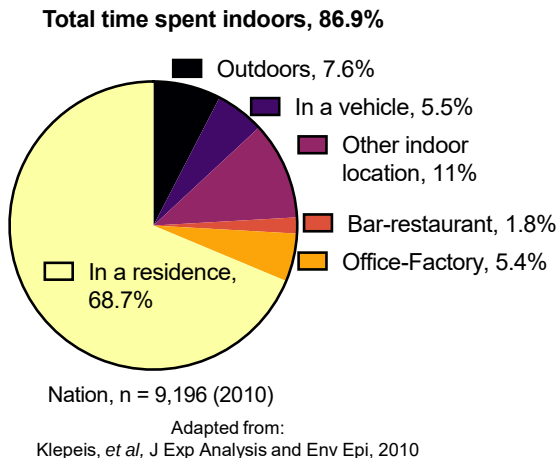
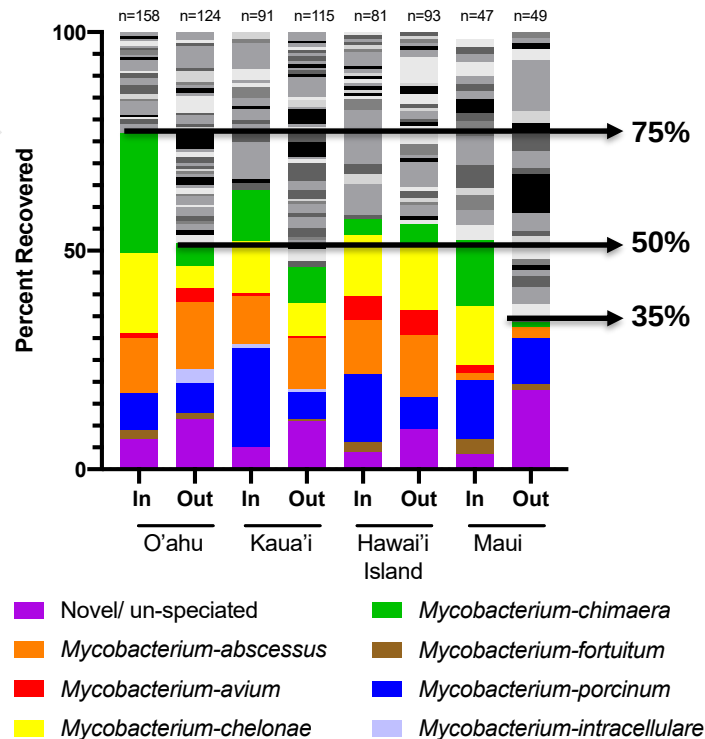


Photo credit: J. Honda



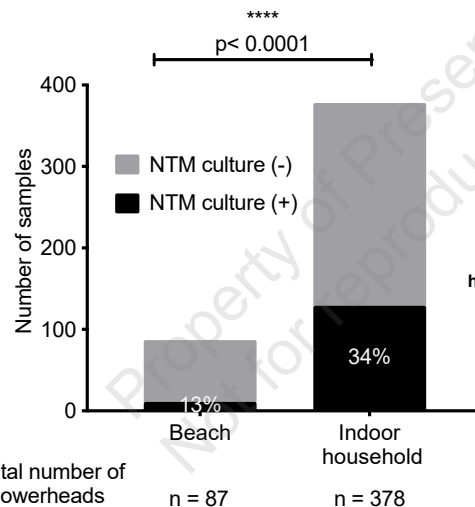
Are all showerhead biofilms created equal?

Indoor
household

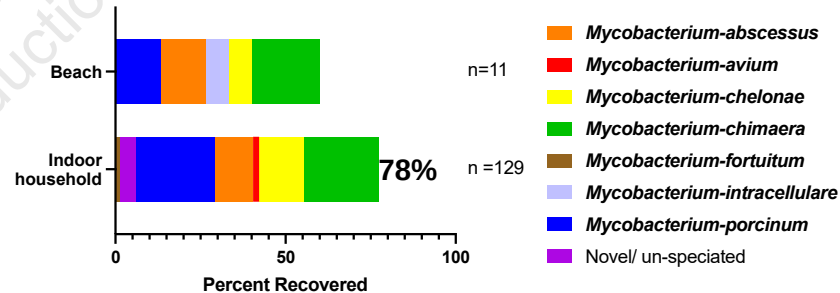
Beach



Photo credit: J. Honda



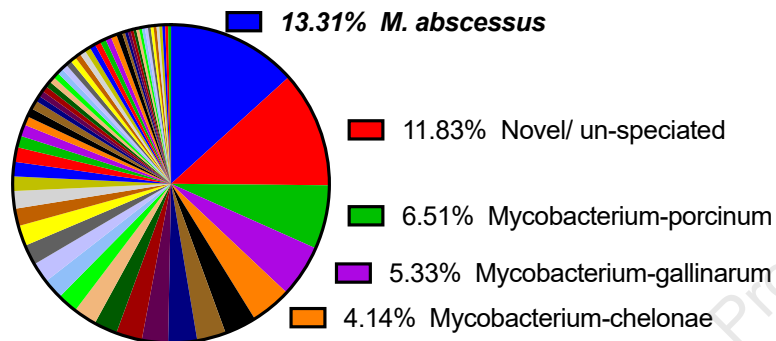
Total number of
showerheads
tested:



Are there unique environmental factors influencing NTM in the Hawai'i environment?

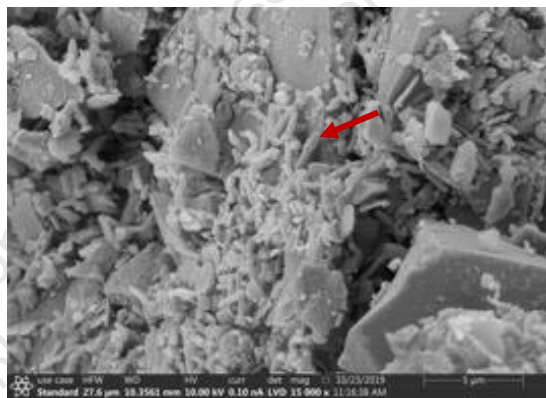
Soil minerals contribute to NTM in Hawai'i

