Equipment Run Charges

Equipment	Price	Units
ABI 7900HT Real-time PCR	\$20.00	per run
Agilent Bioanalyzer 2100	\$12.00	per run
Auto MACs	\$10.00	per hour
Genome Sequencer FLX 454	\$1,000.00	per run
Gentle MACs Dissociator	\$10.00	per hour
OpenArray NT Cycler System	\$16.00	per hour
SOLiD Covaris S2	\$25.00	per hour
Synergy HT Multi-Detection Microplate Reader	\$10.00	per hour

Sequencing

Agilent Bioanalyzer (labor included): The Agilent 2100 Bioanalyzer is a microfluidics-based platform for sizing, quantification and quality control of DNA, RNA, proteins and cells on a single platform. Results are delivered within 30-40 minutes. This is an essential QC check prior to microarray analysis.

RNA Nano Chip	\$58.00/chip
RNA Pico Chip	\$58.00/chip

Microarrays: Pricing is for Gene-Expression microarrays.	Other formats may be requested.
4X44K Feature Gene Expression Arrays	\$350.00/sample
Labor (typically, 5 hours/project)	\$60.00/hour

Capillary Sequencing (labor included): Sanger Sequencing.	
Instrument-Ready Samples	\$4.00 per sample
Reaction/Cleanup/Capillary Electrophoresis	\$6.00 per sample

Autopure LS: Automated DNA extraction from fresh or frozen blood, buffy coat, cell suspensions, saliva and cultured cells using PureGene Chemistry. Empty tubes charged as samples.

1-5 ml samples:	\$10.00 each
5-10 ml samples:	\$18.00 each

Real-Time PCR, OpenArray: High throughput, real-time PCR for gene expression, genotyping or digital PCR using SYBR Green and Taq-Man chemistry on pre-configured gene panels or completely customized content. Each OpenArray plate contains 3072 reactions of 33 nl each. Custom Materials, designed in

Custom Materials, designed in
conjunction w/ Life TechnologiesVariable Pricing (approx.
\$650/plate)Labor\$60.00/hour

Liquid Handling Automation: Tecan Freedom EVO or Qiagen BioRobot Universal available to automate liquid handling operations such as reaction setup, sample dilutions, cherry-picking, normalization, DNA and RNA extractions. Hourly charge applies to protocol development and operation.

Labor

\$60.00/hour

Illumina Veracode Genotyping: Illumina's Golden Gate chemistry is ideal for larger genotyping projects due to efficient multiplexing and content is completely customizable. Prices based on Illumina's list costs and subject to change.

Multiplex	# of Samples	# of Kits with same OPA	Price Per Sample	Approx. Labor/Kit	Price Per SNP	Price Per Kit
	480	1	\$10.50	\$840.00	\$0.28	\$6,384.00
48 plex	960	2	\$8.89		\$0.24	\$5,531.00
	≥1,440	>3	\$6.83		\$0.19	\$4,446.00
	480	1	\$12.48	\$840.00	\$0.16	\$7,429.00
96 plex	960	2	\$10.56		\$0.14	\$6,415.00
	≥1,440	>3	\$8.64		\$0.12	\$5,401.00
	480	1	\$17.28	\$840.00	\$0.14	\$9,963.00
144 plex	960	2	\$14.62		\$0.12	\$8,560.00
	≥1,440	>3	\$12.10		\$0.10	\$7,228.00
	480	1	\$23.04	\$840.00	\$0.14	\$13,005.00
192 plex	960	2	\$19.50		\$0.12	\$11,134.00
_	≥1,440	>3	\$16.13		\$0.10	\$9,356.00
	480	1	\$38.40	\$840.00	\$0.11	\$21,115.00
384 plex	960	2	\$30.72		\$0.09	\$17,060.00
	≥1,440	>3	\$19.20		\$0.06	\$10,977.00

Next Generation Sequencing

Price includes reagents, consumables and a minimum of an hour of labor for library construction, emulsion PCR and sequence run. Prices may vary according to unforeseen sample issues and subsequent troubleshooting.