Total: 251 soil samples



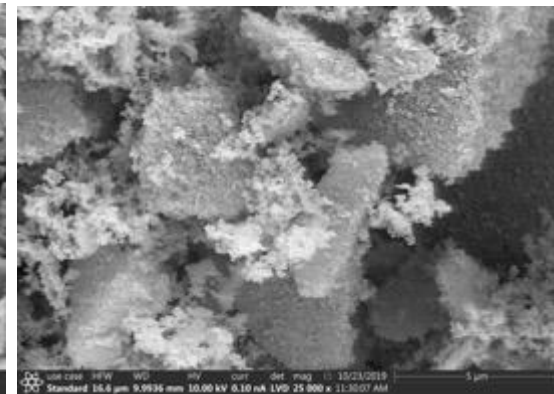
Iron minerals

More *M. abscessus* with **hematite**



Clay minerals

Less with **gibbsite**



NTM probability highest in:

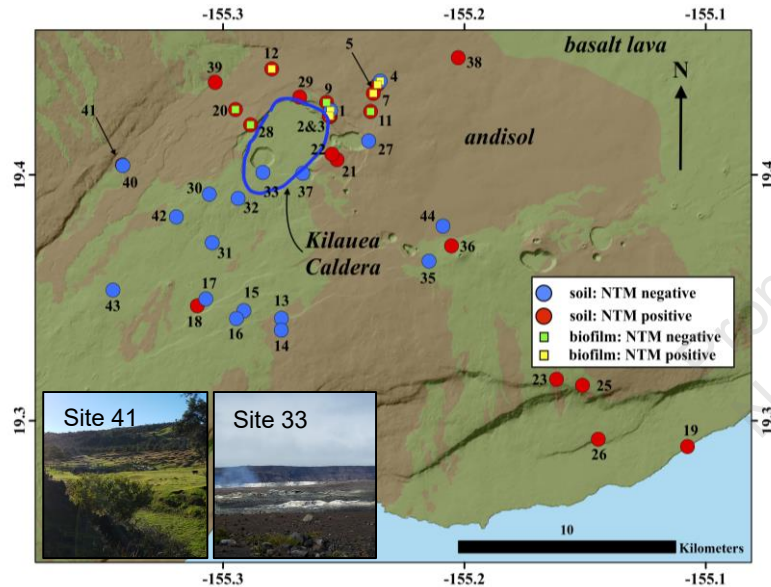
- Rich in ferric oxide.
- Expansive soils with a high-water balance.

Kilauea Volcano, Hawai'i Island

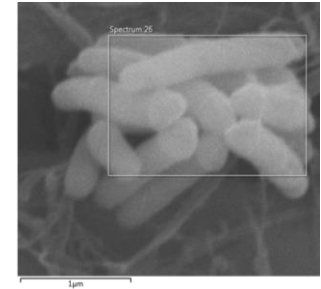
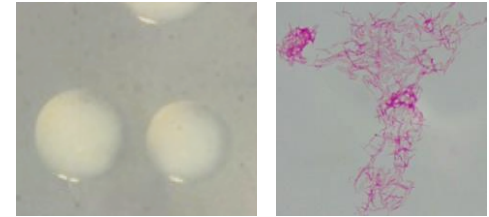


<https://www.usgs.gov/media/before-after/recent-changes-k-lauea-volcano-october-4-2021>
Photo credit: <https://www.usgs.gov/center-news/photo-and-video-chronology-k-lauea-may-29-2018>

Do NTM inhabit volcanic environments?



Kilauea steam vent

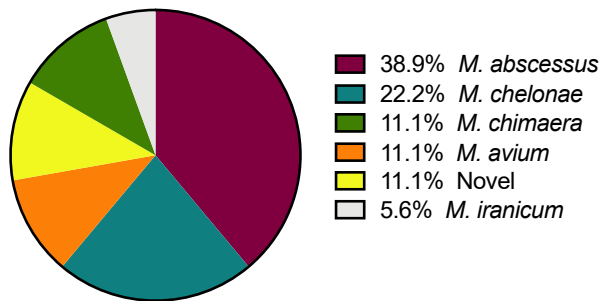


Taxa ID	BLAST ID
<i>Mycobacterium abscessus</i>	99.7%

Diversity of NTM in Kilauea water biofilms and soil

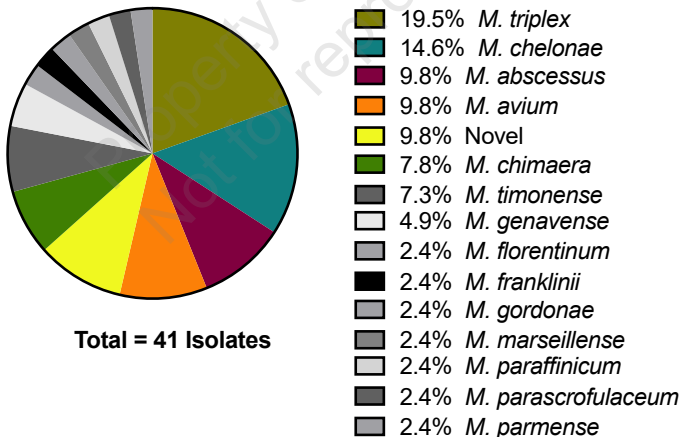
NTM at Kilauea		
Total sites sampled:	No. NTM positive sites:	% NTM Positive:
44	26	59%

Water biofilms

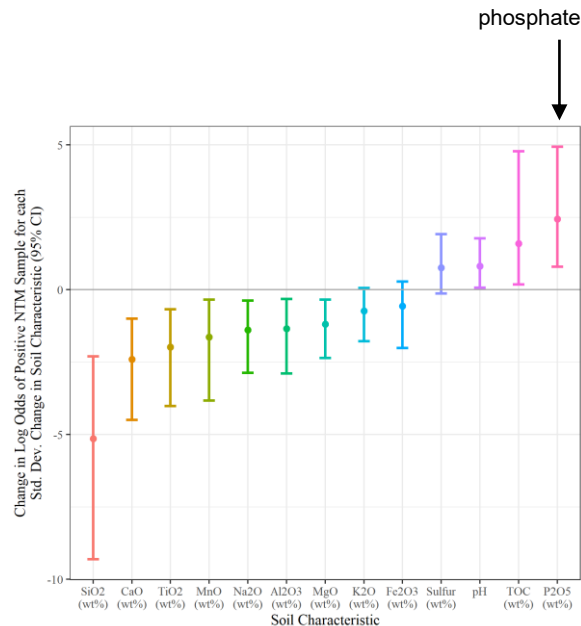


Total = 18 Isolates

Soil



Total = 41 Isolates



Kilauea's Most Recent Explosive Eruption

Jan. 19, 2018 ¹



May 4, 2018 ²



¹ Photo credit: J. Honda

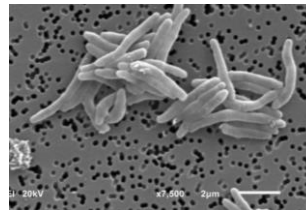
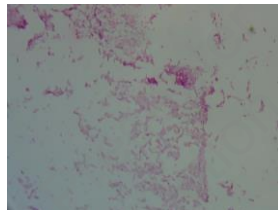
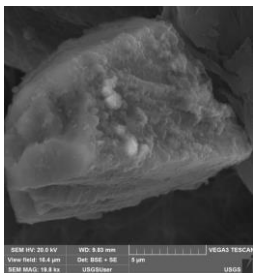
² Photo credit: <https://www.usgs.gov/center-news/photo-and-video-chronology-k-lauea-may-4-2018>