Please contact Eve Farias-Hesson, PhD @ (303) 270-2650 or <u>farias-hessone@njhealth.org</u> for an estimate for a project with a large number of samples or to obtain a quote for a grant proposal.

No of Regions per PTP	Reads per region (x 10 ³)	Throughput per region	Total Reads per PTP	Throughput per PTP
2	450 -650	180 - 280 Mb	~1,000,000	360 - 560 Mb
4	160 - 250	60 - 110 Mb	~800,000	240 - 440 Mb
8	80 - 120	30 - 55 Mb	~800,000	240 - 440 Mb
16	25 - 40	10 - 20 Mb	~512,000	160 - 320 Mb
454 GS FLX Tita	nium Sequence run			
Service ^{1,2}		Price		
454 FLX 2 region	PTP sequence	\$8,000		
454 FLX 4 region	PTP sequence	\$9,000		
454 FLX 8 region	PTP sequence	\$11,000		
454 FLX 1/2 regi	on PTP sequence	\$4,000		
454 FLX 1/4 regio	on PTP sequence	\$2,250		
454 FLX 1/8 regio	on PTP sequence	\$1,375		
emPCR ³				
SV emPCR titratio	on (4x)	\$600		
Library Construc	ction ⁴			
Short fragment rer	noval (per sample)	\$25		
Library Quality assessment (per chip)		\$60		
Shotgun library (g	DNA) construction	\$550		
Library Construc	tion Training			
Per hour		\$100		
Per nour		\$100		

454 GS FLX Titanium Sequencing

454 FLX Titanium Expected Run Results PTP 70 x 75

1- Additional information on pricing is available upon request for the 16 region PTP.

2- Partial PTP regions are available, however, the sequence run will be started upon obtaining the complete number of samples for running a full PTP plate.

3- An additional small volume emulsion PCR titration fee may apply if a sample requires troubleshooting.

4- Please contact us for an estimate of amplicon and paired-end library construction.

SOLID 5500xl

Library construction for DNA, RNA and ChIP samples are available. Please contact us for a cost estimate.

Service	Description	Pricing unit	Expected reads	Price
Library quality assessment	post-library QC	per chip	N/A	\$45
Library Construction Training	DNA, mRNA, ChiP	hour	N/A	\$100
SOLiD 5500xl sequencing	single read 75bp	per lane	170 Million	\$1,270
SOLiD 5500xl sequencing	mate-pair 65 x 65bp	per lane	340 Million	\$1,570
SOLiD 5500xl sequencing	paired end 75 x 35bp	per lane	340 Million	\$2,184

1- Price per lane is subject to change if different users request individual ePCR per lane. Please contact us to get a project cost estimate.

2- Lane run is available, however, sequence run will be started upon obtaining the complete number of samples for running a full flowchip.

Ion Torrent (PGM)

Expected sequence run results

Chip	Throughput	
314 chip	10Mb	
316 chip	100Mb	
318 chip	1Gb	
S	ervice	Price
Library C	onstruction	
Fragment l	ibrary	\$240
100bp rea	d length	
314 chip		\$562
316 chip		\$762
318 chip		\$962
200bp rea	d length	
314 chip		\$912
316 chip		\$1,112
318 chip		\$1,312
Library C	onstruction Traini	ing
Per hour		\$100.00

Bioinformatics Analysis

Analysis of data generated by the Center for Genes, Environment, and Health varies from basic analysis, which produces the output files typical for a given application, to complex characterization of the genes or loci implicated by the experimental study. Fees shown below apply for basic analyses, using typical pipelines, with the output level and form as detailed. The list of options may vary as additional pipelines are developed. For analysis needs that involve the creation of custom analysis scripts or implementation of programs to investigate specific questions of the data, support will be available at the bioinformatics analysis hourly rate; please contact Sonia Leach, PhD, the Director of Bioinformatics @ (303) 270-2568 or leachs@njhealth.org to discuss the details of your project.

These projects will be charged at the bioinformatics analysis rate of \$65 per hour.