Kilauea Volcanic Ash Harbors Viable NTM



David Damby, PhD

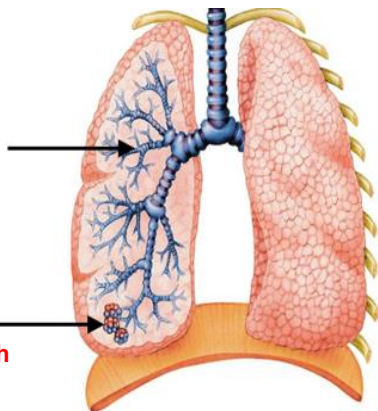
Ash SEM



<i>rpoB</i> Taxa ID	BLAST ID
<i>Mycobacterium abscessus</i>	99.2%

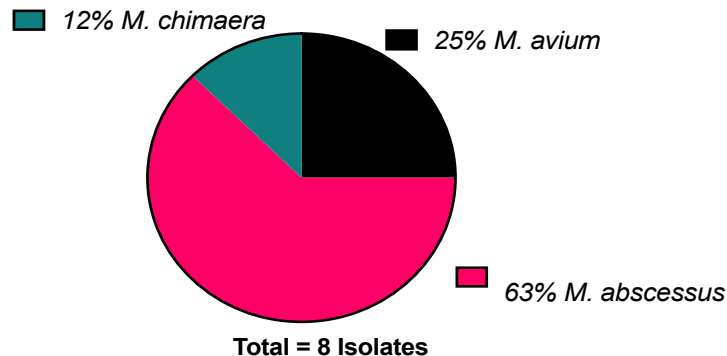
< 32 μm
"Fine" ash

< 5 μm
"Respirable" ash
can reach the
alveoli



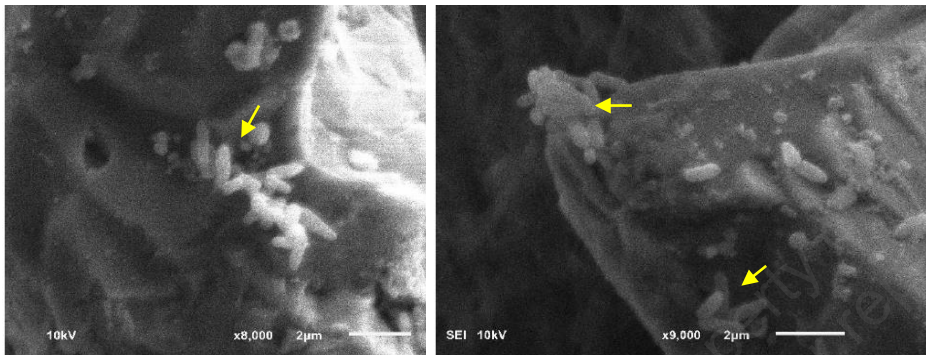
8 NTM-like colonies
were isolated

Kilauea ash, May 2018

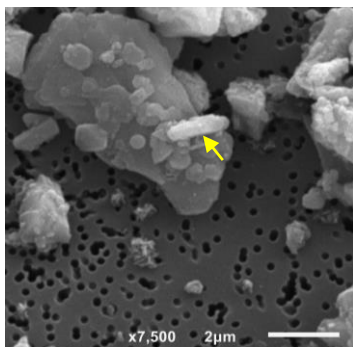


NTM adheres to Kilauea ash

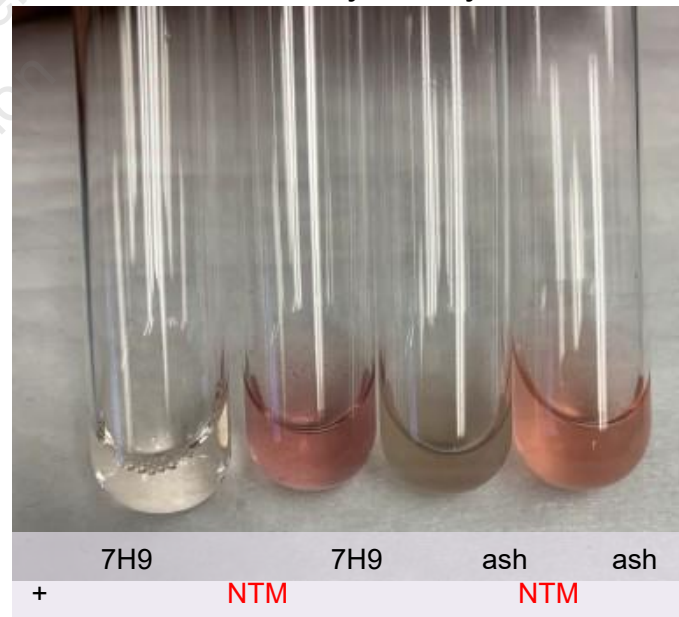
M. abscessus + ash



M. chelonae + ash



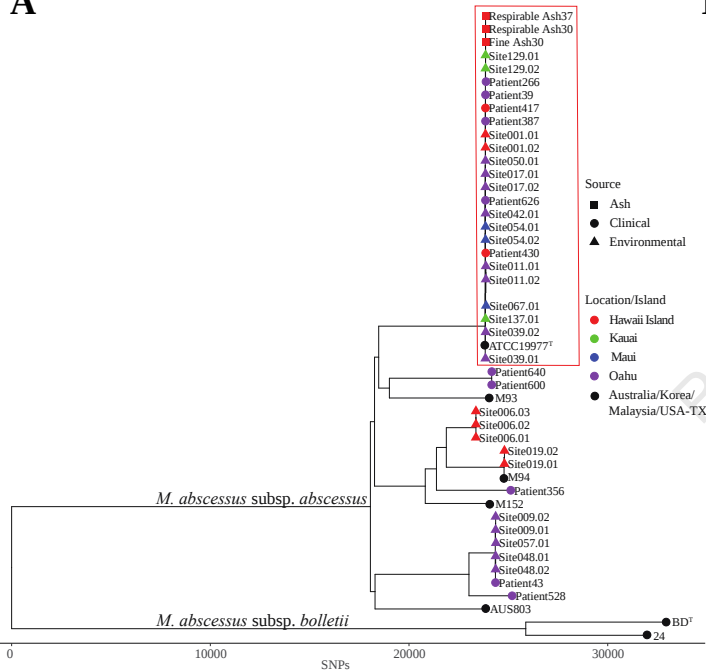
Viability Assay



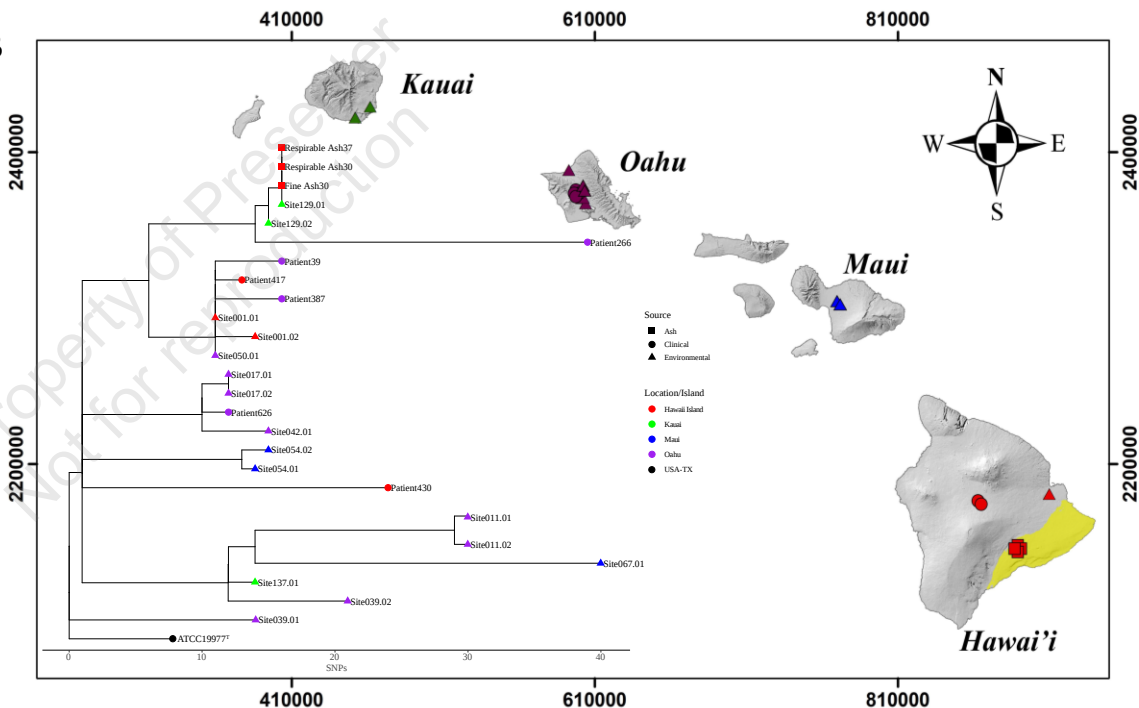
NTM are viable in the presence of ash.

Kilauea ash as a possible fomite for NTM transmission

A

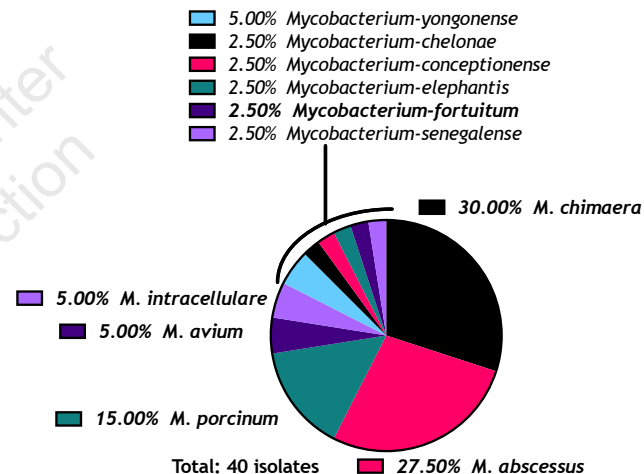


B



NTM in Hawai'i – A Matched Study

	Cases (n=35)	Controls (n=28)	p-value
Gender			
Female	18 (51.4%)	15 (53.6%)	1
Male	17 (48.6%)	13 (46.4%)	1
Age			
40-50	21 (60.0%)	14 (50.0%)	0.59
51-60	2 (5.7%)	1 (3.6%)	1
61-70	4 (11.4%)	4 (14.3%)	1
71-80	7 (20%)	7 (25%)	0.89
81-90	1 (2.9%)	2 (7.1%)	0.58
Race/Ethnicity			
Asian	13 (37.1%)	4 (14.3%)	0.08
Native Hawaiian and Pacific Islander	2 (5.7%)	2 (7.1%)	1
White	15 (42.9%)	12 (42.9%)	1
Other	5 (14.3%)	10 (35.7%)	0.09



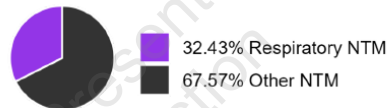
	Kitchen Faucet 	Showerhead 	Soil 
<i>M. abscessus</i>	5/5 (100%)	0/7 (0%)	N/A
<i>M. chimaera</i>	0/7 (0%)	4/4 (100%)	0/2 (0%)