Basic Analysis Support

Analysis Option	Description	Estimated Set-	Relevant Output Files	Fee
		Up and		
		Computation		
		Time		
Mapping to	Platform-specific mapper	~1-24 hours	Mapping and quality	\$50 per
sequenced	or <u>Bowtie</u>		control statistics, *.bam	library
genome†			file of aligned reads	
SNP Detection	Platform-specific or	~2-8 hours for	*.gff or *.vcf file	\$100
and Annotation	samtools identification of	SNP detection.	describing SNPs. Tab	
	SNPs. Determination of	Annotation time	delimited text file	
	functional consequence	depends on	describing effect of	
	(missense, non-sense,	number of	variant (and whether in	
	non-coding, etc.), Human	SNPs and	dbSNP for Human)	
	using <u>SeattleSeq</u> ,	program used.		
	otherwise Annovar for			
	genomes with known gene			
	models			
Small InDel	Detection of insertions	~2 hours for	*.gff or *.vcf file. Tab	\$100
Detection and	(~<20bp) or deletions	detection, ~1	delimited text files	
Annotation	(<10kb). Determination of	hour for	describing effect of	
	functional consequence	annotation.	variant (and whether in	
	(missense, non-sense,		dbSNP for Human)	
	non-coding, etc.), Human			
	using SeattleSeq,			
	otherwise Annovar for			
	genomes with known gene			
	models			

DNA-seq (Whole Genome, Exon, or Targeted)

Analysis Option	Description	Estimated Set-	Relevant Output Files	Fee
		Up and		
		Computation		
		Time		
CNV Detection	Determination of copy	~2 hours	List of genes within CNV	\$100
and Annotation	number variation, with		segment, and *.gff file	
	filtering against known		describing CNV	
	segmental duplications			
Large Indel	Determining large	~1 hour	List of genes within indel,	\$100
Detection and	insertions and deletions		and *.gff file describing	
Annotation			indel	

DNA-seq (Whole Genome, Exon, or Targeted) Cont.

[†]Mapping to specialized sequence reference file will require special consideration

RNA-seq				
Analysis Option	Description	Estimated Set-	Relevant Output Files	Fee
		Up and		
		Computation		
		Time		
Mapping to	Platform-specific mapper	~1-24 hours	Mapping and quality	\$50 per
sequenced			control statistics, *.bam	library
genome†			file of aligned reads	
Mapping using	Alternative 3rd party	~6-24 hours	Mapping and quality	\$50 per
<u>TopHat</u>	mapping algorithm, with		control statistics, *.bam	library
	or without reference gene		file of aligned reads	
	models, identifies splice			
	junctions. Especially for			
	Illumina reads			
Differential	Generates counts of	~3 hours	List of Ensembl	\$75
Expression using	mapped data for reference		transcripts with fold	
<u>DESeq</u>	gene model (e.g.		changes and p-values,	
	Ensembl), differential		*.bedgraph file	
	expression test by			
	negative binomial			
Differential	Assembles transcripts,	~10 hours	List of assembled	\$100
Expression and	estimates abundances,		transcripts (Ensembl id if	
Transcript	tests for differential		known) with FPKM, fold	
Assignment using	expression by t-test		changes, p-values, and	
Cufflinks/Cuffdiff			Microsoft Excel macro-	
			enabled file to explore	
			results in IGV, *.bedgraph	
			file	

ChIP-seq

Analysis Option	Description	Estimated Set-	Relevant Output Files	Fee
		Up and		
		Computation		
		Time		
Mapping to	Platform-specific mapper	~2-10 hours	Mapping and quality	\$50 per
sequenced	or <u>Bowtie</u>		control statistics, *.bed	library
genome†			file	
Peak Calling	Determination of peaks	~ 1 hour	*.bed file with list of	\$75
using <u>MACS</u>	with or without a control		peaks	
	sample			
Peak Calling	Determination of peaks	~1 hour	*.bed file with list of	\$75
using <u>ChIPDiff</u>	and differential		peaks	
	enrichment between two			
	samples			