Possible significance of NTM in Hawai'i

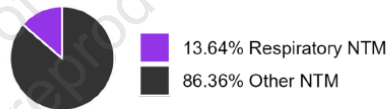
Ranking	Top Environmental:	Top Respiratory:
1	<i>M. abscessus</i>	<i>M. chimaera</i>
2	<i>M. chelonae</i>	<i>M. abscessus</i>
3	<i>M. chimaera</i>	<i>M. avium</i>
4	<i>M. porcinum</i>	<i>M. fortuitum</i>
5	Novel/un-specified	<i>M. porcinum</i>
6	<i>M. gordonae</i>	<i>M. intracellulare</i>
7	<i>M. mucogenicum</i>	<i>M. yongonense</i>
8	<i>M. avium</i>	Novel/un-specified
9	<i>M. iranica</i>	<i>M. timonense</i>
10	<i>M. gallinarum</i>	<i>M. chelonae</i>

Natural areas

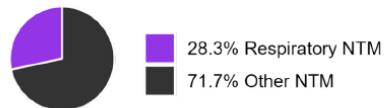
A: Total 37 Water Biofilms



B: Total 22 Water Filters

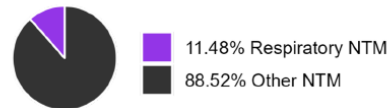


C: Total 212 Soil Samples

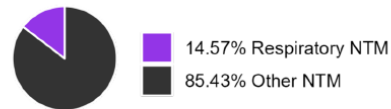


Houses

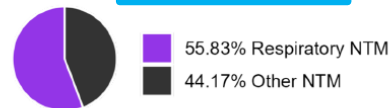
D: Total 61 Dust Samples



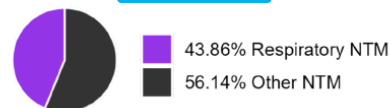
E: Total 151 Soil Samples



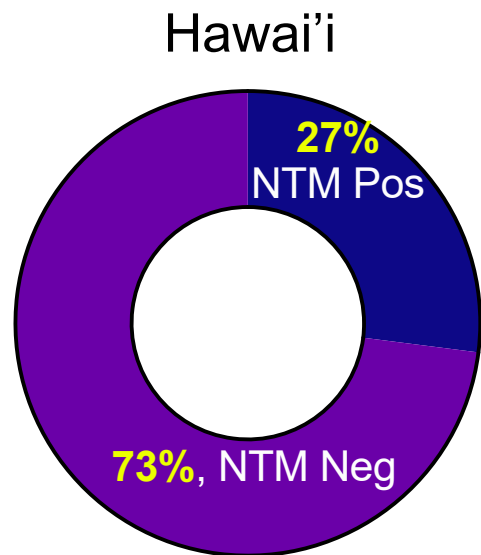
F: Total 120 Showerhead Biofilms



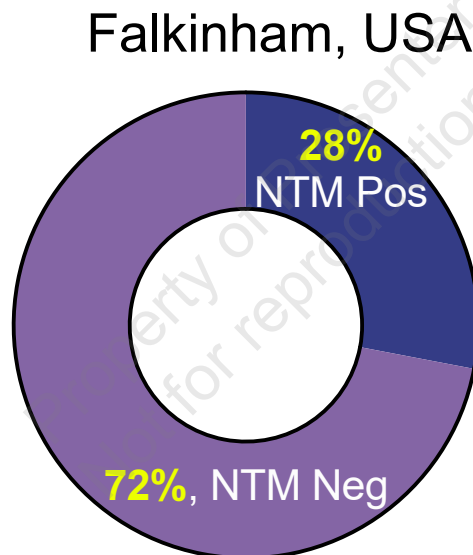
G: Total 114 Sink Biofilms



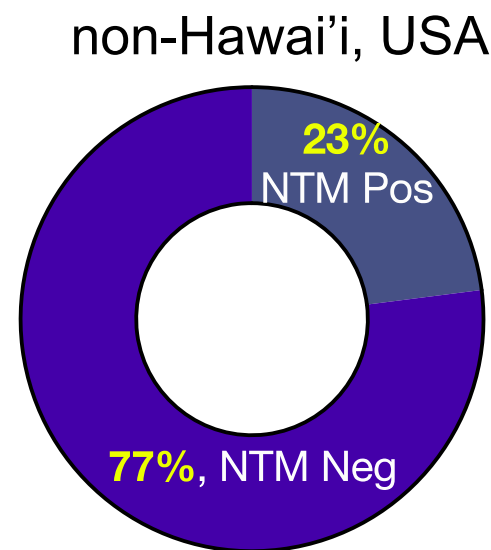
NTM may not be as “ubiquitous” as presumed



n = 2,831 samples



n = 394 samples

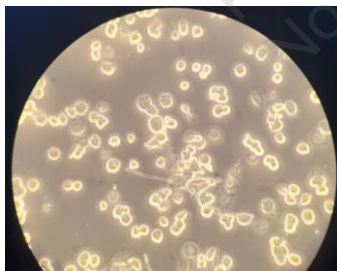
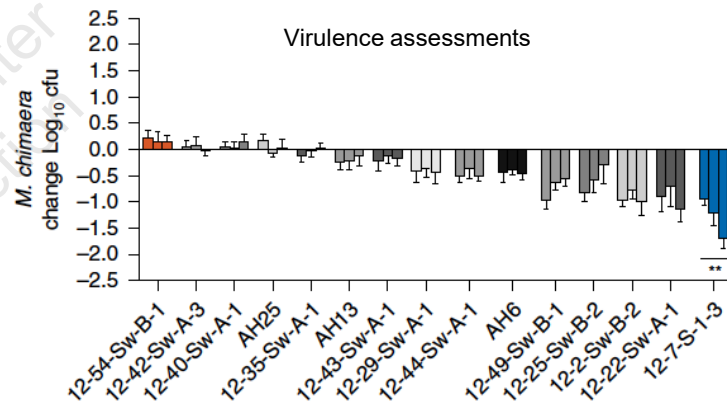
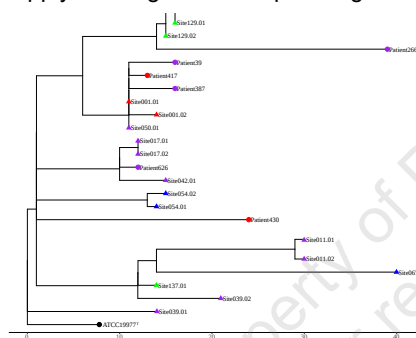


n = 525 samples

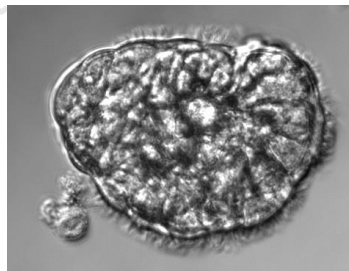
Testing new hypothesis – ongoing work

Biorepository:	
	No. of ENV Isolates:
<i>M. chimaera</i>	154
<i>M. abscessus</i>	182
<i>M. chelonae</i>	153
<i>M. porcinum</i>	183
<i>M. avium</i>	27
<i>M. intracellulare</i>	11
<i>M. fortuitum</i>	20
Other NTM isolates	733
Total:	1,463

Apply whole genome sequencing information



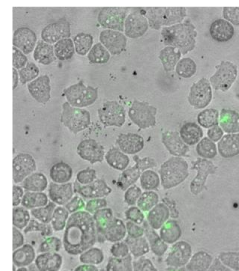
Macrophages



ALI, lung organoids



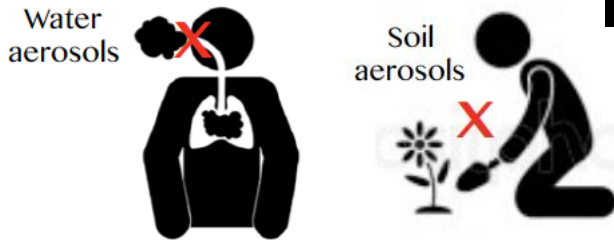
Mice



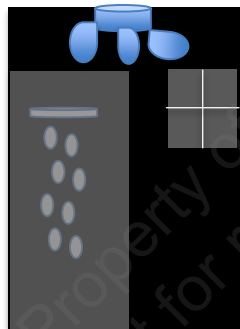
Amoebae

Proposed methods to reduce exposures

Avoid aerosol generating activities



Use fans or open windows in shower area



preferred

Replace or clean showerheads regularly
Avoid fine mist showerheads



**NOT
recommended**

Avoid bubbling water (*i.e.*, hot tubs, nail foot baths, humidifiers)



Avoid smoking



Flush water heater frequently
Increase hot water heater temperature



Avoid using tap water for nasal irrigations

Learning Objectives

- 1). Become familiar with examples of environmental niches for NTM.
- 2). Recognize features that may promote NTM in the environment – showcasing Hawai'i.
- 3). Identify possible methods to reduce environmental NTM exposures.

Steam disinfection of toothbrushes reduces NTM



M. abscessus subsp. *abscessus* (n=2)
M. abscessus subsp. *bolletii* (n=2)
M. abscessus subsp. *massiliense* (n=2)

None of the 80 isolates examined survived at 91°C for 4 min. ¹

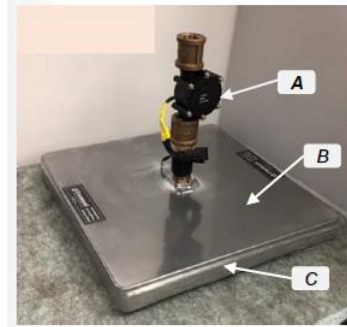
** Boiling water for 10-15min reduces NTM. ²

¹ Millar, et al., Pediatric Pulmonology, 2020

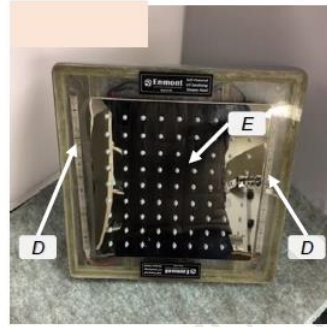
² Falkinham, WhiteJ, 2013

UV disinfection to reduce NTM in freshwater

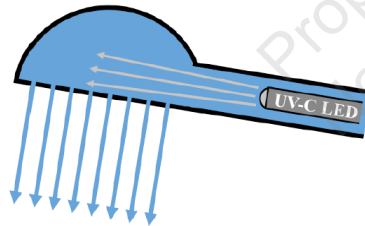
UV-LED adapted showerhead reduces *Legionella* (EPA)



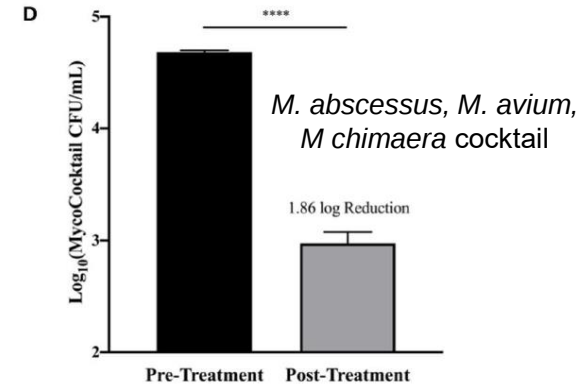
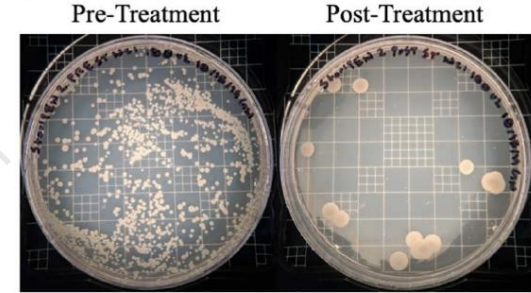
Top View