Functional Characterization

Analysis Option	Description	Estimated Set- Up and Computation Time	Relevant Output Files	Fee
<u>Ingenuity Systems</u> <u>IPA</u> Pathway Analysis (Self run)	Given list of genes, determine over- represented pathways	~2 hours	List of over-represented pathways with p-values and genes annotated to them	\$20 per hour
<u>Ingenuity Systems</u> <u>IPA</u> Pathway Analysis (Center staff run)	Given list of genes, determine over- represented pathways	~2 hours	List of over-represented pathways with p-values and genes annotated to them	\$50 per hour
<u>DAVID</u> Functional Annotation	Given list of genes, perform batch annotation, gene-term enrichment (Gene Ontology, Protein interactions, functional domains, disease associations, pathways, sequence features), and determine over- representation	~1 hour	Gene annotation and list of over-represented pathways with p-values	\$50 per hour

Microarray Analysis

Analysis Option	Description	Estimated Set- Up and Computation Time	Relevant Output Files	Fee
Partek Genomics Suite (Self run)	Determines list of significantly differential genes	~2 hours	List of genes and p- values	\$20 per hour
Partek Genomics Suite (Center staff run)	Determines list of significantly differential genes	~2 hours	List of genes and p- values	\$50 per hour

Other Bioinformatics Analysis

Hourly Bioinformatics Analysis	\$65 per hour
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<u>FlexiVent</u>

Run by CGEH Staff	\$50.00/hour
Self Run *	\$35.00/hour
Data Analysis and Interpretation	\$50.00/hour
FlexiVent Training	\$30.00/hour

*Must be trained on using FlexiVent and receive approval from Center staff. There is a 8 animal minimum for FlexiVent. Contact Leah Teeter to schedule FlexiVent @ (303) 270-2650 or teeterl@njhealth.org

Outside institutions will be charged 58.5%, which is the current NIH indirect rate. A 10% administrative fee will be added to all supplies purchased through the Center.

Please contact the Genetics Core for pricing as charges vary per application, and number of submitted samples.

Before submitting your DNA/RNA sample, please confirm that:

DNA Sample Requirements-

- DNA must be double stranded. If possible, please submit a well-labeled gel image to indicate that the sample is not degraded and is within the concentration range determined by spectrophotometry.
- DNA should not be the result of whole genome amplification (or other similar process which may compromise representativity).
- DNA should not be degraded

- DNA should contain no particulate matter, organic solvents, salts or proteins.
- DNA should have an OD 260/280 ratio ≥ 1.8 .
- Your sample has the minimum amount of DNA for sample preparation. A minimum of 10ug of DNA is recommended for preparing mate-pair libraries of microbial genomes, 20ug for preparing mate-pair libraries of human genome, and 1ug for fragment or paired-end libraries.
- Your sample is in TE buffer (e.g., Qiagen EB) with a minimal concentration of 50 ng/uL (DNA).

RNA Sample Requirements-

- RNA should not be degraded
- RNA should have an RNA integrity number (RIN) greater than 9.
- Whole transcriptome library construction input:
 - 100-500ng of poly(A) RNA or 250-650ng of rRNA-depleted total RNA
 - Suspended in 10uL Nuclease-free water
 - Absent of contaminating rRNA

Library ready for ePCR Sample Requirements-

- Your sample is in TE buffer (e.g., Qiagen EB) with a minimal concentration of 5ng/ul.
- Provide a brief workflow of the library sample preparation.
- Inform origin of adaptors used for making libraries (e.g. IDT HPLC purified oligos).
- If barcoding libraries, inform sequence and barcode series (DNA/RNA) used for preparing each library submitted for sequencing. If sequencing using our SOLiD 5500xl instrument, in order to obtain color balance during sequencing we recommend design experiments to use at least one of the full sets of four barcodes (e.g. barcodes 1-4, 5-8, 9-12, 13-16, 17-20, etc).
- Provide, if available, Bioanalyzer file containing results for the final libraries.