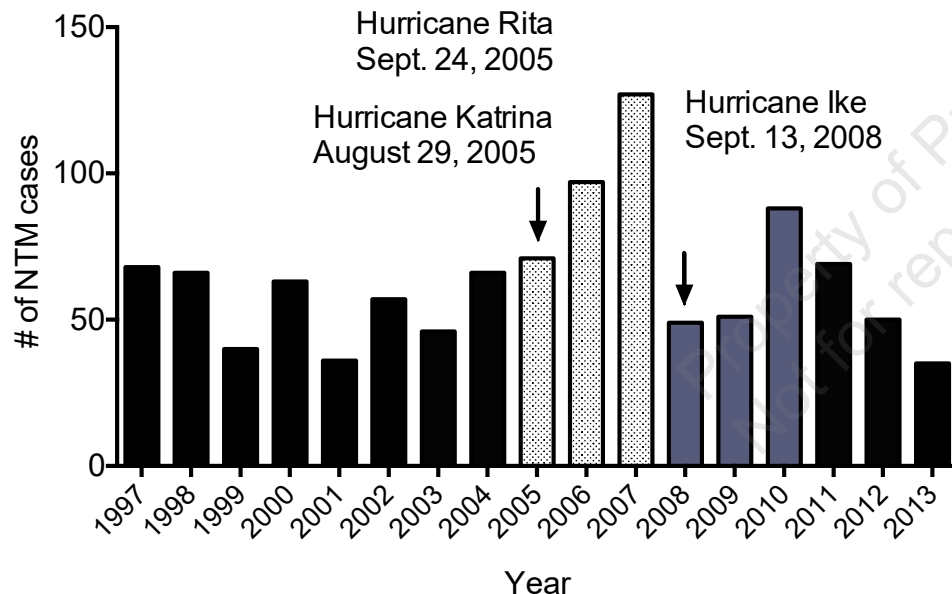
Bottom View



UV Water Bottles reduce NTM from drinking water



Reduce exposure to aerosol generating events



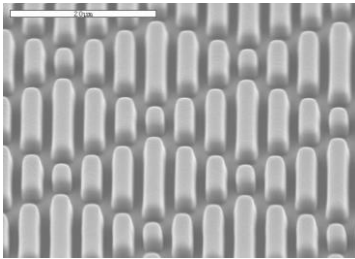
Honda, *et al*, Chest, 2015

Image: <https://www.usgs.gov/special-topic/fire>

Image: https://volcanoes.usgs.gov/volcanic_ash/ash_gas.html



Novel applications to reduce surface NTM adherence

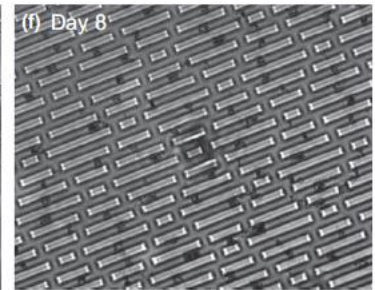
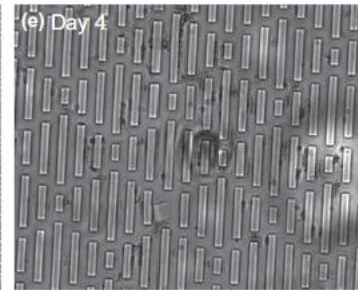
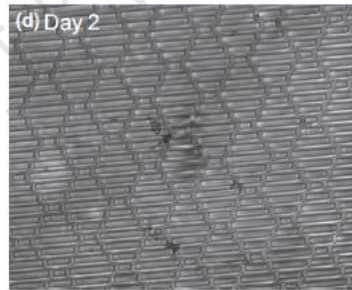
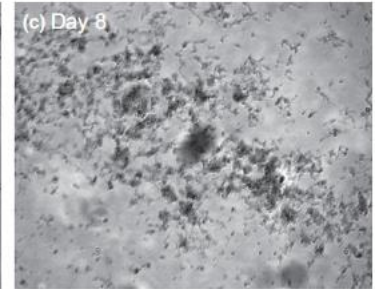
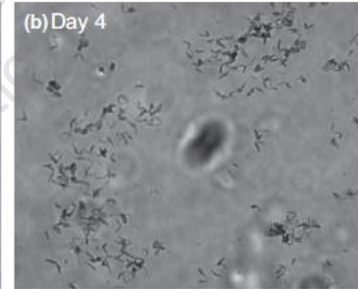
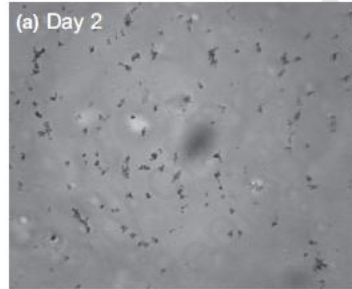


Sharklet(-)

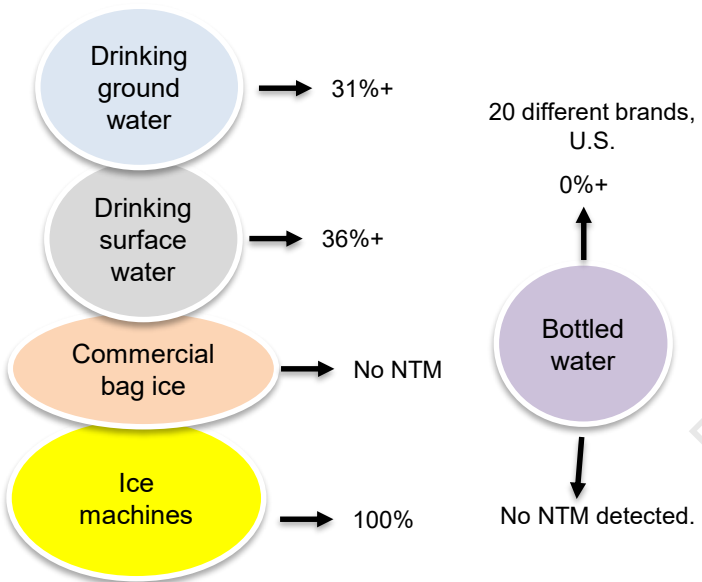


Sharklet(+)

M. abscessus



Expand Drinking Water Awareness



Bottled Water (Honda Lab, 2021):

Water Tested:	Type of water (Source = U.S.A.) unless noted:	Characteristic:	Results:
1	Bottled Water, Brand 1	Natural spring water	None
2	Bottled Water, Brand 2	Purified water	None
3	Bottled Water, Brand 3	Natural spring water	None
4	Bottled Water, Brand 4	Water from snow	None
5	Bottled Water, Brand 5	Volcanic rock filtered water	<i>Mycobacterium neoaurum</i> <i>Mycobacterium phocaicum</i>
6	Bottled Water, Brand 6	Volcanic rock filtered water	None
7	Bottled Water, Brand (non-U.S.A)	Volcanic rock filtered water	None
8	Distilled water	Commercially available	None
9	Sink faucet 1	Municipal water, Colorado	<i>Mycobacterium abscessus</i>
10	Sink faucet 2	Municipal water, Colorado	None
11	Sink faucet 3	Municipal water, Colorado	None
12	Water fountain	Municipal water, Colorado	None
13	Wall mounted water bottle filling station	Municipal water, Colorado	None

Holtzman, *et al*, J Food Protect 1997

Covert, *et al*, AEM, 1999

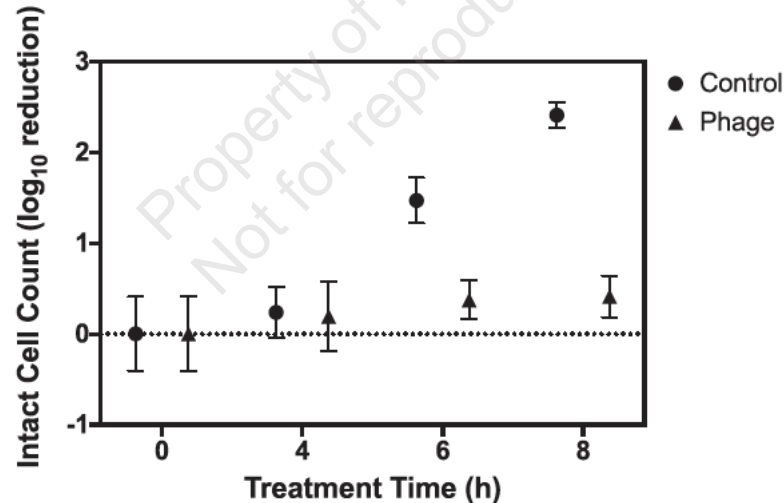
Totaro, *et al*, J Water Health, 2018

https://www.bottledwater.org/public/CCL4%20Microbes%20of%20Interest%20in%20Drinking%20Water_0.pdf0

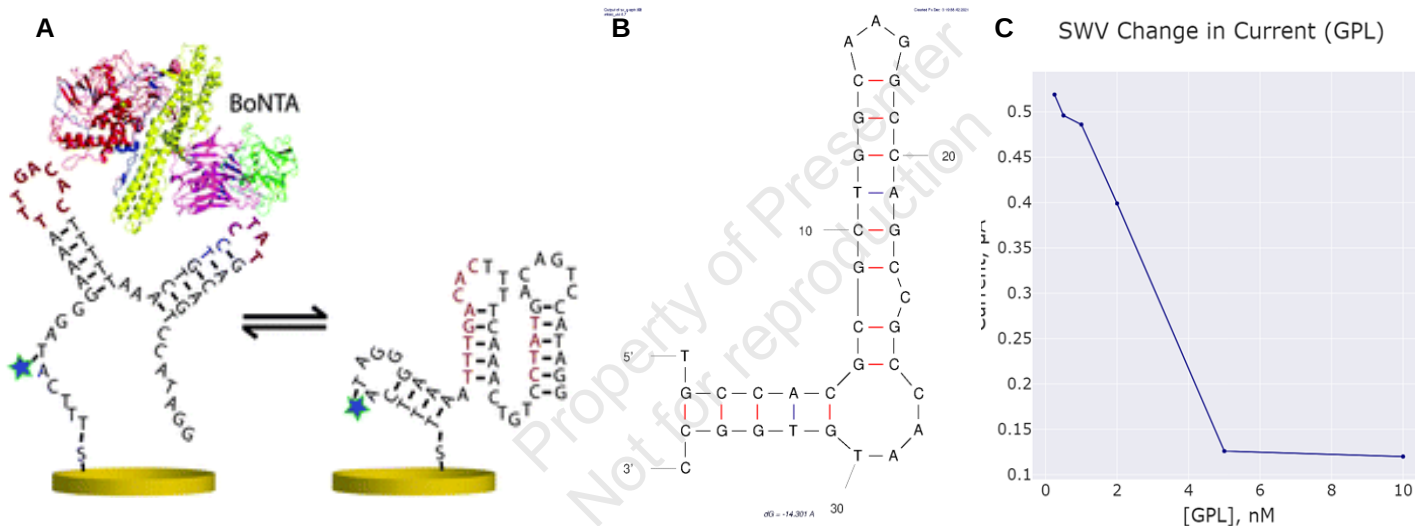
Honda Lab, unpublished.

Phages as possible sanitizers to prevent NTM in the built environment

Bacteriophage Treatment before Chemical Disinfection Can Enhance Removal of Plastic-Surface-Associated *Pseudomonas aeruginosa*



Developing new point of use DNA biosensors to detect environmental NTM (Andrew Bonham, PhD, Metro State University)



Electrochemical DNA-based biosensor for GPL. A) Schematic of the general biosensor approach, wherein a DNA aptamer selected to bind to a target, in this example Botulinum neurotoxin A (BoNTA), is attached to a gold electrode with a redox-active tag (blue star). Binding of the target changes DNA conformation and generates a specific, reproducible change in electrical current. **B)** SELEX-evolved DNA aptamer specific to GPL. **C)** Preliminary graph of electrical current response change when tested against purified GPL.

Acknowledgements

University of Texas Health Science Center at Tyler

Rachel Wilsey

National Jewish Health

Former Honda Lab

Stephanie Dawrs
Charmie Vang
Haley Hendrick
Ravleen Viridi
Grant Norton
Ryan Hoyt

Michael Strong PhD
Nabeeh Hasan, PhD
Elaine Epperson, PhD
Jo Hendrix

Former:

Sean Beagle
Cody Glickman, PhD

James Crooks PhD

Former:

Brady Holst
Melissa Lowe

Virginia Tech

Joe O. Falkinham, III PhD

NTM Hawai'i Consortium

'Iolani School, The Kamehameha Schools, Seabury Hall, Kailua High School, Waianae High School, Kapa'a High School, Island School, University of Hawai'i at Hilo, Kiana Frank, Carl Arume, University of Hawai'i Maui, Brigham Young University Laie, Mike Burnett, Homeschoolers, Department of Tropical Medicine University of Hawai'i, Hawai'i Citizen Scientists

North Carolina State University

Arielle Parsons, PhD (Former)
Krishna Pacifici, PhD

University of Hawai'i

Haley Hendrik (MS)
Sandra Chang, PhD

Pig Hunters Hawai'i

O'ahu Pig Hunters
Hawai'i Dept of Forestry and Wildlife
US. Dept of Agriculture
Nick Vargas
Keoki Oliveros

Brigham Young University, UT

Steve Nelson, PhD

Leeza Brown

Former:

Schuyler Robinson, MS

Hawai'i Volcano Observatory; USGS

David Damby, PhD
Tamar Elias

Diagnostic Lab Services, Hawai'i

Kaiser Permanente Hawai'i

Metro State University, CO

Andrew Bonham, PhD
Dylan Poch
Tyler Sodra

Funding: NSF Ecology, Evolution and Infectious Disease (#1743587), Padosi Foundation

